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***Posidonia oceanica* and *Zostera marina* as Potential Biomarkers of Heavy Metal Contamination in Coastal Systems**

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1. Introduction

In the early 1960s recognition of the adverse effects of environmental contamination due to industrial, pesticide, and agricultural pollution led to the emergence of the field of ecotoxicology (Ramade, 1992). Today, marine estuary and inshore ecosystems continue to be negatively impacted by environmental contamination (Short & Wyllie-Echeverria 1996; Orth et al., 2006; Osborn & Datta, 2006). In order to reduce these negative impacts, bio-surveillance programs are needed to monitor environmental conditions so that changes in ecosystem processes, structure, and the physiological condition of species can be assessed (Blandin, 1986; Tett et al., 2007). An important characteristic of these programs is that indicator species must be capable of rapidly detecting significant changes in the ecosystem so that the cause of deterioration can be addressed early (e.g. Hemminga & Duarte, 2000).

Mussels (Goldberg et al., 1983) and fish (Reichert et al., 1998; Stephensen et al., 2000) are frequently used as indicators of chemical contamination in long-term environmental monitoring programs. However, these programs can be deficient because they only provide information about water column contamination, and these organisms can have limited ranges and often must be introduced to a site as part of the monitoring program. To offset these deficiencies widely distributed indicator organisms in coastal systems that have the capacity to provide contamination information from both water column and sediment environments are needed. Consequently, there is increasing interest in the use of marine macrophytes because they grow in most coastal and estuarine systems (see Green & Short, 2003). These rooted vascular plants interact with the chemical properties of the water column and surface sediment environments within site-specific and basin-wide locations (Brix, 1997; Brix et al., 1983; den Hartog, 1970; Lange and Lambert, 1994; Rainbow and

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Phillips, 1993). For this study the focus was on the seagrasses *Posidonia oceanica* (L.) Delile (Posidoniaceae) and *Zostera marina* L. (Zosteraceae). These were chosen because they are the dominant species in the regions of our inquiry which were, respectively, the Mediterranean Sea (Lipkin et al., 2003; Procaccini et al., 2003) and the Pacific Northwest (Wyllie-Echeverria & Ackerman, 2003). Both species can form vast meadows across the intertidal-subtidal gradient in their respective ecosystems (Molinier & Picard, 1952; Phillips, 1984).

1.1 *P. oceanica* and *Z. marina* as indicators of environmental quality

The potential for these species to provide an early warning of deteriorating environmental quality has been noted for *P. oceanica* and *Z. marina* where both species were found useful at detecting environmental deterioration within local and basin-wide locations (Augier, 1985; Dennison et al., 1993; Pergent, 1991; Pergent-Martini et al., 1999). For example, *P. oceanica* accumulates certain metal pollutants, notably mercury (Pergent-Martini, 1998), which is one of the most abundant marine pollutants. Within the Mediterranean Sea elevated mercury levels have been reported in certain regions (Maserti et al., 1991), and correlations have been drawn between mercury levels in plant tissue and the concentrations of mercury in the water column (Pergent-Martini, 1998). In laboratory studies Lyngby & Brix (1984) and Brix & Lyngby (1984) demonstrated that *Z. marina* can accumulate heavy metals in concentrations above natural levels, and that these concentrations inhibited growth. In addition, based on extensive sampling along the coastline of Limfjord, Denmark, these authors noted that *Z. marina* could be used to monitor heavy metal contamination. Also, a related species *Z. capricorni* has provided valuable information in monitoring iron, aluminium, zinc, chromium, and copper contamination (Prange & Dennison, 2000).

Indicator species that provide an early warning of ecosystem change will likely be those that reveal first order changes in organism function. Molecular, biochemical, and/or cellular changes triggered by pollutants are measurable in biological mediums such as cells, tissues, and/or cellular fluids (McCarthy & Shugart, 1990). For example, oxidation is known to be a significant factor in stress-related organismal weakening, and antioxidant molecules have been used to evaluate organism health (Chen et al., 2007). One group of antioxidant molecules are the widely studied phenolic compounds (Ferrat et al., 2003a) which are known to be induced by reactive oxygen species (Rice-Evans et al., 1995; Vangronsveld et al., 1997).

1.2 Physiological and ecological roles of phenolics and volatile compounds

Phenolic compounds produced via the Shikimic Acid Pathway, and volatiles produced via the Mevalonate Pathway, are known to be important to plant health and survival (Cates, 1996; Fierer et al., 2001; Hartman, 2007; Phillips, 1992; Schimel et al., 1996). They are found in terrestrial higher plants, most notably angiosperms (Goodwin & Mercer, 1983; Hadacek, 2002), some seagrasses (Verges et al., 2007; Zapata and McMillan, 1979), and have a wide range of chemical structures and activities (Hadacek, 2002; Hartman, 2007). Phenolic and volatile compounds contribute significantly to the antioxidant activity of plants, have the capacity to bind heavy metals (Emmons et al., 1999), and are an important mechanism in protecting plants against stress (Swain, 1977). Volatile compounds (e.g. monoterpenes, sesquiterpenes) have been found to serve as energy sources in plants (Croteau & Sood,

1985), are important in the defensive system of higher plants (Cates, 1996; Langenheim, 1994), and influence ecosystem processes such as nutrient cycling (Horner et al., 1988; White, 1986). The production of phenolics and volatiles is under genetic control (Croteau & Gershenzon, 1994; Hartman, 2007), but their qualitative and quantitative production is affected by various environmental factors (Bryant et al., 1983; Gershenzon, 1984; Hartman, 2007; Macheix, 1996; Quackenbush et al., 1986; Ragan & Glombitza, 1986). However, as with other seagrasses, only a very limited number of studies deal with the role of phenolic and volatile compounds from *Posidonia oceanica* (Heglmeier & Zidorn, 2010) and *Zostera marina* (Short & Willie Echeverria, 1996). Only were investigated the impacts of interspecific competition (Dumay et al., 2004), nutrient variation, diseases (Vergeer & Develi, 1997) and grazing (Cannac et al., 2006), or general anthropization of water masses (Short & Wyllie-Echeverria, 1996, Agostini et al., 1998).

1.3 Objective

The objective of this study was to determine if *P. oceanica* and *Z. marina* might be reliable candidates as bio-surveillance organisms with regard to heavy metal pollution. We choose to consider different environmental conditions and to monitor physiological changes through two different seasons. Our assumption was that heavy metal contamination would adversely impact adult *P. oceanica* and *Z. marina* plants, and that plant response to these impacts could be assessed by differences in phenolic and volatile compound content of tissue from impacted and non-impacted sites.

We assessed differences in heavy metal content of plant tissues from sites with documented heavy metal pollution versus controls with no sources of heavy metal pollution. Then, we tested the hypothesis that the presence of identified contaminants could induce a bio-indicator response in these seagrass species. To do this we measured changes in total phenolic content in the leaf and sheath tissue of *P. oceanica*, and total phenolic and volatile compound content in above-and below-ground tissue of *Z. marina*.

2. Materials and methods

2.1 Site location and sample collection

In June 2000 and January 2001, 30 adult shoots of *P. oceanica* were collected by SCUBA at ~10 m in the sub-tidal region at two sites located in the northwestern Mediterranean Sea. The Bay of Bonifacio, a control site, is a pristine area relatively free of industrial pollution located in the south of Corsica (Tonnara - France; 41.4000 N; 9.0830 E; Capiomont et al., 2000). The Bay of Rosignano site south of Livorno (Italy; 43.4000 N; 10.4166 E) is a polluted site. At this site, a chlor-alkali plant has discharged industrial wastes rich in mercury since 1920 (130 kg per year; Ferrara et al., 1989). Water temperature ranged from 18°C in June 2000 to 14°C in January 2001 at all sites but salinity was relatively constant at 38.5 PSU within the study zone (i.e., 10 m depth contour; Villefranche sur Mer Observatory and Di Martino, personal communication).

For *P. oceanica*, foliage leaf and sheath tissue was analyzed for mercury and phenolic content. Tissue was obtained by separating the foliage leaf and sheath tissue from the roots and rhizomes following the procedure of Giraud (1977); root and rhizome tissue was discarded. The chlorophyllous foliage leaves were then separated from non-chlorophyllous

sheaths that are located at the leaf base. Foliage leaves from three adult shoots were dissected according to Giraud (1977) and combined to form one sample. Sheaths from the same three shoots were combined to form each sheath sample. After epiphytes were removed from leaf and sheath samples using a glass slide, each sample was rinsed with ultra-pure water and frozen (-20°C) until analysis. To determine mercury and phenolic content, we extracted 0.5 g dry wt. of each tissue sample ($n=10$).

During maximum low tide, *Z. marina* adult shoots were hand-collected from the lower intertidal region of two sites in Northern Puget Sound, Washington, USA in April and June 2000. The site located near Anacortes, WA (48.4263°N ; 122.5897°W) was documented as having heavy metal pollution (http://www.ecy.wa.gov/programs/wq/permits/permit_pdfs/dakota/factsheet.pdf), and the other was a pristine location with no industrial activity on the southeast side of Shaw Island (48.33942°N ; $122.55448^{\circ}\text{W}$) that served as the control site (Wyllie-Echeverria & Ackerman, 2003). Water temperature ranged between 9°C in April to 12°C in June 2000 at all sites, and salinity was relatively constant at 30 PSU during this time period (Wyllie-Echeverria, unpublished data).

Three samples were collected from each site, and each sample consisted of at least 0.5 g dry wt (Cuny et al., 1995) of eight to ten sterile (non-reproductive) shoots which were separated into above- and below-ground parts. Above-ground tissue consisted of the foliage leaf (i.e. basal leaf sheath and distal leaf blade; Kuo & den Hartog, 2006) excised from the rhizome at the node primordia (Tomlinson, 1974). The remaining rhizome and associated nodes and roots formed the below-ground sample. Epiphytes were scraped from the above-ground tissue and sediment was rinsed from the roots and rhizomes (Brackup & Capone, 1985). Each above- and below-ground sample was placed in labelled bags, kept moist and cool in a refrigerator, and shipped overnight to the Chemical Ecology Laboratory at Brigham Young University. Three replicate samples of above-ground tissue from each site, and three replicates of below-ground tissue from each site, were frozen at -80°C until extracted for heavy metals, phenolics or volatiles. Samples were stored at -80°C to preserve the volatile compounds in the tissues.

2.2 Qualitative and quantitative analysis of plant tissues for heavy metal content

Foliage leaves and sheaths of *P. oceanica* and above- and below-ground tissues of *Z. marina* were analyzed qualitatively and quantitatively for heavy metals. For *P. oceanica*, only mercury content, which is the predominant heavy metal pollutant at the Rosignano site (Lafabrie et al., 2007), was analyzed. Three individual shoots (three foliage leaves and three sheaths) that had been separately freeze dried were ground to a powder, and an aliquot of 0.05 g dry wt was digested. Digestion was performed in a 100-ml Teflon® advanced composite vessel reactor with 5 ml HNO_3 and 1 ml H_2O_2 (30%). Microwave digestion (Mars 5, CEM Chemistry, Engineering and Microwave, Matthews, NC, USA) was carried out using a temperature ramp of 8 min up to 200°C followed by a heating plateau of 20 min at 200°C . After digestion, the samples were increased to 25 ml with ultra-pure water and then filtered. Total mercury was determined using a flameless atomic absorption spectrophotometer flow injection (Perkin-Elmer System 100; Norwalk, CT, USA). The procedure consisted of reduction with 1.1% tin chloride ($\text{SnCl}_2 \cdot 2\text{H}_2\text{O}$) in 3% HCl and 0.5% hydroxylammonium chloride (NH_2OH , HCl). A standard addition method for total mercury was used to calibrate the protocol. The analytic procedure was verified using a moss as the certified

reference material (*Lagarosiphon major*, Certified Reference Material 60, Community Bureau of Reference, Commission of the European Community, Brussels, Belgium). Data are expressed as ng per g dry wt.

For *Z. marina*, heavy metal content was analyzed using the EPA Method 3052 Procedure. All elements were wet-ashed to prevent loss of elements and reduce the potential of confounding data due to silica content. Above- and below-ground tissue (0.5 g dry wt) was placed in a 50 ml folin tube, and 5 ml concentrated nitric acid was added. Samples sat overnight, and then were placed on a block digester at 200°C for 5-10 minutes. Tubes were removed, cooled, and then digested with 1 ml hydrofluoric acid. Samples were placed back on the block digester for 45-60 minutes. Tubes were removed and brought to a 50 ml volume with distilled water. Stoppered tubes were shaken and then analyzed by inductively coupled plasma atomic emission spectrometry (Iris Intrepid II XSP, model 14463001; Thermo Electron Corporation, Franklin, MA) equipped with an ASX-520 autosampler. Data are expressed as ppm (Table 2).

2.3 Extraction and determination of phenolic content in the tissues of *P. oceanica*, and phenolic and volatile content of above-ground tissues of *Z. marina*

Total phenolic content for both species, and total volatile content for *Z. marina*, were determined to ascertain whether tissue collected from impacted (heavy metal pollution for both species) and control sites differed. A different method is used for the definition of the phenolic and volatile compounds, because the measurements were realized in different labs. For *P. oceanica*, extraction of total phenolic compounds was carried out on 0.5 g dry wt freeze-dried foliage leaf or sheath tissue. Extraction followed Cuny et al. (1995) and consisted of infusing each sample at 40°C in 50 % (v/v) aqueous ethanol in darkness for 3 h. The extract was acidified with a few drops of 2N HCl, the ethanol was evaporated under vacuum, and the aqueous residue extracted with ethanol/acetic acid. The organic phase was dried using anhydrous Na₂SO₄. Concentration of total phenolic compounds was measured by colorimetry (Swain & Hillis, 1959) using Folin-Denis reagent (Folin and Dennis, 1915). Phloroglucinol (Frantzis, 1992) was used for elaboration of standard curves. For *Z. marina*, phenolics were extracted using MeOH/CH₂CH₂ (50/50) from 200 mg dry wt of freeze dried above-ground tissue, filtered using VWR grade 415 filter paper, and blown dry using nitrogen gas to prevent oxidation. After redissolving in MeOH/CH₂CH₂ (50/50), the extract was again filtered, placed in an auto-sampler vial (Chromatography Research Supplies, Addison, IL) and injected into a high pressure liquid chromatograph (HPLC) (HP Model 1100; Agilent 1100 Series, Model G1313A; Santa Clara, CA) equipped with a diode-array detector (Model G1316A) and a C₁₈ reverse phase 5µm column (Phenomenex, Torrance, CA). The HPLC solvents were A = water/acetic acid (98:2); B = acetonitrile/acetic acid (98:2). Temperature was 50°C, flow rate 1ml/min, and wavelength of the detector set at 280 nm (optimized for *Z. marina* phenolic compounds). Phenolic content is expressed as total peak height /200 mg dry wt. To obtain volatile compound content in *Z. marina* samples, 3 g fresh wt of above-ground tissue was ground to a fine powder in liquid nitrogen and hexane. The extract was then filtered, and the filtrate injected into a capillary gas chromatograph (HP Model 6890) equipped with a head-space sampler (Perkin-Elmer HS 40 XL; Waltham, MA) and a HP-1 column. Oven temperature was 80°C, needle temperature 85°C, transfer temperature 120°C, thermostat time 10 min, pressurizing time 0.6 min, injection time 0.2

min, and withdrawal time 0.5 min. The ramp GC program was 40-210°C at intervals of 3°C ramp/min. Total volatile compound content is expressed as total peak height per 3 g fresh wt tissue.

2.4 Statistical analysis

Data from *P. oceanica* samples were analyzed using a three-way ANOVA to allow comparisons between the phenolic compounds and mercury levels according to tissue, site and sampling period. Since the interaction among these factors was significant, one-way analyses followed by a Tukey test (for analyses over the annual cycle) or Student-t test (for analyses of tissue and site factors at given months) were performed (Zar, 1999). Normality and homoscedasticity were verified by Shapiro Wilks and Bartlett tests, respectively (Zar, 1999). The relationships between phenolic compounds and mercury level were assessed using correlation and regression analyses in Statgraphics plus (ver 3.1) for Windows. Data from *Z. marina* are expressed as ppm for heavy metals, total peak area per 200 mg freeze dried tissue for phenolics, and total peak area per 3 g fresh wt for volatiles. Since all samples were randomly collected along a transect, each sample is treated as an independent experimental unit. Comparison of heavy metal content between impacted and control sites in *Z. marina* above- and below-ground tissues, and for phenolic and volatile content in above-ground tissues, was conducted using a one-way ANOVA, SAS GLM program (SAS, 1996).

3. Results

3.1 Site and tissue differences in heavy metal contamination

Foliage leaf and sheath tissue of *P. oceanica* from the industrially impacted Rosignano site showed large and significant ($p<0.05$) differences in mercury content when compared to the control Tonnara site (Table 1).

Tissue Type		Mercury impacted site (Rosignano)	Control site (Tonnara)
Foliage Leaves	June 2000	233 ± 23	77 ± 11
	January 2001	317 ± 41	79 ± 15
Sheaths	June 2000	368 ± 26	64 ± 8
	January 2001	215 ± 16	80 ± 19

Table 1. Mercury levels (ng/g dw) in foliage leaf and sheath tissues of *P. oceanica* collected at different sites and different sampling periods.

Samples of above-ground tissue collected in April 2000 from *Z. marina* plants growing in the impacted site were higher in iron, aluminium, and copper when compared to tissue from the control site (Table 2). However, above-ground tissue from the control site was significantly higher in zinc, nickel, molybdenum, and mercury when compared to the impacted site (Table 2). For the July 2000 samples, the only significant differences were that nickel and copper were in highest concentration in plants from the impacted site when compared to plants from control site (Table 2). For below-ground tissue of *Z. marina* in April, samples from the industrially impacted site were significantly higher ($p<0.05$) for iron, aluminium, nickel, manganese, copper, cadmium, chromium, and lead when

compared to the control site (Table 2). None of the heavy metals was higher in concentration in the control site for samples taken in April 2000. For the July 2000 samples, barium, iron, aluminium, zinc, manganese, copper, cadmium, arsenic, and chromium were higher in the plants from the impacted site when compared to the control site, and cobalt and strontium were higher in plants from the control site (Table 2).

Heavy Metals	Site (ppm)*							
	Above-ground Tissue				Below-ground Tissue			
	April		July		April		July	
	Industrially impacted site	Control site	Industrially impacted site	Control site	Industrially impacted site	Control site	Industrially impacted site	Control site
Barium	323(41) ^a	364(32) ^a	279(107) ^a	312(55) ^a	466(136) ^a	420(100) ^a	570(148) ^a	315(84) ^b
Iron	320(127) ^a	180(58) ^b	204(87) ^a	142(94) ^a	5801(2846) ^a	1068(540) ^b	5591(1503) ^a	576(263) ^b
Aluminum	183(75) ^a	119(43) ^b	88(42) ^a	100(81) ^a	1626(1341) ^a	665(435) ^b	1737(494) ^a	503(336) ^b
Zinc	100(11) ^a	119(13) ^b	102(22) ^a	110(15) ^a	134(45) ^a	133(44) ^a	169(46) ^a	96(16) ^b
Nickel	55(21) ^a	104(25) ^b	45(16) ^a	23(15) ^b	127(78) ^a	34(13) ^b	63(20) ^a	64(31) ^a
Manganese	37(6) ^a	42(6) ^a	48(16) ^a	51(7) ^a	38(33) ^a	11(6) ^b	26(7) ^a	10(4) ^b
Copper	14(2) ^a	12(2) ^b	16(4) ^a	10(1) ^b	40(21) ^a	19(29) ^b	43(12) ^a	10(3) ^b
Molybdenum	5(2) ^a	6(1) ^b	7(2) ^a	8(1) ^a	0(0)	----**	0(0) ^a	0(1) ^a
Cadmium	2(1) ^a	1(1) ^a	2(1) ^a	2(1) ^a	13(8) ^a	4(2) ^b	11(3) ^a	3(1) ^b
Arsenic	4(2) ^a	3(2) ^a	3(2) ^a	3(2) ^a	8(7) ^a	8(1) ^a	10(6) ^a	1(2) ^b
Cobalt	2(1) ^a	4(1) ^a	2(1) ^a	2(1) ^a	2(1) ^a	1(1) ^a	1(1) ^a	2(1) ^b
Mercury	1(1) ^a	2(1) ^b	0(1) ^a	1(1) ^a	3(3) ^a	2(1) ^a	4(1) ^a	4(2) ^a
Strontium	1(2) ^a	2(3) ^a	4(3) ^a	4(2) ^a	3(4) ^a	6(4) ^a	0(0) ^a	1(2) ^b
Chromium	1(1) ^a	2(1) ^a	1(0) ^a	1(0) ^a	7(4) ^a	2(1) ^b	6(2) ^a	1(1) ^b
Lead	0(0) ^a	1(2) ^a	0(0) ^a	0(0) ^a	13(23) ^a	6(3) ^b	0(1) ^a	0(1) ^a

Table 2. Differences in accumulation of heavy metals in above- and below- ground tissues of *Z. marina* between impacted and control sites [April, July 2000; x, -]. *Means followed by different letters are significantly different at $p < 0.05$; Means followed by the same letter (i.e. “a”) are not significantly different at $p < 0.05$. **Insufficient sample for analysis.

3.2 Production of phenolic and volatile compound content in plant tissues between impacted and control sites

Foliage leaves from Tonnara (20.5 mg.g⁻¹) were significantly higher (Tukey test, $p < 0.05$) in phenolic content in January 2001 compared to plants from the mercury impacted Rosignano site (13.2 mg.g⁻¹), but were not significantly different in the June 2000 samples (Fig. 1). For sheaths, the levels of total phenolic compounds from Tonnara plants in June and January (9.2 and 15.2 mg.g⁻¹, respectively) were significantly higher than those measured in plants at

the Rosignano site (5.0 and 6.4 mg.g⁻¹, respectively) (Tukey test, $p<0.05$). Phenolic content was higher across sites and sampling times in *P. oceanica* foliage leaves compared to sheaths in all comparisons (Mann and Whitney test, $p>0.05$).

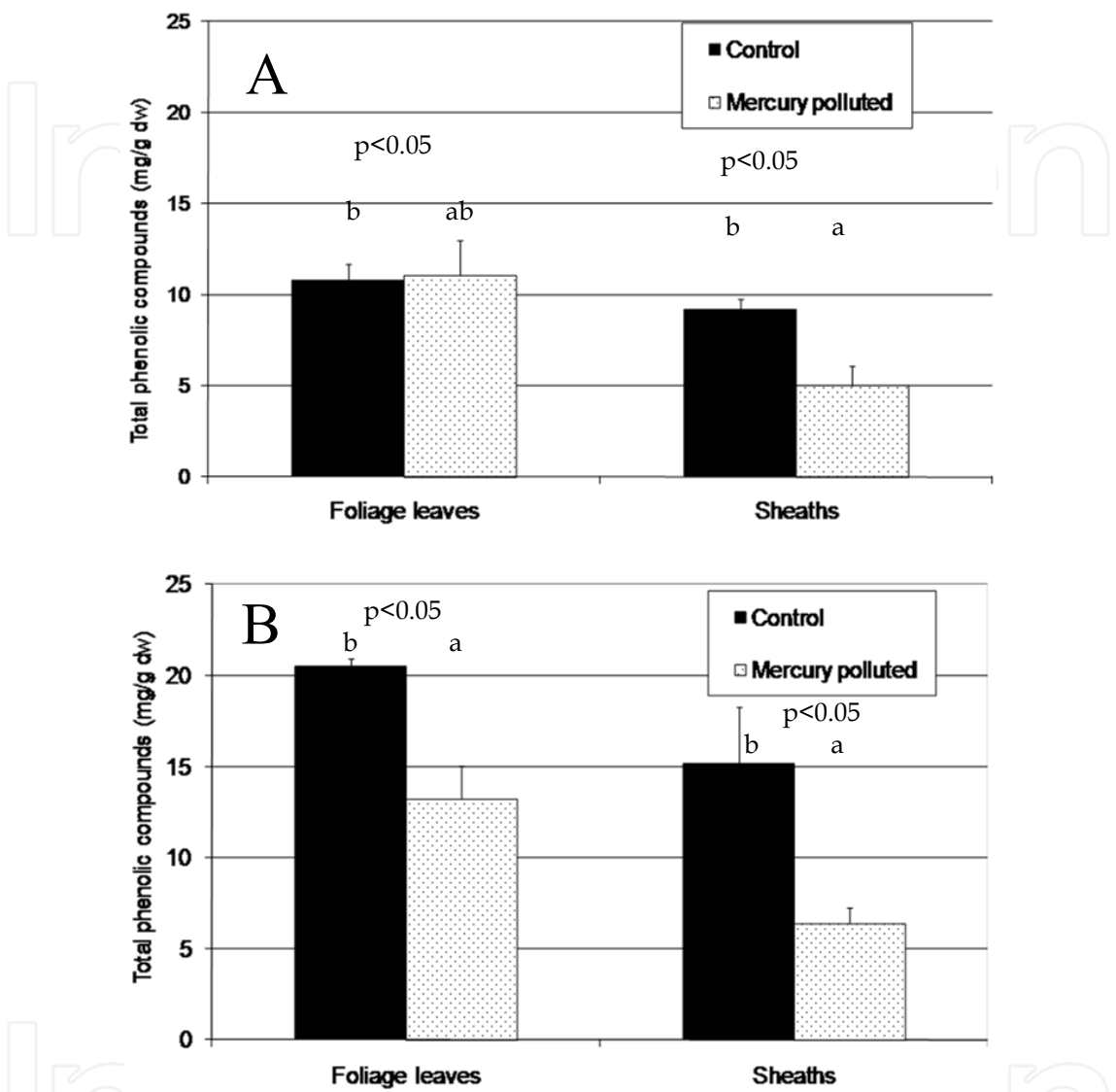


Fig. 1. Total phenolic concentration (mg.g⁻¹ dw) in foliage leaf and sheath tissues of *P. oceanica* in Tonnara (control) and Rosignano (mercury polluted) in June 2000 (A) and January 2001 (B).

For *Z. marina* total phenolic content in above-ground tissues collected from plants at the control site always was higher when compared to above-ground tissues collected from the impacted site for both April and July 2000 (Fig. 2). However, the only significant difference was in July where the control site produced a higher amount of total phenolic (65.8 vs 50.8 peak area / 200 mg dry wt, respectively; $p<0.05$). Total volatile compound production also was higher at both sampling periods, but the only significant difference occurred in the April 2000 sampling where above-ground tissues from the control site showed an average peak area of 551 per 200 mg dry wt tissue compared to 352 at the impacted site ($p<0.05$).

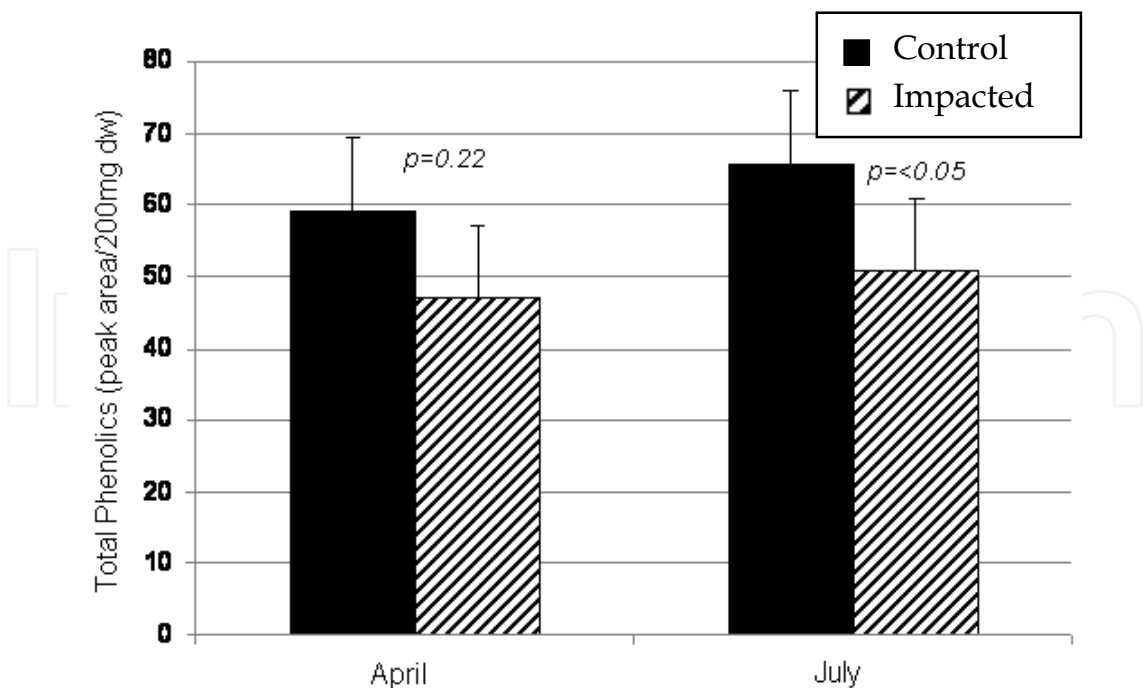


Fig. 2. Total phenolic content in above-ground tissue from *Z. marina* plants growing in heavy metal impacted and control sites (April and July, 2000).

4. Discussion

4.1 Tissue and site differences in heavy metal content

Results presented indicate that plant tissues of *P. oceanica* and *Z. marina* significantly accumulated high levels of heavy metals when growing on heavy metal-impacted sites (Tables 1 & 2). At the Rosignano site, when compared to the control Tonnara site, foliage leaves and sheaths contained two to over six times the amount of mercury. These patterns of accumulation are consistent with findings by other authors who have studied the same sites (Capiomont et al., 2000; Ferrat, 2001; Ferrat et al., 2003b; Maserti & Ferrara, 1991).

Z. marina plants from the heavy-metal impacted site accumulated significantly higher concentrations of iron, aluminum, nickel, and copper in their above-ground tissues when compared to the control site (Table 2). In addition, below-ground tissue of *Z. marina* plants from the industrially-impacted site accumulated over three, and up to five, times the levels of heavy metals compared to plants from the control site. A striking difference between above- and below-ground tissue, is that below-ground tissue from the impacted site accumulated 12 heavy metals (barium, iron, aluminum, zinc, nickel, manganese, copper, cadmium, arsenic, cobalt, chromium, lead; Table 2) while above-ground tissue only accumulated four heavy metals (iron, aluminum, nickel, copper) (Table 2). Another major difference is that the quantity of heavy metals accumulated in the below-ground tissue was higher for most of the heavy metals compared to that in the above-ground tissue.

Variation in metallic accumulation between above- and below-ground seagrass tissue has been discussed by various authors (see synthesis in Pergent Martini & Pergent, 2000), and could be a function of differences in binding sites or seasonal translocation between above- and below-ground structures (Libes & Boudouresque, 1987; Ward, 1987). The level of

environmental contamination within a particular site also may be an important factor. For example Capiomont et al. (2000) found that mercury content was higher in the interstitial water than in the water column at our Rosignano sampling location.

Heavy metals are known to have adverse affects on the physiology of *P. oceanica* and *Z. marina* as well as other seagrasses (Ward, 1987). Lyngby & Brix (1984) have shown that the order of heavy metal inhibition of growth of *Z. marina* from greatest to least is mercury, copper, cadmium, zinc, chromium, and lead. Interestingly mercury was not significantly accumulated by *Z. marina* at our impacted site but the other five generally followed the pattern described by Lynby & Brix (1984) (Table 2).

4.2 Phenolic and volatile compound production in plant tissues between impacted and control sites

Our results suggest that total phenolic compound levels within seagrass tissue could be an indicator of site quality. Differences in production of phenolics in tissues from both species were noted between impacted and control sites. For foliage leaves and sheaths of *P. oceanica* collected in January, and above-ground tissue of *Z. marina* collected in July, total phenolic content was significantly lower in plants collected from industrial sites (Fig. 1 & 2). This is supported by Vergeer et al. (1995) who concluded that a decrease of total phenolic compounds in the tissue of *Z. marina* indicated plants may be growing in unsuitable environmental conditions. Noteworthy is that correlation analysis indicated a significant ($p < 0.05$) inverse relationship between heavy metal content and the health of plants as measured by phenolic content for *P. oceanica* ($r^2 = 69.8\%$, linear model of regression: mercury = $0.22 - 0.0055 * \text{phenol}$ for sheaths).

Additionally, gas chromatography analysis of volatile compounds from *Z. marina* indicated that above-ground tissue from plants growing in the impacted site was significantly lower in volatiles from the April collection, when growth begins in Northern Puget Sound (Phillips, 1984) compared to tissue from the control site (Fig. 3). However, no significant differences occurred in volatile compound production between impacted and control sites in the July collection.

4.3 Phenolic compound production with regard to tissue and time collection

For *P. oceanica*, the concentration of phenolic compounds differed between foliage leaves and sheaths being higher in leaf tissue regardless of site. Similarly, Agostini et al. (1998) found higher concentrations (6 mg.g^{-1}) in the apical parts and youngest leaves and lower concentrations in sheaths (0.1 mg.g^{-1}). Also, in our study significant variation was observed between seasons; for example, phenolic levels were found to be higher in the January 2001 samples compared to the June 2000 samples.

Differences occur in the natural products analyzed depending on month of collection for both *P. oceanica* and *Z. marina* (Fig. 1-3). For example, *P. oceanica* foliage leaves and sheaths in January 2001 were higher in phenolic content than those collected in June 2000 (Fig. 1). While phenolic content in above-ground *Z. marina* tissue was similar in concentration between April and July (Fig. 2), but volatile compounds in above-ground tissue collected in April were significantly higher than those collected in July (Fig. 3). April and July were selected as sampling times for *Z. marina* because they represent early and mature tissue

growth in the Northern Puget Sound (Phillips, 1984). However, in a preliminary study in which *Z. marina* shoots were collected in February 2000, plants from the heavy metal-impacted site produced only 19% of the total phenolic content when compared to plants from the control site (Zou et al., unpublished data). In order to establish when phenolics and volatiles may best indicate plant health, experimental designs need to involve sampling plants every two months throughout the year.

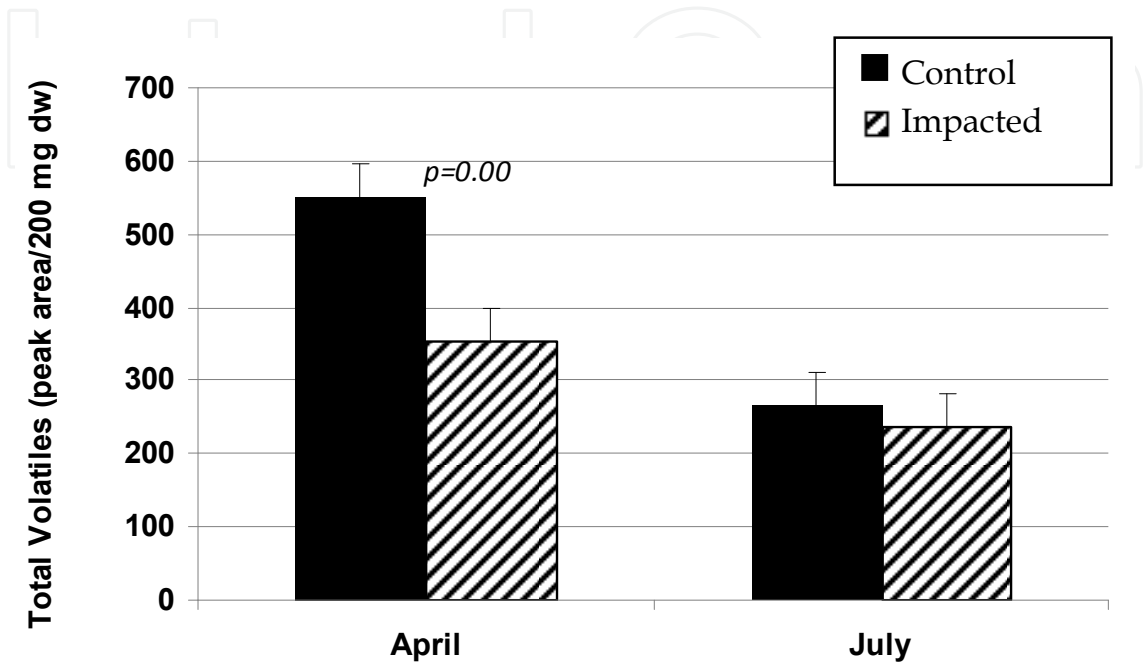


Fig. 3. Total volatile content of above-ground tissue from *Z. marina* plants growing in heavy metal impacted and control sites (April and July, 2000).

Finally, based on the response of different seagrass genotypes to disturbance (e.g. Ehlers et al., 2008; Hughes & Stachowicz, 2009; Wyllie-Echeverria et al., 2010), we suspect that variation in the type and concentration of heavy metal uptake may exist within different genotypes. However, this aspect of heavy metal accumulation in needs investigation in controlled conditions with seagrass species from different locations.

5. Conclusions

Significant differences were found in the accumulation of mercury in leaf and sheath tissues of *P. oceanica* when plants were growing on impacted sites as compared to sites not impacted heavily by mercury (Table 1). *Z. marina* plants growing in a site impacted by heavy metals associated with industrial pollution accumulated significantly higher amounts of iron, aluminium, nickel, and copper in above-ground tissues as compared to a non-impacted site, and higher amounts of barium, iron, aluminium, zinc, nickel, manganese, copper, cadmium, arsenic, chromium, and lead in below-ground tissues at the impacted site (Table 2). For *P. oceanica*, total phenolics were significantly higher in leaves at the control site when compared to the mercury impacted site for the January sampling period (Fig. 1). For sheath tissue total phenolics from the control site were significantly higher when compared to the mercury impacted site for both sampling periods (Fig. 1). For *Z. marina*, total phenolic content was higher in both sampling periods at the non-impacted site compared to the

control site, but only significantly so for the July 2000 sampling period (Fig. 2). Total volatile content also was higher at the control site for both sampling periods, but only significantly higher for the April sampling period (Fig. 3). These results support the hypotheses that *P. oceanica* and *Z. marina* accumulate significant amounts of heavy metals from impacted sites, and that these accumulations are associated with reduced total phenolic and volatile compound content. Based on these supportive data, we conclude that *P. oceanica* and *Z. marina* are potential candidates as bio-surveillance organisms especially with regard to heavy metal pollution of coastal and estuarine ecosystems.

Since we observed variation in the production of phenolics and volatiles with regard to sampling time and season, a priority is the identification of individual phenolic and volatile compounds in the tissue of these two species. In our labs we have identified in one or both species using gas chromatography/mass spectroscopy and high pressure liquid chromatography several cinnamic acid and benzoic acid derivatives; these results are comparable to those found by Quackenbush et al. (1986). Additionally, these analyses indicate not only a quantitative decrease in total phenolic and volatile compounds, but also qualitative differences between plants growing on impacted and non-impacted sites (Ferrat et al., unpublished data for *P. oceanica*; Zou et al., unpublished data for *Z. marina*). Finally, since various environmental perturbations may adversely affect seagrass health (impact of human activity reviewed in Short & Wyllie-Echeverria, 1996), and thereby phenolic and volatile compound production, collaboration among scientists working at a diversity of sites would greatly facilitate progress toward this bio-surveillance effort.

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7. References

- Agostini, S.; Desjobert, J.M. & Pergent, G. (1998). Distribution of phenolic compounds in the seagrass *Posidonia oceanica*. *Phytochemistry*, Vol.48, No.4, (June 1998), pp. 611-617, ISSN 0031-9422
- Augier, H. (1985). L'herbier à *Posidonia oceanica*, son importance pour le littoral Méditerranéen, sa valeur comme indicateur biologique de l'état de santé de la mer, son utilisation dans la surveillance du milieu, les bilans écologiques et les études d'impact. *Vie Marine*, Vol.7, (1985), pp. 85-113
- Blandin, P. (1986). Bioindicateurs et diagnostic des systèmes écologiques. *Bulletin d'Ecologie*, Vol.17, No.4, (April 1986), pp. 211-307, ISSN 0395-7217
- Brackup, I. & Capone, D.G. (1985). The effect of several metal and organic pollutants on nitrogen-fixation (acetylene reduction) by the roots and rhizomes of *Zostera marina* L. *Environmental and Experimental Botany*, Vol.25, No.2, (May 1985), pp. 145-151, ISSN 0098-8472

- Brix, H.; Lyngby, J.E. & Schierup, H.H. (1983). Eelgrass (*Zostera marina* L.) as an indicator organism of trace metals in the Limfjord, Denmark. *Marine Environmental Research*, Vol.8, No.3, (March 1983), pp. 165-181, ISSN 0141-1136
- Brix, H. & Lyngby, J.E. (1984). A survey of the metallic composition of *Zostera marina* L. in the Limfjord, Denmark. *Archiv für Hydrobiologie*, Vol.99, No.3, (March 1984), pp. 347-359, ISSN 0003-9136
- Brix, H. (1997). Do macrophytes play a role in constructed treatment wetlands? *Water Science and Technology*, Vol.35, No.5, (May 1997), pp. 11-17, ISSN 0273-1223
- Bryant, J.P.; Chapin F.S. & Klein, D.R. (1983). Carbon/nutrient balance of boreal plants in relation to vertebrate herbivory. *Oikos*, Vol.40, No.3, (March 1983), pp. 357-368, ISSN 0030-1299
- Capiomont, A.; Piazzzi, L. & Pergent, G. (2000). Seasonal variations of total mercury in foliar tissues of *Posidonia oceanica*. *Journal of the Marine Biological Association of the United Kingdom*, Vol.80, No.6, (June 2000), pp. 1119-1123, ISSN 0025-3154
- Cates, R.G. (1996). The role of mixtures and variation in the production of terpenoids in conifer-insect-pathogen interactions, In: *Phytochemical Diversity and Redundancy in Ecological Interactions, Serie Recent Advances in Phytochemistry*, Romeo, J.T.; Saunders, J.A.; Barbosa, P., Vol.30, pp. 179-216, Plenum Press, ISBN 978-0-306-45500-1, New York
- Chen, C.; Arjomandi, M.; Balmes, J.; Tager, I. & Holland, N. (2007). Effects of chronic and acute ozone exposure on lipid peroxidation and antioxidant capacity in healthy young adults. *Environmental Health Perspectives*, Vol.115, No.12, (December 2007), pp. 1732-1737, ISSN 0091-6765
- Croteau, R. & Gershenzon, J. (1994). Genetic control of monoterpene biosynthesis in mints (*Mentha*: Lamiaceae), In: *Genetic Engineering of Plant Secondary Metabolism, Serie Recent Advances in Phytochemistry*, Ellis, B.E.; Kuroki, G.W.; Stafford, H.A., Vol.28, pp. 193-229, Plenum Press, ISBN 978-0-306-44804-1, New York
- Croteau, R. & Sood, V. (1985). Metabolism of monoterpenes. Evidence for the function of monoterpenes and catabolism in peppermint (*Mentha piperita*). *Plant Physiology*, Vol.77, No.4, (April 1985), pp. 801-806, ISSN 0032-0889
- Cuny, P.; Serve, L.; Jupin, H. & Boudouresque, C.F. (1995). Water soluble phenolic compounds of the marine phanerogam *Posidonia oceanica* in a Mediterranean area colonized by the introduced chlorophyte *Caulerpa taxifolia*. *Aquatic Botany*, Vol.52, No.3, (December 1995), pp. 237-242, ISSN 0304-3770
- den Hartog, C.D. (1970). *The Seagrasses of the World*, Verhand Koninklijke Nederl Akad. Wetenschap Afd. Nat. Tweede Reeks, North-Holland Publication, Amsterdam
- Dennison, W.C.; Orth, R.J.; Moore, K.A.; Stevenson, J.C.; Carter, V.; Kollar, S.; Bergstrom, P.W. & Batiuk, R.A. (1993). Assessing water quality with submerged aquatic vegetation: Habitat requirements as barometers of Chesapeake Bay health. *BioScience*, Vol.43, No.2, (February 1993) pp. 86-94, ISSN 0006-3568
- Ehlers, A.; Worm, B. & Reusch, T.B.H. (2008). Importance of genetic diversity in eelgrass *Zostera marina* for its resilience to global warming. *Marine Ecology Progress Series*, Vol.355, (February 2008), pp. 1-7, ISSN 0171-8630
- Emmons, C.L.; Peterson, D.M. & Paul, G.L. (1999). Antioxidant capacity of oat (*Avena sativa* L.) extracts. 2. *In vitro* antioxidant activity and contents of phenolic and tocol antioxidants. *Journal of Agricultural and Food Chemistry*, Vol.47, (December 1999), pp. 4894-4898, ISSN 0021-8561

- Fierer, N.; Schimel, J.P.; Cates, R.G. & Zou, J. (2001). Influence of balsam poplar tannin fractions on carbon and nitrogen dynamics in Alaskan taiga floodplain soils. *Soil Biology and Biochemistry*, Vol.33, No.12-13, (October 2001), pp. 1827-1839, ISSN 0038-0717
- Ferrat, L. (2001). *Réactions de la phanérogame marine Posidonia oceanica en réponse à des stress environnementaux*, Thèse de doctorat, Université de Corse, France
- Ferrat, L.; Pergent-Martini C. & Roméo M. (2003a). Assessment of the use of biomarkers in aquatic plants for the evaluation of environmental quality: application to seagrasses. *Aquatic Toxicology*, Vol.65, No.2, (October 2003), pp. 187-204, ISSN 0166-445X
- Ferrat, L.; Gnassia-Barelli, M.; Pergent-Martini, C. & Roméo, M. (2003b). Mercury and non-protein thiol compounds in the seagrass *Posidonia oceanica*. *Comparative Biochemistry and Physiology c*, Vol.134, No.1, (January 2003), pp. 147-155, ISSN 0742-8413
- Ferrara, R.; Maserti, B.E. & Paterno, P. (1989). Mercury distribution in marine sediment and its correlation with the *Posidonia oceanica* prairie in a coastal area affected by a chlor-alkali complex. *Toxicological and Environmental Chemistry*, Vol.22, No.1-4, (April 1989), pp. 131-134, ISSN 0277-2248
- Folin, O. & Denis, W. (1915). A colorimetric method for the determination of phenols (and phenol derivatives) in urine. *Journal of Biological Chemistry*, Vol.22, (1915), pp. 305-308, ISSN 0021-9258
- Frantzis, A. (1992). *Etude expérimentale des niveaux de consommation et d'utilisation des macrophytes et des détritux dérivés par deux invertébrés benthiques Paracentrotus lividus et Abra ovata*, Thèse de doctorat, Université Aix Marseille II, France
- Gershenzon, J. (1984). Changes in the level of plant secondary metabolite production under water and nutrient stress. In: *Phytochemical adaptation to stress, Serie Recent Advances in Phytochemistry*, Loewus F.A.; Timmermann B.N.; Steelink C., Vol.24, pp. 273-320, Plenum Press, ISBN 0306417200, New York
- Giraud, G. (1977). *Contribution à la description et à la phénologie quantitative des herbiers à Posidonia oceanica (L.) Delile*, Thèse de Doctorat 3ème cycle, Université Aix-Marseille II, France
- Goldberg, E.D.; Koide, M. & Hodeg, V. (1983). U.S. Mussel watch: 1977-1978 results on trace metals and radionuclides. *Estuarine Coastal and Shelf Science*, Vol.16, No.1, (January 1983), pp. 69-93, ISSN 0272-7714
- Goodwin, T.W. & Mercer, E.I. (1983). *Introduction to Plant Biochemistry*, Pergamon Press, Oxford, ISBN 0080249221 9780080249223 0080249213 9780080249216, England
- Green, E.P. & Short, F.T. (2003). *World Atlas of Seagrasses*, UNEP World Conservation Monitoring Centre, University of California Press, Berkeley, ISBN 0-520-24047-2, USA
- Hadacek, F. (2002). Secondary Metabolites as Plant Traits: Current Assessment and Future Perspectives. *Critical Reviews in Plant Sciences*, Vol.21, No.4, (April 2002), pp. 273-322, ISSN 0735-2689
- Hartman, T. (2007). *From waste products to ecochemicals: fifty years research of plant secondary metabolism*. *Phytochemistry*, Vol.68, No.22-24, (November-December 2007), pp. 2831-2846, ISSN 0031-9422

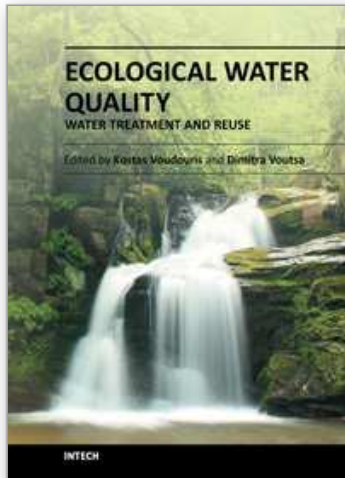
- Heglmeier, A. & Zidorn, C. (2010). Secondary metabolites of *Posidonia oceanica* (Posidoniaceae). *Biochemical Systematics and Ecology*, Vol.38, No.5, (May 2010), pp. 1-7, ISSN 03051978
- Hemminga, M. & Duarte, C. (2000). *Seagrass Ecology*, Cambridge University Press, ISBN 0-521-66184-6, USA
- Horner, J.D.; Gosz, J.R. & Cates, R.G. (1988). *The role of carbon-based plant secondary metabolites in decomposition in terrestrial ecosystems*. *American Naturalist*, Vol.132, No.6, (June 1988), pp. 869-883, ISSN 0003-0147
- Hughes, A.R. & Stachowicz, J.J. (2009). Ecological impacts of genetic diversity in the clonal seagrass *Zostera marina*. *Ecology*, Vol.90, No.5, (May 2009), pp. 1412-1419, ISSN 0012-9658
- Kuo, J. & den Hartog, C. (2006). Seagrass morphology, anatomy and ultrastructure, In: *Seagrasses: Biology, Ecology, and Conservation*, Larkum, A.W.D.; Orth, R.J.; Duarte, C.M., pp. 51-87, Springer-Verlag, ISBN 978-1402029424, The Netherlands
- Lafabrie, C.; Pergent, G.; Kantin, R.; Pergent-Martini, C. & Gonzalez, J-L. (2007). Trace metals assessment in water, sediment, mussel and seagrass species – Validation of the use of *Posidonia oceanica* as a metal biomonitor. *Chemosphere*, Vol.68, No.11, (August 2007), pp. 2033-2039, ISSN 0045-6535
- Lange, C.R. & Lambert, K.E. (1994). Biomonitoring. *Water Environmental Research*, Vol.66, No.4, (June 1994), pp. 642-651, ISSN 1061-4303
- Langenheim, J. (1994). Higher plant terpenoids: A phytocentric overview of their ecological roles. *Journal of Chemical Ecology*, Vol.20, No.6, (June 1994), pp. 1223-1280, ISSN 0098-0331
- Libes, M. & Boudouresque C.F. (1987). Uptake and long-distance transport of carbon in the marine phanerogram *Posidonia oceanica*. *Marine Ecology Progress Series*, Vol.38, (June 1987), pp. 177-186, ISSN 0171-8630
- Lipkin, Y.; Beer S. & Zakai D. (2003). The seagrasses of the Eastern Mediterranean and the Red Sea, In: *World Atlas of Seagrasses*, Green, E.P.; Short F.T., pp. 65-73, UNEP World Conservation Monitoring Centre, University of California Press, Berkeley, ISBN 0-520-24047-2, USA
- Lyngby, J.E. & Brix, H. (1984). The uptake of heavy metals in eelgrass *Zostera marina* and their effect on growth. *Ecological Bulletins*, Vol.36, (1984), pp. 81-84
- Macheix, J.J. (1996). Les composés phénoliques des végétaux : quelles perspectives à la fin du XXème siècle? *Acta Botanica Gallica*, Vol.143, No.6, (June 1996), pp. 473-479, ISSN 1253-8078
- Maserti, B.E. & Ferrara, R. (1991). Mercury in plants, soil and atmosphere near a chlor-alkali complex. *Water, Air, Soil Pollution*, Vol.56, No.1, (April 1991), pp. 15-20, ISSN 1567-7230
- Maserti, B.E.; Ferrara, R. & Morelli, M. (1991). *Posidonia oceanica*: uptake and mobilization of mercury in the Mediterranean basin. In: *Proceedings of the FAO/UNEP/IAEA Workshop on the biological effects of pollutants on marine organisms*, Gabrielides, G.P., Vol. 59, pp. 243-249, MAP Technical Reports Series, Athens, Greece
- McCarthy, J.F. & Shugart, L. (1990). *Biomarkers of Environmental Contamination*, Lewis Publishers: Boca Raton, ISBN 0873712846, Florida, USA

- Molinier, R. & Picard, J. (1952). Recherches sur les herbiers de phanérogames marines du littoral méditerranéen français. *Annales de l'Institut Océanographique*, Vol.27, No.3, (1952), pp. 157-234, ISSN 0078-9682
- Orth, R.J.; Carruthers, T.J.B.; Dennison, W.C.; Duarte, C.M.; Fourqurean, J.W.; Heck, Jr.K.L.; Hughes, A.R.; Kendrick, G.A.; Kenworthy, W.J.; Olyarnik, S.; Short, F.T.; Waycott, M. & Williams, S.L. (2006). A global crisis for seagrass ecosystems. *Bioscience*, Vol.56, No.12, (December 2006), pp. 987-996, ISSN 0006-3568
- Osborn, D. & Datta A. (2006). Institutional and policy cocktails for protecting coastal and marine environments from land-based sources of pollution. *Ocean and Coastal Management*, Vol.49, No.9-10, (January 2006), pp. 576-596, ISSN 0964-5691
- Pergent, G. (1991). Les indicateurs écologiques de la qualité du milieu marin en Méditerranée. *Oceanis*, Vol.17, No.4, (April 1991), pp. 341-350, ISSN 0182-0745
- Pergent-Martini, C. (1998). *Posidonia oceanica*: a biological indicator of past and present mercury contamination in the Mediterranean Sea. *Marine Environmental Research*, Vol.45, No.2, (March 1998), pp. 101-111, ISSN 0141-1136
- Pergent-Martini, C. & Pergent G. (2000). Are marine phanerogams a valuable tool in the evaluation of marine trace-metal contamination: example of the Mediterranean sea? *International Journal of Environment and Pollution*, Vol.13, No.1-6, (January 2000), pp. 126-147, ISSN 0957-4352
- Pergent-Martini, C.; Pergent, G.; Fernandez, C. & Ferrat, L. (1999). Value and use of *Posidonia oceanica* as a biological indicator, In: *Proceedings MEDCOAST-EMECS 99 Joint Conference Land-ocean interactions: managing coastal ecosystems*, Vol.1, pp. 73-90, MEDCOAST, Middle East Technical University Publication, ISBN 975-429-142-X, Ankara, Greece
- Phillips, D. (1992). Flavonoids: plant signals to soil microbes. In: *Phenolic Metabolism in Plants, Serie Recent Advances in Phytochemistry*, Stafford, H.; Ibrahim, R., Vol.26, pp. 201-231, Plenum Press, ISBN 9780306442315, New York, USA
- Phillips, R.C. (1984). *The ecology of eelgrass meadows in the Pacific Northwest: A community profile*, FWS/OBS-84/24, U.S. Dept. of the Interior, Washington, USA
- Prange, J.A. & Dennison, W.C. (2000). Physiological responses of five seagrass species to trace metals. *Marine Pollution Bulletin*, Vol.41, No.7-12, (July 2000), pp. 327-336, ISSN 0025-326X
- Procaccini, G.; Buia, M.C.; Gambi, M.C.; Perez, M.; Pergent, G.; Pergent-Martini, C. & Romero, J. (2003). The seagrasses of the Western Mediterranean, In: *World Atlas of Seagrasses*, Green, E.P.; Short F.T., pp. 48-58, UNEP World Conservation Monitoring Centre, University of California Press, Berkeley, ISBN 0-520-24047-2, USA
- Quackenbush, R.C.; Bunn, D. & Lingren, W. (1986). HPLC determination of phenolic acids in the water-soluble extract of *Zostera marina* L. (eelgrass). *Aquatic Botany*, Vol. 24, No.1, (January 1986), pp. 83-89, ISSN 0304-3770
- Ragan, M.A. & Glombitza, K.W. (1986). Phlorotannins, brown algal polyphenols. In: *Progress in Phycological Research*, Round, F.E.; Chapman D.J., Vol.4, pp. 130-241, Elsevier Biomedical Press, ISBN 094873700X, 9780948737008, UK
- Rainbow, P.S. & Phillips, D.J.H. (1993). Cosmopolitan biomonitors of trace metals. *Marine Pollution Bulletin*, Vol.26, No.11, (November 1993), pp. 593-601, ISSN 0025-326X
- Ramade, F. (1992). *Précis d'écotoxicologie*, Masson, Collection d'écologie, ISBN 2-225-82578-5, France

- Reichert, W.L.; Myers, M.S.; Peck-Miller, K.; French, B.; Anulacion, B. F.; Collier, T. K.; Stein, J.E. & Varanasi, U. (1998). Molecular epizootiology of genotoxic events in marine fish: Linking contaminant exposure, DNA damage, and tissue-level alterations. *Mutation Research*, Vol.411, No.3, (November 1998) pp. 215-225, ISSN 0027-5107
- Rice-Evans, C.A.; Miller, N.J.; Bolwell, P.G.; Bramley, P.M. & Pridham, J.B. (1995). The relative antioxidant activities of plant-derived polyphenolic flavonoids. *Free Radical Research*, Vol.22, No.4, (April 1995), pp. 375-383, ISSN 1071-5762
- SAS Institute, Inc. (1996). *SAS User's Guide: Statistics*, Carey, ISBN 0-917382-01-3, USA
- Schimmel, J.P.; Cates, R.G.; Clausen, T.P. & Reichardt, P.B. (1996). Effects of balsam poplar (*Populus balsamifera*) tannins and low molecular weight phenolics on microbial activity in taiga floodplain soil: Implications for changes in N cycling during succession. *Canadian Journal of Botany*, Vol.74, No.1, (January 1996), pp. 84-90, ISSN 0008-4026
- Short, F.T. & Wyllie-Echeverria, S. (1996). Human-induced disturbance in seagrasses. *Environmental Conservation*, Vol.23, No.1, (January 1996), pp. 17-27, ISSN 0376-8929
- Stephensen, E.; Savarsson, J.; Sturve, J.; Ericson, G.; Adolfsson-Erici, M. & Forlin, L. (2000). Biochemical indicators of pollution exposure in shorthorn sculpin (*Myoxocephalus scorpius*), caught in four harbours on the southwest coast of Iceland. *Aquatic Toxicology*, Vol.48, No.4, (April 2000), pp. 431-442, ISSN 0166-445X
- Swain, T. & Hillis, W.E. (1959). *The phenolic constituents of Prunus domestica. I. The quantitative analysis of phenolic constituents*. *Journal of the Science of Food and Agriculture*, Vol.10, No.1, (1959), pp. 63-68, ISSN 0022-5142
- Swain, T. (1977). Secondary compounds as protective agents. *Annals Review of Plant Physiology*, Vol.28, (1977), pp. 479-501, ISSN 0066-4294
- Tett, P.; Gowen, R.; Mills, D.; Fernandes, T.; Gilpin, L.; Huxham, M.; Kennington, K.; Read, P.; Service, M.; Wilkinson, M. & Malcolm, S. (2007). Defining and detecting undesirable disturbance in the context of marine eutrophication. *Marine Pollution Bulletin*, Vol.55, No.1-6, (2007), pp. 282-297, ISSN 0025-326X
- Tomlinson, P.B. (1974). Vegetative morphology and meristem dependence: The formation of productivity in seagrasses. *Aquaculture*, Vol.4, (1974), pp. 107-130, ISSN 0044-8486
- Vergeer, L.H.T.; Aarts, T.L. & Groot, J.D. (1995). The wasting disease and the effect of abiotic factors (light intensity, temperature, salinity) and infection with *Labyrinthula zosterae* on the phenolic content of *Zostera marina* shoots. *Aquatic Botany*, Vol.52, No.1, (September 1995), pp. 35-44, ISSN 0304-3770
- Vangronsveld, J.; Mocquot, B.; Mench, M. & Clijsters, H. (1997). Biomarqueurs du stress oxydant chez les végétaux, In: *Biomarqueurs en écotoxicologie - Aspects fondamentaux*, Lagadic L.; Caquet T.; Amiard J.C.; Ramade F., pp. 165-181, Dunod, ISBN 2225830533, Paris, France
- Verges, A.; Becerro, M.A.; Alcoverro, T. & Romero, J. (2007). Experimental evidence of chemical deterrence against multiple herbivores in the seagrass *Posidonia oceanica*. *Marine Ecology Progress Series*, Vol.343, (August 2007), pp. 107-114, ISSN 0171-8630
- Ward, T.J. (1987). Temporal variation of metals in the seagrass *Posidonia australis* and its potential as a sentinel accumulator near a lead smelter. *Marine Biology*, Vol.95, No.2, (February 1987), pp. 315-321, ISSN 0025-3162

- White, C.S. (1986). Volatile and water-soluble inhibitors of nitrogen mineralization and nitrification in a ponderosa pine ecosystem. *Biology and Fertility of Soils*, Vol.2, (1986), pp. 97-104, ISSN 0178-2762
- Wyllie-Echeverria, S. & Ackerman J.D. (2003). The seagrasses of the Pacific Coast of North America, In: *World Atlas of Seagrasses*, Green, E.P.; Short F.T., pp. 199-206, UNEP World Conservation Monitoring Centre, University of California Press, Berkeley, ISBN 0-520-24047-2, USA
- Wyllie-Echeverria, S.; Talbot S.L. & Rearick J.R. (2010). Genetic structure and diversity of *Zostera marina* (eelgrass) in the San Juan Archipelago, Washington, USA. *Estuaries and Coasts*, Vol.33, No.4, (July 2010), pp. 811-827, ISSN 1559-2723
- Zapata, O. & Mc Millan, C. (1979). Phenolic acids in seagrasses. *Aquatic Botany*, Vol.7, (1979), pp. 307-317, ISSN 0304-3770
- Zar, J.H. (1999). *Biostatistical analysis*, Prentice-Hall International, ISBN 9780130815422, U.K

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