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# Platelets

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Gökhan Cüce and Tahsin Murad Aktan

Additional information is available at the end of the chapter

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## 1. Introduction

General information about platelets, origin of platelets and granule contents of platelets were summarized.

## 2. Platelets

These cell fragments are morphologically small scale but functionally vital under life threatening conditions (1). They originate from megakaryocytes located mainly in the bone marrow, found in circulating blood and stored in spleen (2). Platelets don't contain a nucleus and during their inactive state they have a discoid morphology with a diameter of 2-4 micrometer (3, 4). But whenever they are active they can change their morphology very rapidly to an irregular branched spread form (5). Currently platelets are being used at wide spread clinic treatments from cosmetic needs to supporting insufficient heart (6, 7).

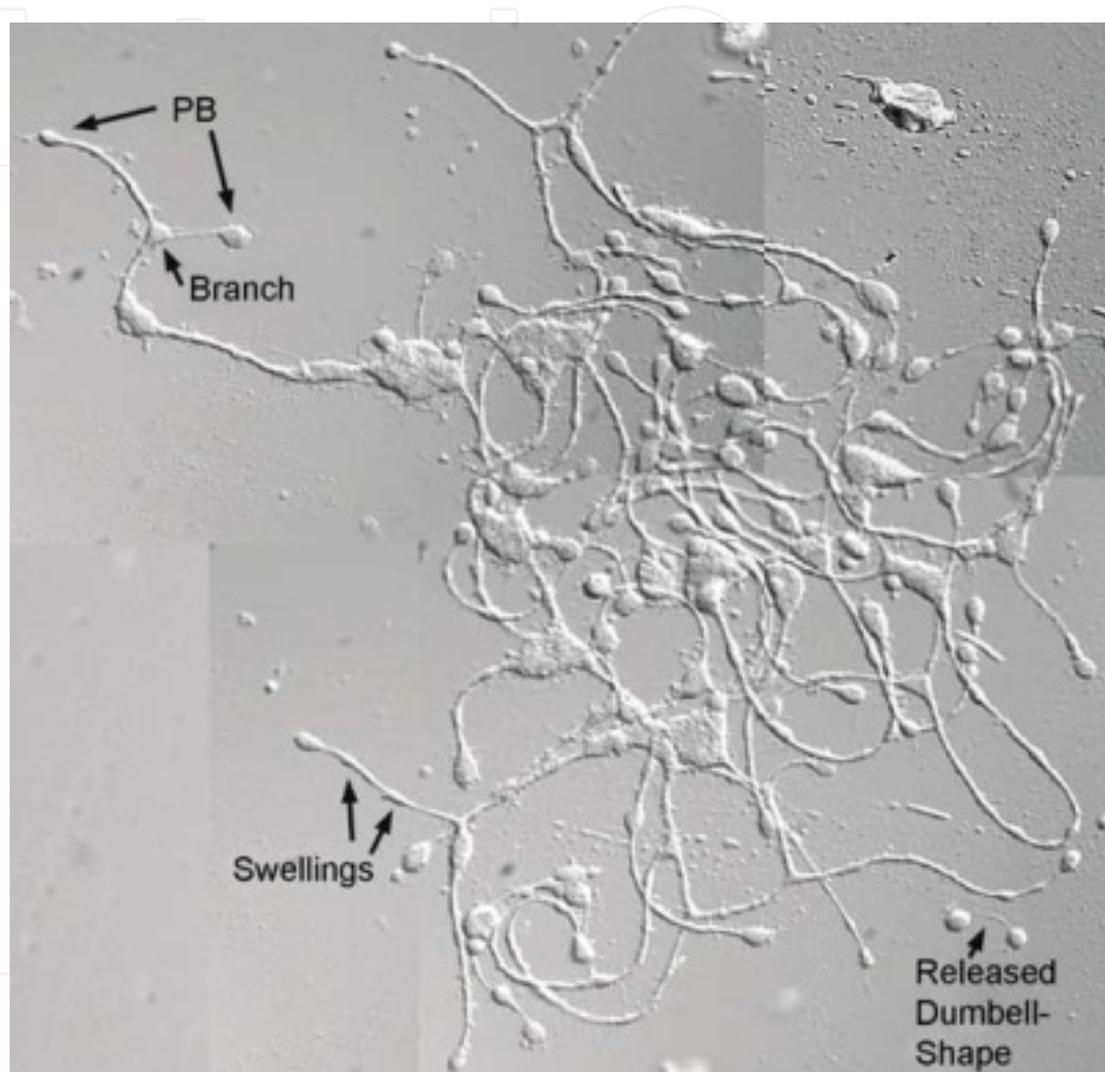
### 2.1. Development of Platelets

It is not exactly explained how platelets originate from megakaryocytes. There are several models to explain formation of platelets.

Megakaryocytes seem to locate as triple form. With their VEGF secretion capacity they hold vessel endothelial cells close to themselves (8). The most scientifically accepted three models are mentioned as,

1. Simply blebbing from the cell membrane of megakaryocytes (1).
2. In megakaryocytes there are special cell fields defined as "Demarcation Membrane System" where granules of platelets condense and fragments break away (9).
3. The most popular theory seems to be "Proplatelet Formation". Here megakaryocytes have long thin branch like extensions at the blood circulating site of blood vessels of

bone marrow and on these branches there are uprising small bodies where by the help of blood shear force platelets enter directly to circulating blood stream. It was suggested that the concept of platelet like bodies arise from pseudopods of Megakaryocytes, the forming platelets were named as “proplatelet” (10).



**Figure 1.** Megakaryocyte branches with Platelet Buds (PB) are seen. Proplatelets are released as Dumbbell shaped bodies. This image is referenced from Hartwig and Italiano 2003 (Thanks for the kind permission of John Wiley and Sons to use this image) (11) .

Kinetics of platelets; they have a life span as 7-10 days and in 1 liter human blood it is estimated that there are  $150-400 \times 10^9$  platelets so for a balanced number they are formed  $15 \times 10^9 - 40 \times 10^9$  daily. Megakaryocytes located in the bone marrow sinusoids form a barrier to other bone marrow cells, it forms a physical barrier preventing direct contact to blood circulation. But there are canalicular openings in megakaryocyte membrane which permits cell migration to other cells to enter blood stream; this is named as “Emperipolesis” (8).

These small cell fragments have complex properties; 2 cytoplasmic regions can be seen in platelets

1. **Hyalomere:** The light blue homogeneous region of the peripheral cytoplasm is called Hyalomere. Hyalomere includes cytoplasmic filaments and circumferential microtubule bundle under the cell membrane. These elements of the cytoskeleton provide the movement and the protection of the platelets' shapes.
2. **Granulomere (Chromomere):** This is the central region and tight area. It is ranging in color from blue to purple-staining. Granulomere includes small Golgi complex, smooth endoplasmic reticulum, lysosome, scattered granules surrounded by a membrane and a variety of mitochondria (4).

Platelets have a simple appearance but carry very complex functional properties. By dividing this simple cell fragment to four regions helps for a better understanding of the functions of platelets.

1. **Peripheral Zone:**  
This region is composed from unit membrane with open canalicular system. Three parts are defined as;
  - a. **Exterior outer layer:**  
This is a glycocalix membrane with 10-20 nm thickness and thicker than the other blood cells, rich from glycoproteins that are mainly receptors for cell-cell and cell-vessel interactions(1, 8).
  - b. **Platelet Unit Membrane:**  
Platelet unit membrane has some similarities and appearance with other unit membranes of cells, it is composed from bilipid layer rich of phospholipids (12), it can distribute molecules according to phsico-chemical properties for passing the membrane. The membrane has anionic and cationic pumps. Platelet unit membrane is an important catalyst for liquid phase coagulation.
  - c. **Submembrane Zone:**  
Just located under the unit membrane a layer composed of microflament network. This network is anatomically and functionally related to membrane glycoproteins and cytoplasmic filament system.
2. **Sol-Jel Zone:**  
This is cytoplasm corresponding part of the cellular fragment, platelet. It is in soluble or gel phase according to changes of polymerization of the filaments; actin and microtubules(1).  
Just under the submembrane zone there are microtubules forming a peripheral ring which helps platelet to maintain its discoid shape in inactive form. When activated, the microtubules surround the organelles and with the contribution of other filaments (13), the organelles are tightly contracted. During silent form only 30-40 % of actin filaments are polymerized, when platelets are activated the polymerized amount increases(1).

3. Organel Zone:

This is the zone where granule's, peroxisome's, lysosome's and mitochondria's are localized. There are enzymes, adenine nucleotids, calcium, serotonin and many other proteins in this region (1).

4. Membrane Zone

There is a distinguishing feature of platelets that their plasma membrane contains wide spread invaginations that forms a network inside platelet. Finally with pore openings the inner network is directly in contact with outer zone. This system is named as "open canallicular system" (OCS) and with this system an extensive amount of surface area stays as potential in silent state. With this system also platelet gains a large area for molecular trafficking. A second canal system is composed from endoplasmic reticulum networks and named as "Dense Tubular System" (DTS). Here in DTS many enzymes and calcium ions that are important for activation are located. DTS is not directly connected to outer membrane (1, 14) but has close connections with OCS. These two systems actively exchange molecules (1).

The granules have diameters ranging between 200 to 500 nm and they are found as spherical or oval structures (15). There are 3 types of granules in platelets, Alfa Granules, Dense granules, lysosomes. Alpha granules are most prominent in terms of material content and majority. These granules include inflammatory molecules, cytokines, cell-activating molecules, proteins, Growth Factors, adhesion molecules, integrins and other proteins These granules are filled by megakaryocytes (3).

### 3. Alpha granules

It is widely accepted that these granules come from the budding of trans golgi apparatus organel of megakaryocytes (16, 17).

These are 200-400 nm diameter granules widespread in the cytoplasm (16) which gives the granular appearance in Romanoski stained smear preparations, each platelet contains around 50-80 of these granules. The content of granules is very diverse; a brief list is given in table 1 (14, 18, 19, 20, 21).

When platelets are activated these alpha granules fuse with each other, OCS and plasma membrane. The secretion of alpha granules is mediated by some proteins (such as SNARE) and membrane lipids (19).

The secretions effect platelet and cells in the environment (such as endothelial, leukocytes) for migration, adhesion and proliferation(14).

A rare syndrome named as Gray Thrombocyte Syndrome (GTS) is both involved with the quantity and quality of platelets which cases susceptibility for bleeding. In GTS the proteins synthesized by megakaryocytes are abnormal and don't enter platelets as they do in normal individuals and additionally the endocytotic mechanisms don't work properly. As a result the secretions spread to bone marrow and a fibrosis forms (miyelifibrozis)(22, 23).

Thrombospondin P-selectin platelet factor 4 beta thromboglobulin Factors V, XI, XIII fibrinogen von Willebrand factor fibronectin vitronectin high molecular weight complexes kininogen chemokines mitogenic growth factors (platelet-derived growth factor) vascular endothelial growth factor TGF-beta
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**Table 1.** Some main components of alpha granules.

#### 4. Dense granules

These are smaller granules with 150 nm diameter (24), because of the calcium and phosphate content their image seems dense under electron microscopic (EM) observation (21, 25). Each platelet contains 3-8 of these granules (14). The components of dense granules are briefly given in Table 2 (10, 14, 19, 20).

Ca Mg P pyrophosphate Nucleotides ATP, GTP, ADP, GDP Membrane proteins CD63 (granulophysin) LAMP 2 Serotonin GPIIb, GPIIb/IIIa P-Selectin Histamine Epinephrine
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**Table 2.** Some main components of dense granules.

In activated platelet these granules fuse with plasma membrane and expel their ingredients to their environment which causes other platelets to aggregate and a local vasoconstriction (especially by serotonin) in the involved vessels. Also the ADP content is a very important participant for homeostasis (14).

The importance of the components of dense granules for homeostasis is recognized when the diseases of the deficiency of dense granules was defined as Hermansky-Pudlak

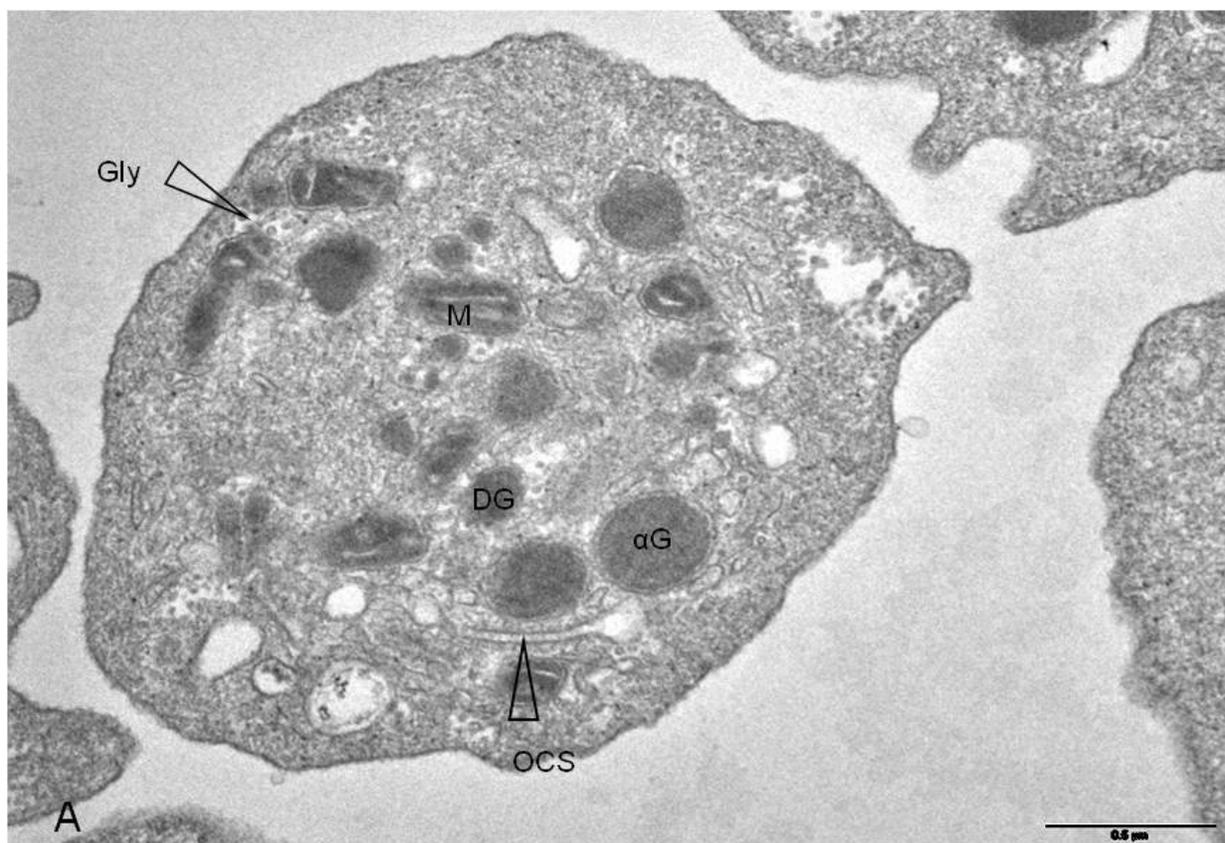
Syndrome (26, 27, 28) and Chediak Higashi Syndrome. In both syndromes stoppage of bleeding is defective based on the impairment in dense granules (14).

## 5. Lysosomes

They have a diameter of 200-250 nm which places them to middle size granule (14). They can't be distinguished from alpha granules under EM observation because of the similarities in dense electron appearance. By the content of acid phosphates and arylsulphates cytochemical staining techniques can effectively distinguish lysosomes from alpha granules. In an activated platelet they expel their contents to environment as the other two granules by membrane fusing mechanisms. The difference for lysosomes to be involved in activation is that they need a more potent stimulus. The role of lysosomal components in homeostasis is not well understood as the other granules contribution. They are involved in thrombus formation and extracellular matrix remodeling (8).

It seems that lysosomes in platelets don't have any distinguished features, they share the common features with other cells lysosomes (29).

The components of dense granules are briefly given in Table 3 (8, 18, 30, 31, 32).



**Figure 2.** M: Mitochondria, αG: alfa-granules, DG: dense granules, Gly: glycogen particles and OCS: open canalicular system. The morphology can be seen in equatorial section of a human platelet. This image is referenced from Zufferey 2011 (Thanks for the kind permission of John Wiley and Sons to use this image)(33).

PF3
Acid phosphatase
Glucose-6 phosphatase
Arabinosidase
N-Acetyl-galactosaminidase
ATP = adenosine triphosphate
TGF
CD63
Cathepsin
lysosomal membrane proteins (LAMP-1, LAMP-2)
acid hydrolases
cathepsins

**Table 3.** Some main components of platelet lysosomes

## 6. Autologous platelet rich plasma (PRP)

The application of growth factors in medical practice is one of the areas where basic clinical research has focused its attention but there are many problems associated with their local administration. For example, recombinant human growth factors are not cost effective, they have limited shelf life, and ineffectively delivered to target cells and in addition, to get efficient therapy, large doses are needed. The use of autologous platelets concentrates for tissue regeneration and wound healing has now become an alternative easy and cheap way to obtain high concentrations of these growth factors (34).

The autologous blood collected from a patient just before surgery can be prepared as platelet concentrates, platelet-rich plasma (PRP) and platelet gel for the treatment the patient specifically needs (35). These forms are prepared by gradient density centrifugation techniques to obtain high (x5) concentration of platelets (36). This autologous concentration includes a large amount of growth factors, especially PRP is an easy and inexpensive technique to accelerate the wound healing (37).

This quite new field is open for research, there are a lot of techniques still under development stage such as platelet gels can be obtained by adding thrombin to autologous platelet-rich plasma. The initiation of fibrin polymerization and the release of platelets factors and cytokines can be achieved by the specific activators such as thrombin, glass, freeze-thaw cycle to platelet-rich plasma depending on what is required during the surgery (35).

In spite of the distinct features of platelet-rich plasma (PRP) and its use by different fields of medicine, no adverse reactions were documented until now(38, 39, 40, 41).

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## 7. References

- [1] Becker RC. Platelet Biology: The Role of Platelets in Hemostasis, Thrombosis and Inflammation. Platelets in Cardiovascular Disease. In: Bhatt DL. Imperial College Press. London, 2008:1-3.
- [2] Mason KD, Carpinelli MR, Fletcher JI, Collinge JE, Hilton AA, Ellis S, Kelly PN, Ekert PG, Metcalf D, Roberts AW, Huang DC, Kile BT. Programmed anuclear cell death delimits platelet life span. *Cell*. 2007;128(6):1173-86.
- [3] Rozman P, Bolta Z. Use of platelet growth factors in treating wounds and soft-tissue injuries. *Acta Dermatovenerol Alp Panonica Adriat*. 2007;16(4):156-65.
- [4] Ovalle WK, Nahirney PC. *Netter Essential Histology*. Saunders; 2007;166.
- [5] Klages B, Brandt U, Simon MI, Schultz G, Offermanns S. Activation of G12/G13 results in shape change and Rho/Rho-kinase-mediated myosin light chain phosphorylation in mouse platelets. *J Cell Biol*. 1999;144(4):745-54.
- [6] Anitua E, Sánchez M, Nurden AT, Nurden P, Orive G, Andía I. New insights into and novel applications for platelet-rich fibrin therapies. *Trends Biotechnol*. 2006(5):227-34.
- [7] Mishra A, Velotta J, Brinton TJ, Wang X, Chang S, Palmer O, Sheikh A, Chung J, Yang PC, Robbins R, Fischbein M. RevaTen platelet rich plasma improves cardiac function after myocardial injury. *Cardiovasc Revasc Med*. 2011;12(3):158-63.
- [8] Drouin A, Cramer EM. Production of Platelets. Editor: Gresele P, Page CP, Fuster V, Vermynen J. *Platelets in Thrombotic and Non-Thrombotic Disorders: Pathophysiology, Pharmacology and Therapeutics*. Cambridge University Press; 2002;25.USA.
- [9] Schulze H, Korpál M, Hurov J, Kim SW, Zhang J, Cantley LC, Graf T, Shivdasani RA. Characterization of the megakaryocyte demarcation membrane system and its role in thrombopoiesis. *Blood*. 2006;107(10):3868-75.
- [10] Italiano JE Jr, Shivdasani RA. Megakaryocytes and beyond: the birth of platelets. *J Thromb Haemost*. 2003;1(6):1174-82.
- [11] Hartwig J, Italiano J Jr. The birth of the platelet. *J Thromb Haemost*. 2003;1(7):1580-6.
- [12] White JG. Platelet Structure. Editor: Michelson AD. *Platelets*. Elsevier: USA, Second Edition, 2007;45
- [13] White JG. Views of the platelet cytoskeleton at rest and at work. *Ann N Y Acad Sci*. 1987;509:156-76.
- [14] Rumbaut RE, Thiagarajan P. Platelet-Vessel Wall Interactions in Hemostasis and Thrombosis. Editor: Granger DN, Granger JP. *Colloquium Series on Integrated Systems*

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- Physiology: From Molecule to Function to Disease. Morgan & Claypool Life Sciences; 2009-2011:5.
- [15] Gassling VL, Açıl Y, Springer IN, Hubert N, Wiltfang J. Platelet-rich plasma and platelet-rich fibrin in human cell culture. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod.* 2009 ;108(1):48-55.
- [16] King SM, Reed GL. Development of platelet secretory granules. *Semin Cell Dev Biol.* 2002(4):293-302.
- [17] Blair P, Flaumenhaft R. Platelet alpha-granules: basic biology and clinical correlates. *Blood Rev.* 2009(4):177-89.
- [18] McNicol A, Israels SJ. Platelet dense granules: structure, function and implications for haemostasis. *Thromb Res.* 1999;95(1):1-18.
- [19] Reed GL. Platelet secretory mechanisms. *Semin Thromb Hemost.* 2004;30(4):441-50.
- [20] Askari AT, Messerli AW, Lincoff M. *Thrombosis and Antithrombotics in Vascular Disease. Management Strategies in Antithrombotic Therapy.* Editör: Askari AT, Messerli AW. Wiley; USA. 2008:3.
- [21] Ma AD, Key NS. *Molecüler Basis of Hemostatic and thrombotic Diseases.* Editör: Coleman WB, Tsongalis GJ, London. *Molecular Pathology: The Molecular Basis of Human Disease.* Academic Press; 1 edition, 2009:258.
- [22] Di Paola J, Johnson J. Thrombocytopenias due to gray platelet syndrome or THC2 mutations. *Semin Thromb Hemost.* 2011(6):690-7.
- [23] Nurden AT, Nurden P. The gray platelet syndrome: clinical spectrum of the disease. *Blood Rev.* 2007(1):21-36.
- [24] Rendu F, Brohard-Bohn B. The platelet release reaction: granules' constituents, secretion secretion and functions. *Platelets.* 2001;12(5):261-73.
- [25] Ruiz FA, Lea CR, Oldfield E, Docampo R. Human platelet dense granules contain polyphosphate and are similar to acidocalcisomes of bacteria and unicellular eukaryotes. *J Biol Chem.* 2004;279(43):44250-7.
- [26] King SM, McNamee RA, Houng AK, Patel R, Brands M, Reed GL. Platelet dense-granule secretion plays a critical role in thrombosis and subsequent vascular remodeling in atherosclerotic mice. *Circulation.* 2009;120(9):785-91.
- [27] Nisal M, Pavord S, Oppenheimer CA, Francis S, Khare M. Hermansky-Pudlak syndrome: management of a rare bleeding disorder in a twin pregnancy. *J Obstet Gynaecol.* 2012 ;32(2):185-6.
- [28] Saftig P, Klumperman J. Lysosome biogenesis and lysosomal membrane proteins: trafficking meets function. *Nat Rev Mol Cell Biol.* 2009;10(9):623-35.
- [29] Skoglund C. *Platelets in inflammation.* Linköping University Medical Dissertations. 2010-Sweden;14.
- [30] Gerrard JM, Phillips DR, Rao GH, Plow EF, Walz DA, Ross R, Harker LA, White JG. Biochemical studies of two patients with the gray platelet syndrome. Selective deficiency of platelet alpha granules. *J Clin Invest.* 1980;66(1):102-9.
- [31] Nishibori M, Cham B, McNicol A, Shalev A, Jain N, Gerrard JM. The protein CD63 is in platelet dense granules, is deficient in a patient with Hermansky-Pudlak syndrome, and appears identical to granulophysin. *J Clin Invest.* 1993;91(4):1775-82.

- [32] Grau AJ, Reiners S, Lichy C, Buggle F, Ruf A. Platelet function under aspirin, clopidogrel, and both after ischemic stroke: a case-crossover study. *Stroke*. 2003;34(4):849-54.
- [33] Zufferey A, Fontana P, Reny JL, Nolli S, Sanchez JC. Platelet proteomics. *Mass Spectrometry Reviews*, 2011; 31, 331–351.
- [34] Nikolidakis D, Jansen JA. The biology of platelet-rich plasma and its application in oral surgery: literature review. *Tissue Eng Part B Rev*. 2008 Sep;14(3):249-58.
- [35] Soffer E, Ouhayoun JP, Anagnostou F. Fibrin sealants and platelet preparations in bone and periodontal healing. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod*. 2003;95(5):521-8.
- [36] Huang Q, Wang YD, Wu T, Jiang S, Hu YL, Pei GX. Preliminary separation of the growth factors in platelet-rich plasma: effects on the proliferation of human marrow-derived mesenchymal stem cells. *Chin Med J (Engl)*. 2009;122(1):83-7.
- [37] Napolitano M, Matera S, Bossio M, Crescibene A, Costabile E, Almolla J, Almolla H, Togo F, Giannuzzi C, Guido G. Autologous platelet gel for tissue regeneration in degenerative disorders of the knee. *Blood Transfus*. 2011;25:1-6.
- [38] Edwards SG, Calandruccio JH. Autologous blood injection for refractory lateral epicondylitis. *J Hand Surg [Am]*. 2003;28(2):272-278.
- [39] Mishra A, Pavelko T. Treatment of chronic elbow tendinosis with buffered platelet-rich plasma. *Am J Sports Med*. 2006;34(11):1774-1778.
- [40] Kajikawa Y, Morihara T, Sakamoto H, et al. Platelet-rich plasma enhances the initial mobilization of circulation-derived cells for tendon healing. *Cell Physiol*. 2008;215(3):837-845.
- [41] Sánchez M, Anitua E, Azofra J, Aguirre JJ, Andia I. Intra-articular injection of an autologous preparation rich in growth factors for the treatment of knee OA: a retrospective cohort study. *Clin Exp Rheumatol*. 2008;26(5):910-913.