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Rickettsiosis as Threat for the Traveller

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1. Introduction

Over the past six decades, tourism has experienced continued expansion and diversification becoming one of the largest and fastest growing economic sectors in the world. Many new destinations have emerged alongside the traditional ones of Europe and North America. In the next years an increase of travelling is expected, and the number of related infections will also be higher (http://www.unwto.org/facts/menu.html). Rickettsioses are an important chapter in the field of travel medicine.

Rickettsiae are small gram-negative intracellular bacteria (belonging to the alpha-1 proteobacteria) mainly transmitted by arthropods (lice, fleas, ticks and other acari) with two genera: *Orientia* with a unique specie (*Orientia tsutsugamushi*) and *Rickettsia* with several species. The clinical pictures that they cause are named rickettsioses (Raoult, 2010a).

Rickettsioses have been a threat all along the History and nowadays they are an important cause of morbi-mortality in some areas of the world. To know the distribution of the different diseases caused by these bacteria and how the clinical pictures are recognized may be essential for a quick diagnoses and starting the correct treatment. Some of these infections can be also easily prevented with basic rules. Main rickettsioses with their distribution area are showed in the table 1.

In the 21st Century in most parts of the world hygienic conditions have improved and epidemic typhus is absent. To acquire this condition it is necessary to be in contact with body lice. Furthermore, if people have personal hygiene and change their clothing, body lice are removed. Nevertheless it is possible that if we travel for cooperation to catastrophic areas or other places with poverty, we may take body lice (refugees' camps) and may develop exanthematic typhus.

There are a lot of references of rickettsioses acquired by travellers and considered imported diseases (McDonald et al., 1988; Bottieau et al. 2006; Freedman et al., 2006; Askling et al. 2009; Chen & Wilson, 2009; Jensenius et al., 2009; Stokes & Walters, 2009).

Nowadays ticks cause most travel-associated rickettsioses. Ticks are considered to be one of the most important vectors of infectious diseases in the world, preceded only by mosquitoes. Therefore, tick-borne rickettsioses are endemic all over the world (Hechemy et al., 2006). The majority of travel-associated rickettsioses refer to Sub-Saharan Africa tourists who develop African tick-bite fever (ATBF), mainly transmitted by *Amblyomma hebraeum* (Figure 1). In addition to malaria, ATBF is an important cause of fever in people returning from the tropic (Field et al., 2010). Other reports describe Mediterranean spotted fever (MSF) acquired by tourists bitten by *Rhipicephalus* spp. ticks (Figure 2) when visiting Europe, being

more scarce references about other rickettsioses. Flea-borne rickettsioses and chigger-transmitted rickettsioses are less frequent in travellers and tourists, and some of them as murine typhus are associated with poor hygienic conditions. Most travel-acquired rickettsioses are related to outdoors leisure activities, like camping, trekking, hunting, safaris, etc.

It will be impossible to describe all rickettsioses in few pages. Since rickettsioses have very similar clinical pictures and they can be grouped in different syndromes, we will describe these syndromes emphasizing the typical features (i.e.: Presence of eschar or type of rash). Afterwards, distribution can be observed in the table 1. We will also write a specific paragraph for some infections (i.e.: Diseases caused by *Rickettsia akari* and *Orientia tsutsugamushi*).



Fig. 1. *Amblyomma hebraeum*, the principal vector of African-tick bite fever (ATBF) in southern Africa.



Fig. 2. Rhipicephalus spp., the main vector of Mediterranean spotted fever (MSF).

DISEASE	CAUSATIVE AGENT	VECTOR	DISTRIBUTION
Epidemic typhus	R. prowazekii	Body lice (Pediculus humanus corporis)	Peru, northern Africa, Senegal, Burundi, Rwanda, Russia, sporadic cases in USA associated with flying squirrels. Potentially, all over the world associated to poverty and dirt.
Murine typhus	R. typhi	Fleas (Xenopsylla cheopis and Ctenocephalides felis)	All over the world (more prevalent in tropical and subtropical areas)
Flea-borne spotted fever	R. felis	Cat fleas (Ctenocephalides felis)	All over the world

DISEASE	CAUSATIVE	VECTOR	DISTRIBUTION	
21021102	AGENT	VECTOR	DISTRIBUTION	
Scrub typhus	Orientia tsutsugamushi	Trombiculid mite larvae (chiggers)	gers) Pakistan, Kashmir, Sri- Lanka, Afghanistan, Nepal, China, Japan, Korea, Indonesia.	
			Philippines, Papua-New Guinea, Australia	
Rickettsialpox	R. akari	Mouse mites Eastern Europe, Korea (Liponyssoides sanguineus) South Africa, USA		
RMSF ¹	R. rickettsii	Dermacentor variabilis and other American ticks	Dermacentor variabilis USA, Mexico, Colombia, and other American Brazil, Argentina,	
RMSF-like	R. parkeri	Amblyomma spp. ticks	USA, Uruguay, Brazil, Argentina	
MSF ²	R. conorii conorii, R. conorii israelensis, R. conorii caspia, R. conorii indica	Rhipicephalus spp. ticks	Mediterranean area, Central Europe, Russia, India and Africa	
MSF-like	R. monacensis	Ixodes ricinus ticks	Europe	
MSF-like	R. massiliae	Rhipicephalus sanguineus ticks	Mediterranean area, Argentina, USA?	
MSF-like	R. aeschlimannii	Hyalomma marginatum ticks	Africa, Europe?	
DEBONEL / TIBOLA ²	R. slovaca R. rioja R. raoultii	Dermacentor marginatus ticks	Europe	
LAR ⁴	R. sibirica mongolitimonae	Hyalomma spp. and Rhipicephalus pusillus ticks	Europe, Africa.	
ATBF ⁵	R. africae	Amblyomma spp. ticks		
Siberian tick typhus	R. sibirica sibirica	Dermacentor spp. ticks Russia, Pakistan, G		
R. helvetica infection	R. helvetica	Ixodes ricinus ticks	Central and northern Europe, Asia	
Japanese spotted fever	R. japonica	Ticks	Japan	
Queensland tick typhus	R. australis	Ixodes spp. ticks	Eastern Australia	
Flinder's Island spotted fever	R. honei	Ticks	Australia, Southeast Asia	
Far Eastern spotted fever	R. heilonjgiangensis	Dermacentor spp. ticks	Eastern Asia	

¹RMSF: Rocky Mountain spotted fever; ²MSF: Mediterranean spotted fever; ³DEBONEL/TIBOLA: *Dermacentor*-borne, necrosis, erythema, lymphadenopathy/Tick-borne lymphadenopathy; ⁴LAR: Lymphangitis-associated rickettsiosis; ⁵ATBF: African tick-bite fever.

Table 1. Rickettsia spp. causing medical diseases, vectors and distribution

2. Typhus syndrome

Typhus syndrome refers to a febrile syndrome with mental status impairment and rash. It is caused by *Rickettsia prowazekii* (epidemic typhus) and *Rickettsia typhi* (formerly, *R. mooseri*). *Rickettsia felis* may also produce a typhus syndrome named flea-borne spotted fever, which is similar to *R. typhi* infection (perhaps less severe) (Walker & Raoult, 2010; Dumler & Walker 2010; Oteo et al., 2006).

Nowadays epidemic typhus is only present in some regions of Africa, Russia and in Peru. It is associated with bad hygienic conditions that are necessary for body lice parasitization. Sporadic cases associated with contact with flying squirrels and their parasite arthropods, which have been involved as new reservoirs of the infection, have also been reported in USA. A possible source of *R. prowazekii* infection may be a recrudescent case (Brill-Zinsser disease) of *R. prowazekii* infection. If hygienic conditions are altered and an epidemic of body lice appears may be an epidemic of typhus, as occurred in Burundi with hundreds of affected people (Raoult et al., 1998). Some cases of louse-borne typhus in travellers have been published (Zanetti et al., 1998; Kelly et al., 2002).

Endemic typhus or murine typhus is associated with the presence of fleas. The main vector is the rat flea (*Xenopsylla cheopis*) associated with dirt and poor hygienic conditions. Fleaborne spotted fever is associated with the cat flea, and in this case bad hygienic conditions are not necessary. Murine typhus and flea-borne spotted fever are distributed all over the world. Although they are more frequent in tropical and subtropical areas, cases have also been reported in the Mediterranean area (Greece, Italy, Spain, France and Portugal) (Bernabeu-Wittel et al., 1999; Angel-Moreno et al., 2006; Gikas et al., 2009; Pérez-Arellano et al., 2005; Oteo et al., 2006).

The clinical pictures of murine typhus and flea-borne spotted fever are less severe than the one of epidemic typhus. Thus, 1-2 weeks after the flea exposure, patients begin with fever, headache, myalgia, nausea and vomiting. Rash can be difficult to see in some cases, but is present until 80%. For *R. typhi* infection, rash is macular or maculo-papular and typically affects trunk and less frequently extremities. In epidemic typhus, petechial rash is more frequent than in endemic typhus, and cough, nausea and vomiting are frequent features. On the contrary of tick-borne rickettsioses or scrub typhus, an inoculation eschar (*tache noire*) is not observed. In most cases, fever and rash disappear in a few weeks but complications can be developed (central nervous, kidney involvement with renal insufficiency, respiratory failure, etc.). These complications are more frequent for epidemic typhus and in older people or patients suffering chronic diseases (Walker & Raoult, 2010; Dumler & Walker 2010). In all these conditions a raise in hepatic enzymes, C reactive protein as well as in leucocytes and platelets counts can be observed. We can also observe hepatosplenomegaly. In severe cases mainly associated with epidemic typhus, evolution to a multiple-organ dysfunction syndrome and coagulation disorders may appear.

Some references related to travellers are: Zanetti et al., 1998; Niang et al., 1999; Kelly et al., 2002; Jensenius et al., 2004; Azuma et al., 2006; Angelakis et al., 2010; Walter et al., 2011.

3. Tick-borne spotted fever

Tick-borne spotted fever are worldwide distributed and the clinical picture is very similar, although the severity is different related with the *Rickettsia* species involved.

ATBF and MSF are the most frequent tick-borne spotted fever rickettsioses in travellers (Smoak et al., 1996; Fournier et al., 1998; Oteo et al., 2004a; Raoult et al., 2001; Caruso et al., 2002; Jensenius et al., 2003; Roch et al., 2008; Tsai et al., 2008; Consigny et al., 2009; Stephany et al., 2009; Althaus et al., 2010; Jensenius et al., 2004; Boillat et al., 2008; Laurent et al., 2009). For this reason, we will refer to these conditions taking into account that few differences in the incubation period and severity may exist. For Rocky Mountain spotted fever (RMSF) and MSF caused by *R. conorii israelensis*, higher mortality than with the rest of spotted fever rickettsioses has been communicated (de Sousa et al., 2003). Some features of the main spotted fever rickettsioses are shown in table 2.

From 4 to 21 days after the tick bite, fever suddenly starts in 100% cases (less severe in ATBF). A characteristic inoculation lesion (eschar) (figure 3) is typically found until 72% of MSF cases and until 95% for ATBF. Multiple eschars are observed in some cases. This is more frequent in ATBF. Fever is accompanied of chills, headache, etc. (table 2). From 3 to 5 days after the onset of fever, the rash appears. This is a maculo-papular rash with purpuric elements in some cases (figure 4). It is more frequent in extremities and typically affects palms and soles. In ATBF the rash can be vesicular (figure 5), as occurs in *R. akari* and *R. australis* infections. For *R. sibirica mongolitimonae* infection, known as lymphangitis-associated rickettsiosis (LAR), lymphangitis from the eschar may appear in approximately



Fig. 3. Eschar (*tache noire*) and maculo-papular rash in a patient with Mediterranean spotted fever.

50% cases. It also can be observed in ATBF (Figure 6). There are few reported cases of tick-borne spotted fever caused by other of *Rickettsia* species (*R. monacensis, R. aeschlimannii, R. massiliae, R. helvetica, R. sibirica mongolitimonae, R. parkeri, R. japonica* and *R. honei*, among others) but it seems that the clinical pictures are very similar to the one of MSF cases. Data about the incidence of these infections among travellers to endemic areas are very scarce (Jensenius et al., 2004; Socolovschi et al., 2010).

For *R. helvetica* infections rash can be absent and fever is often the unique clinical manifestation. All these diseases are more frequent in spring and summer, when the vectors are more active. In all these conditions a raise in hepatic enzymes, C reactive protein and in leucocytes and platelets counts can be observed. We can also observe hepatosplenomegaly. In severe cases mainly associated to RMSF or MSF, evolution to a multiple-organ dysfunction syndrome and coagulation disorders may appear.

Distribution of human cases of tick-borne rickettsioses in Europe, Africa and Americas are showed in figures 7-10. Human cases of tick-borne rickettsioses and scrub typhus in Asia and Oceania are showed in figure 11.

DISEASE	RASH	SPECIFITIES	ESCHAR	FEVER
MSF ¹	>95%	10% purpuric rash	72%. Multiple in 32%	100%
			(children)	
RMSF ²	90%	45% purpuric rash	<1%	100%
ATBF ³	30%	Vesicular rash	100%. Frequently multiple	100%
DEBONEL/TIBOLA4	Possible	Lymph nodes	100%. Larger than in other rickettsioses	30%
LAR ⁵	>90%	50% lymphangitis	Frequent	100%
R. aeschlimannii infection	Possible	-	Possible	100%
	Possible	-	Absent	Not
Tu never in incertain	1 0001212		Tibbeth	always
R. massilliae infection	Possible	Rash can be purpuric	Possible	100%
R. monacensis infection	Possible	Ī ,	?	100%
Queensland tick typhus	100%	Vesicular rash	65%	100%
Flinder's Island spotted fever	85%	8% purpuric rash	28%	100%
Siberian tick typhus	100%	-	77%	100%
Japanese spotted fever	100%	-	90%	100%
Far eastern spotted fever	Possible	-	Possible	100%

¹MSF: Mediterranean spotted fever; ²RMSF: Rocky Mountain spotted fever; ³ATBF: African tick-bite fever; ⁴DEBONEL/TIBOLA: *Dermacentor*-borne, necrosis, erythema, lymphadenopathy/Tick-borne lymphadenopathy; ⁵LAR: Lymphangitis-associated rickettsiosis.

Table 2. Main clinical characteristics of tick-borne rickettsioses



Fig. 4. Vasculitic rash affecting soles in a patient with Mediterranean spotted fever.



Fig. 5. Vesicular rash in a patient with African-tick bite fever.



Fig. 6. Eschar and lymphangitis in a patient with African tick-bite fever.

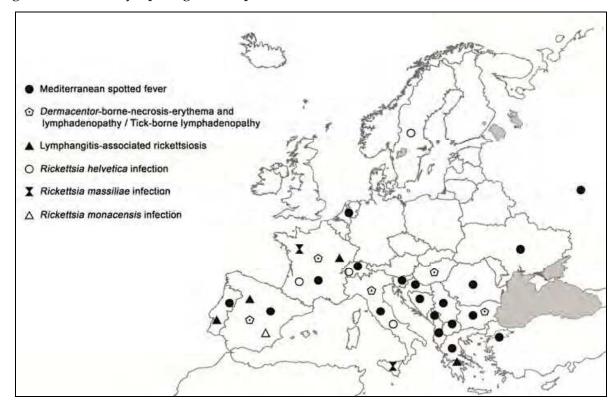


Fig. 7. Map showing distribution of human cases of tick-borne rickettsioses in Europe.

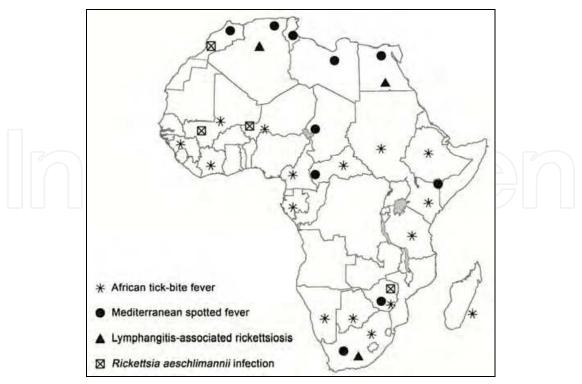


Fig. 8. Map showing distribution of human cases of tick-borne rickettsioses in Africa.



Fig. 9. Map showing distribution of human cases of tick-borne rickettsioses in Latin America.



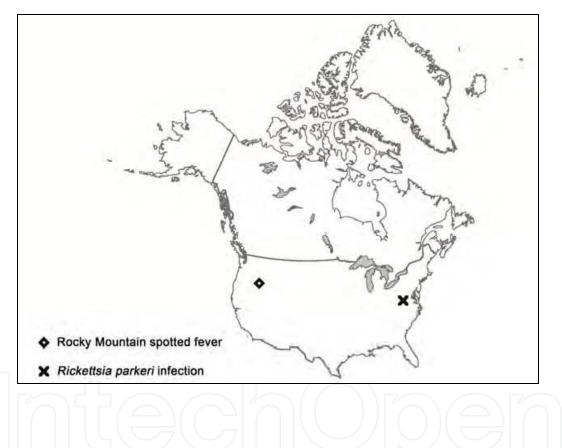


Fig. 10. Map showing distribution of human cases of tick-borne rickettsioses in North America.

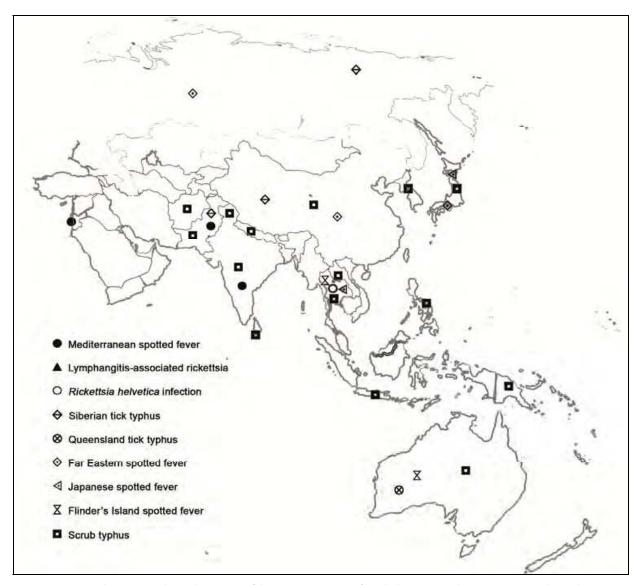


Fig. 11. Map showing distribution of human cases of tick-borne rickettsioses and srub typhus in Asia and Oceania.

4. Eschar and lymphadenopathy

This clinical syndrome has been recently reported in Europe, where it is named TIBOLA (TIck-BOrne LimphAdenopaty) or DEBONEL (*DErmacentor*-BOrne Necrosis-Erythema-Lymphadenopathy). *R. slovaca, R. rioja* and *R. raoultii* are the etiological agents, and *Dermacentor marginatus* is the main vector. This tick species is distributed all over Europe as well as in the North of Africa. Since this rickettsiosis appears in the coldest months of the year, the risk of acquisition for the travellers is lower than for the rickettsioses that are prevalent in spring and summer. In most cases (>90%) the tick-bite is located on the scalp (head) and always in the upper site of the body. After 1-15 days (mean: 4.8 days) of incubation period, the characteristic skin lesion starts as a crusted lesion at the site of the tick-bite (frequently on the scalp). A honey-like discharge from the lesion is observed in some cases. Few days later, a necrotic eschar appears (figure 12). This eschar is usually bigger than the one observed in MSF cases, and it is surrounded by an erythema. When the

tick-bite is out of the head, the skin lesion resembles the erythema migrans of Lyme disease. Other typical manifestation, which is always present when the bite is on the head, is the presence of regional and very painful lymphadenopathies.

On the contrary of other rickettsioses, in DEBONEL/TIBOLA there are not systemic clinical signs (or they are rare), such as fever or maculo-papular rash (Oteo et al., 2004b). The clinical course is sub-acute and no severe complications have been described.



Fig. 12. DEBONEL/TIBOLA patient with the typical crusted lesion on the scalp.

5. Scrub typhus

The etiological agent of scrub typhus is *Orientia tsutsugamushi*, which is transmitted by chigger bites (trombiculid mite larvae). It is mainly distributed in Afghanistan, India, Pakistan, Sri-Lanka, Kashmir, China, Nepal, Japan, Korea, Vietnam, Indonesia, Laos, Philippines, Papua New Guinea and Australia (Figure 11). Cases are mainly observed in autumn and spring, in temperate zones where the bite of this arthropod, which is on vegetation, is frequent. The incubation period is about 10 or more days and the clinical signs and symptoms are similar to typhus syndrome, including the rash which is transient and easily missed. A difference with typhus syndrome is the presence of eschar that is frequently multiple. The presence of regional lymphadenopathy is also more frequent. The mortality can be high despite the correct antimicrobial treatment. Outbreaks related to military operations have been reported (Pages et al., 2010). Most travel acquired cases of scrub typhus occur in patients returning from Southeast Asia (Jensenius et al., 2004, 2006).

6. Rickettsialpox

Rickettsialpox is a worldwide (North America, Eastern Europe, Korea and southern Africa) rickettsiosis caused by *Rickettsia akari* and transmitted by the bite of the mouse mite *Lyponyssoides sanguineus*. We can consider it a remerging infection since several cases have been detected in New York City after September 11 attacks (Paddock et al., 2003). Patients have fever, a prominent eschar -which is the best sign of the disease- and rash that, as occurs in ATBF and Queensland tick typhus, may be vesicular. Palms and soles are not involved. The presence of regional lymphadenopathy is common. Patients recover without treatment in most cases (Raoult, 2010b).

7. Laboratory diagnostic tools

As occurs for all infectious diseases, the most definitive diagnostic method is the rickettsial isolation in culture. The main problem is that *Rickettsia* spp. are strictly intracellular bacteria, conventional growth media cannot be used, and a laboratory with P3 safety level (not generally available in clinical microbiology labs) is necessary. Furthermore, culture is not very sensitive and the yield decreases when clinical samples are taken after antibiotic treatment or when samples are not processed within 24 hours. It is a slow technique that is used for research purposes but not for the routine clinical practice. Centrifugation shell-vial technique is a commercially available adaptation of cell cultures that is easier to handle, faster and less hazardous. Isolation attempts on cell cultures may be performed using buffy coat or tissue samples (eschar biopsies when possible). If not processed within 24 h, samples must be frozen at -70°C or in liquid nitrogen.

Detection of rickettsiae by Giménez or Giemsa staining from blood and tissue samples would allow the confirmation of the diagnosis, but these techniques are non-specific and their sensitivity is very low.

In some laboratories molecular biology tools, such as polymerase chain reaction (PCR) and sequencing, are also available. PCR-based assays from anticoagulated blood, biopsies and arthropod tissue samples targeting *Rickettsia* spp. genes are quite sensitive and useful for a quick diagnosis of these infections. The evaluation of several primer sets for the molecular diagnosis of rickettsioses demonstrated that the performance of three sequential PCRs (nested or semi-nested ones) allowed the detection and identification of *Rickettsia* species in a high percentage of the samples with previous clinical diagnosis or microbiological confirmation (serological analysis) of rickettsiosis (Santibáñez et al., 2011). Blood and tissue samples should be stored at -20°C or lower if PCR-based diagnosis is delayed for more than 24 hours. The European guidelines for the diagnosis of tick-borne bacterial diseases contain useful information for clinicians and microbiologists (Brouqui et al., 2004).

Indirect diagnostic tests and specifically, immunofluorescence assays (IFA) are considered the standard tests. Besides, since most traveller patients are investigated after returning, IFA are the most available tools for diagnosis. Acute and convalescent sera (collected 4-6 weeks after the onset of the illness) should be taken. In many cases we cannot observe seroconvertion but a high titre of antibodies. Cross-reactions among *Rickettsia* spp. make very difficult to definitively identify the causative agent by means of IFA. This can only be achieved in reference centres in which different antigens and other serological assays, such as western-blot, are available. Serum samples can be preserved at -20°C or lower for several months without significant degradation of antibodies.

Ticks removed from patients can be used as tools for the diagnosis of tick-borne rickettsioses. The strategy includes the identification of the tick to the species level, and the detection or isolation of rickettsias (Table 3).

- 1. Identification of the ticks to the species level
- 2. Detection of bacteria in ticks with the use of staining tests (haemolymph for viable ticks; salivary glands if ticks were frozen), or PCR-based methods (using one-half of the tick, the other half being kept frozen). PCR may also be done using only ticks that stain positive.
- 3. Sequencing of the amplified PCR fragment and comparison with available sequences in sequence databases.
- 4. If there is 100% similarity between the tested sequence and the corresponding sequence of a known organism, the presumptive identification is confirmed
- 5. If the tested sequence appears to be different from all corresponding sequences available, the organism is probably a new strain and should be isolated and characterized from the stored frozen part of the tick

Table 3. Strategy for detecting and/or isolating rickettsias from ticks

Diagnostic scores with epidemiological, clinical and laboratory tests for some tick-borne rickettsioses (ATBF and MSF) have been proposed (Tables 4 and 5).

- a. Direct evidence of *R. africae* infection by culture and/or PCR or
- b. Clinical and epidemiological features highly suggestive of ATBF, such as multiple inoculation eschars and / or regional lymphadenitis and / or a vesicular rash and / or similar symptoms among other members of the same group of travellers coming back from an endemic area (sub-Saharan Africa or French West Indies) and

Positive serology against spotted fever group rickettsiae

c. Clinical and epidemiological features consistent with a spotted fever group rickettsiosis such as fever and/or any cutaneous rash and/or single inoculation eschar after travel to sub-Saharan Africa or French West Indies

Serology specific for a recent R. africae infection (seroconversion or presence of IgM \ddagger 1:32), with antibodies to R. africae greater than those to R. conorii by at least two dilutions, and/or a Western blot or cross-absorption showing antibodies specific for R. africae

Table 4. Diagnostic criteria for African-tick bite fever (ATBF). A patient is considered to have ATBF when criteria A, B or C are met

CRITERIA			
Epidemiological criteria			
Stay in endemic area	2		
Occurrence in May-October	2		
Contact (certain or possible) with dog ticks	2		
Clinical criteria			
Fever > 39°C	5		
Eschar	5		
Maculopapular or purpuric rash	5		
Two of the above criteria	3		
All three of the above criteria	5		
Non-specific laboratory findings			
Platelets < 150 G/L			
SGOT or SGPT > 50 U/L			
Bacteriological criteria			
Blood culture positive for <i>Rickettsia conorii</i>			
Detection of <i>Rickettsia conorii</i> in a skin biopsy			
Serological criteria			
Single serum and IgG > 1/128			
Single serum and IgG > 1/128 and IgM > 1/64			
Four-fold increase in two sera obtained within a 2-week interval			

SGOT, serum glutamate-oxaloacetate transaminase; SGPT, serum glutamate-pyruvate transaminase. a A positive diagnosis is made when the overall score is \geq 25.

Table 5. Diagnostic criteria for Mediterranean spotted fever caused by Rickettsia conorii

8. Prophylaxis

An important chapter in the field of rickettsioses is related to prophylaxis. Since the majority of rickettsioses associated to travels are transmitted by ticks, the main preventive measure is to avoid tick-bites. Measures to avoid chiggers' attacks are the same as the ones used against ticks. Only fleas can be more difficult to avoid when cats and other pets are abundant. If there is risk of getting lice, hygiene measures such as changing clothing (they live in the seams of clothing) may be sufficient.

How can we avoid tick-bites? There are some rules that can be useful to avoid arthropodbites:

- 1. You must not wear dark clothes to see the ticks and remove them before attaching. Curiously, dark clothes attract less arthropods than clear ones. But, in our opinion, to look for the arthropods and remove them as soon as possible is more effective.
- 2. For outdoor activities (grass areas or mountains) you do not have to exposure your body to ticks. Thus, it is very useful to wear clothing that covers the majority of your body. The trousers must be tucked in your shocks with boots. Long sleeves shirt must be tucked into trousers. You must also wear a cap (especially children).
- 3. Permethrin-based repellents can be used on clothing, although their effect is short in time and the application should be repeated every few hours.
- 4. A careful inspection of clothing and body looking for ticks after returning from outdoors activities in endemic areas as well as removing them correctly has been

effective for the prevention of Lyme disease. The tick needs at least 24-48 hours for the transmission of *Borrelia burgdorferi*. This measure can be less efficient for *Rickettsia* spp. because these microorganisms can be transmitted since the first hours. But, anyway, the removal of the tick has to be done.

5. The contact with parasitized pets and wild animals must be avoided.

There are two questions that physicians have linked up with tick-bites: How must I remove the tick? Must I take prophylactic drugs after a tick-bite?

The first question is easy to answer. The most useful method to remove an attached tick is using forceps. Smooth forceps (without teeth) must be introduced between the tick's head and the skin in a 90° angle and then pull (Oteo et al., 1996). Other traditional methods as using oil, burning or freezing must be forgotten.

The other question is the use of prophylactic drugs after arthropod bites. There are no studies to answer this question. The transmission of rickettsias may be very quick, so we cannot extrapolate the recommendations for Lyme disease. Anyway, when people have been bitten by several ticks in an endemic area for a determinate disease (i.e.: Kruger National Park in South Africa and ATBF) and if the patient is anxious, we can offer doxycycline. It has been demonstrated that 3 doses of 100 mg. every 12 hours is safety and sufficient as treatment for the majority of rickettsioses. We must be cautious with the sun to avoid photo-sensibility. Children can take doxycycline for a short period of time. It is only contraindicated for pregnant women and in this case we can use macrolides (i.e. azythromycin).

Vaccine approaches for prevention of rickettsial diseases have been developed since the past century, but currently no vaccine is available. Major surface protein antigens (OmpA and OmpB) of *R. rickettsii* and *R. conorii* are candidate vaccine antigens. Molecular biology techniques such as selection, cloning and expression of genes encoding *R. prowazekii* virulence-associated proteins, offer the opportunity to develop new rickettsial vaccines against typhus group rickettsiae. Further research is needed to develop effective vaccines without undesirable toxic reactions (Azad & Radulovic, 2003; Walker, 2009).

9. Treatment

The treatment of rickettsiosis should be initiated as soon as possible. Antibiotics are very effective and may avoid severe complications and death. In all cases if rickettsiosis is suspected, samples should be sent for laboratory confirmation. In DEBONEL/TIBOLA, in which the clinical signs and symptoms are less severe, recovery without antimicrobials occurs but the use of antibiotics shortens the clinical course and improves the clinical picture (Ibarra et al., 2005).

Doxycycline is the most useful drug in children and adults. Doxycycline can be administered in short course (100 mg. every 12 hours for one day) for the treatment of typhus and scrub typhus. In the case of MSF, 2 doses of 200 mg./12 hours are also very effective (in children, 5 mg./kg./12hours); although most physicians use 100 mg. every 12 hours for 3-7 days after fever disappears. The same can be recommended for ATBF. This antibiotic regimen could probably be followed in other tick-borne rickettsioses but there are not good evidences (clinical assays) to support a recommendation. In RMSF the administration of doxycycline for 7 days is recommended. Other drugs that can be prescribed when not using doxycycline (allergy or pregnancy) are chloramphenicol (50-75 mg./kg./day given in 4 doses for 7-10 days) and azythromycin (500 mg./day for 5 days). Doxycycline for 7 days is the treatment of choice for rickettsialpox. Although there is *in vitro*

susceptibility to quinolones, the use of these drugs has been associated with worse clinical course (Botelho-Nevers et al., 2011).

10. Conclusion

In conclusion, rickettsioses are a worldwide threat that must be suspected in travellers returning from endemic areas. Most cases are caused by tick-bites, although in some areas of the world old diseases as typhus are present, and the risk exists. Rickettsiosis must be suspected in all patients with fever, exanthema with or without rash. Starting treatment with doxycycline when possible may be essential to rapidly recover and avoid complications. ATBF along with malaria is the leading cause of fever after returning from Sub-Saharan Africa.

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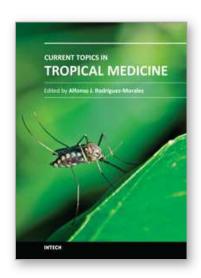
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