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Allelopathic Weed Suppression Through the Use of Cover Crops

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1. Introduction

There has long been observed an inhibitive response by plant species to certain neighboring plants. The Greek philosopher and botanist, Theophrastus, noted this effect from cabbage as early as 300 BC (Willis 1985). Since that time, others have documented similar plant interactions. In 1937, Austrian botanist, Hans Molisch, described this phenomenon as allelopathy, which he determined to be the result of biochemical interactions between plants (Molisch 1937; Putnam and Duke 1978). When first described, allelopathy referred to both deleterious and beneficial interactions between species; since that time, however, allelopathy has been applied to only adverse plant interactions, rather than to both. First described by a Roman scholar during the first century, black walnut (*Juglans nigra* L.) has long served as the common example of allelopathic effects with its ability to inhibit growth of surrounding plants either through decaying leaves or nuts or from the tree itself (Weir et al. 2004). Researchers have continued to examine allelopathy and the mechanism for biochemical inhibition, which was initially scrutinized by many since differentiation between this effect and plant competition remained uncertain (Weir et al. 2004). Subsequent bioassays involving specific chemical compounds extracted from plants have confirmed that certain species do, in fact, produce biochemicals that can inhibit plant germination and growth in the absence of resource competition (Einhellig 1994a).

With confirmation of allelopathy, many investigations have been conducted in order to determine how best to utilize this effect for possible weed control in agricultural settings (Khanh et al. 2005; Olofsdotter 2001; Weston 1996). The ability to inhibit weed growth through the implementation of cover crops into a crop rotation has been a focal point for this research for several reasons. In addition to weed suppression and control through allelopathy, as well as a mulching effect, cover crops provide substantial environmental benefits such as reduced erosion and water runoff (Price et al. 2006; Truman et al. 2003). Moreover, cover crops are readily available and easily adapted to many agricultural situations. Because of these many benefits, including natural weed suppression through allelopathy, the use of cover crops has become a vital component of sustainable agriculture systems, as well as organic production.

Ensuring sufficient food and fiber production for future generations can be hampered by limited options for weed control, particularly in developing countries where yields are reduced by up to 25% by weed competition. Identifying and describing sustainable weed control measures that can be implemented to reduce weed pressure in a number of settings can help safeguard the productivity of agriculture. Therefore, the objectives of this chapter are to describe the fundamentals of allelopathy and how to utilize allelopathic compounds for weed control through cover crop use. The chapter also highlights many of the identified biochemicals, their structures, and the respective cover crops in which they are found. Lastly, we describe the degree of allelopathic potential for a number of cover crops, as determined by laboratory testing.

2. Production and release of allelopathic compounds

Allelochemicals enter the environment from plants in a number of ways, such as plant degradation, volatilization, leaching from plant leaves, and from root exudation (Bertin et al. 2003; Weir et al. 2004). During active plant growth, particularly in early growth stages or during periods of stress, root exudation, either through diffusion, ion channels, or vesicle transport, is the primary method for release of many organic and inorganic compounds into the rhizosphere (Battey and Blackbourn 1993; Uren 2000). These compounds serve a multitude of functions such as improving nutrient uptake, root lubrication, plant growth regulation, microorganism defense, and waste removal (Bertin et al. 2003; Fan et al. 1997; Uren 2000).

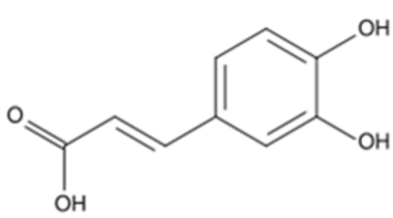
A large proportion of identified allelochemicals are noted to be secondary compounds formed during photosynthetic processes (Einhellig 1994b; Swain 1977). Since many allelopathic chemicals appear to perform no primary metabolic functions, although some compounds such as cinnamic acid and salicylic acid do serve other functions within a plant, it is unclear at this point as to what regulates the release of these compounds (Einhellig 1994a). Many environmental plant stressors have been observed to increase allelochemical release but not necessarily chemical production (Bertin et al. 2003; Inderjit and Weston 2003; Sterling et al. 1987). Plant stressors such as elevated temperature, reduced water availability, and herbivory may cause increased allelochemical release; however, a definitive correlation between environmental factors and allelopathic compounds has yet to be made (Bertin et al. 2003; Pramanik et al. 2000). Continued research directed at isolating and identifying individual root exudates while manipulating environmental stress factors may help to increase our understanding of allelochemical release into the rhizosphere.

3. Allelopathic compounds

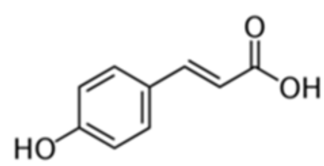
Many allelochemicals have been identified since experiments began to isolate and determine allelopathic potentials of plant compounds. Compounds that have been identified thus far include a variety of chemical classes such as phenolic acids, coumarins, benzoquinones, terpenoids, glucosinolates, and tannins (Chung et al. 2002; Putnam and Duke 1978; Seigler 1996; Swain 1977; Vyvyan 2002). These and other allelochemicals are found in many plant species from woody to herbaceous plants, grasses and broadleaves, weeds and crops. There are many details left to be determined such as regulation and production stimuli and mode of action for inhibition. It is also not readily understood to what extent allelopathic compounds interact with each other and other chemical compounds within the rhizosphere to inhibit surrounding plants. The following sections present several of the structural classes of recognized allelochemicals as well as specific compounds within each group.

3.1 Phenolic acids

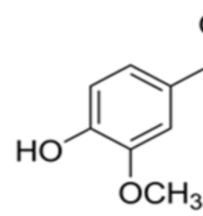
Like most allelochemicals, phenolic acids are secondary plant compounds typified by a hydroxylated aromatic ring structure. To date, a number of phenolic acids have been determined to have allelopathic properties and have been measured in extracts from a variety of plant species (Figure 1). Species which have been noted to produce phenolic acids include: rice (*Oryza sativa* L.), wheat (*Triticum aestivum* L.), mango (*Mangifera indica* L.), and spotted knapweed (*Centaurea stoebe* L.) (Bais et al. 2003; Chung et al. 2002; El-Rokiek et al. 2010; Fitter 2003). Many species, such as rice, contain multiple phenolic compounds along with other allelopathic compounds. In two studies, researchers isolated nine individual phenolic acids from rice hull extracts and 14 different phenolic acids from buffalograss [*Buchloe dactyloides* (Nutt.) Engelm] (Chung et al. 2002; Wu et al. 1998). At this time, however, it is not clear to what degree individual allelochemicals interact to produce plant inhibition. Some reports show a synergistic effect when allelochemicals are in a mixture, while other studies indicate decreased plant inhibition in the presence of a mixture when compared to individual chemical inhibition (Chung et al. 2002; Einhellig 1996).



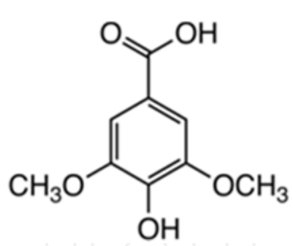
Caffeic acid



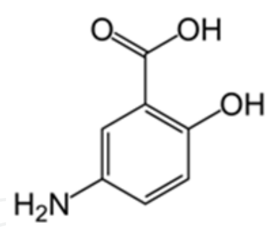
Coumaric acid



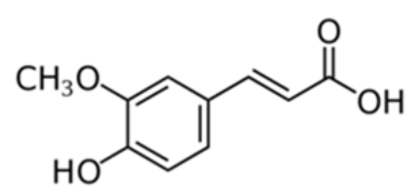
Vanillic acid



Syringic acid



p-hydrobenzoic



Ferulic acid

Fig. 1. Phenolic acids identified in many plant species, such as oat (*Avena sativa* L.) and rice (*Oryza sativa* L.), have been found to have allelopathic properties.

Although modes of action for allelopathic chemicals are not readily understood for each identified allelochemical, phenolic acids have been the focus of many studies designed to establish the basis of their allelopathy (Putnam 1985). Early research with phenolic acids indicated that some phenolic acids could function though increasing cell membrane permeability, thus affecting ion transport and metabolism (Glass and Dunlop 1974). More recent studies report disruption of cell division and malformed cellular structures in plants exposed to phenolic acids (Li et al. 2010). Reduced respiration and reduced photosynthetic rates, due to decreased photosynthetic products such as chlorophyll, have also been reported in the presence of phenolic acids (Patterson 1981; Yu et al. 2003). Other studies have cited altered plant enzymatic functions, inhibited protein synthesis, and inactivated plant hormones as inhibitory mechanisms from these allelochemicals (Batish et al. 2008; Li et al. 2010). Each mechanism of plant inhibition can lead to the reduced growth and/or death of an exposed plant; however, it is likely multiple functions within a plant are being affected simultaneously due to the mixture of allelochemicals released from a plant species. Despite the extensive research with phenolic acids, target sites for allelochemical activity within affected plant species remain to be determined for many phenolic compounds.

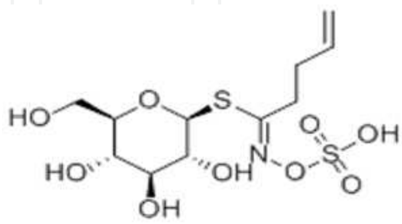
3.2 Glucosinolates

Glucosinolates occur in many plant species, but are widely known to be produced by species within the Brassicaceae family (Figure 2) (Haramoto and Gallandt 2005; Malik et al. 2008; Mithen 2001). Members of this family include: wild radish (*Raphanus raphanistrum* L.), white mustard (*Sinapis alba* L.), turnip (*Brassica campestris* L.), and rapeseed (*Brassica napus* L.). Glucosinolates, secondary metabolites containing sulfur and nitrogen, are enzymatically hydrolyzed by myrosinase in the presence of water to form isothiocynates, the active allelochemicals (Haramoto and Gallandt 2005; Norsworthy and Meehan 2005; Petersen et al. 2001; Price et al. 2005). Previous research examining extracts from glucosinolate-producing plant species have shown inhibition of other species through reduced germination, reduced seedling emergence and reduced size, as well as delayed seed germination (Al-Khatib et al. 1997; Brown and Morra 1996; Malik et al. 2008; Norsworthy et al. 2007; Wolf et al. 1984). Although specific modes of action have not been thoroughly investigated for each compound, it is evident that some plant species are able to tolerate these allelochemicals more readily than other species (Norsworthy and Meehan 2005). Some suggest that seed size variability plays a role in determining inhibitory effects of these allelochemicals; however, this may not be the only determinant for tolerance to these compounds (Haramoto and Gallandt 2005; Westoby et al. 1996). Future research with these allelopathic compounds will likely seek to answer this question, along with identifying the mode of action for plant inhibition, in order to utilize these compounds more effectively in agricultural production.

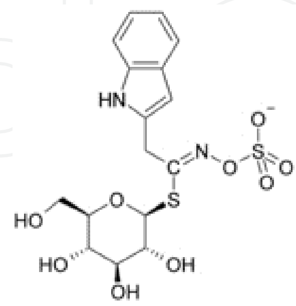
3.3 Coumarins

Coumarin compounds (Figure 3) are found in a range of plant species, particularly from the Apiaceae, Asteraceae and Fabaceae families (Razavi 2011). Coumarins and their derivatives have been identified in plants such as lettuce (*Lactuca sativa* L.), wild oat (*Avena sativa* L.), sweet vernalgrass (*Anthoxanthum odoratum* L.), and a number of other species (Abenavoli et al. 2004; Razavi 2011). Like many other allelochemicals, coumarins have been found to inhibit plant growth by reduced seedling germination and reduced root and shoot growth, likely with interference in photosynthesis, respiration, nutrient uptake and metabolism (Abenavoli et al. 2001; Abenavoli et al. 2004; Razavi et al 2010; Yamamoto 2008).

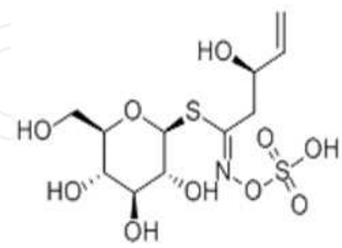
In addition to plant inhibition, biological activity of coumarins includes antibacterial, nematicidal, antifungal, and insecticidal activity; moreover, pharmacological activity of coumarins has been commonly noted in a number of instances with specific compounds functioning to reduce edema and inflammation (Casley-Smith and Casley-Smith 1992; Hoult and Paya 1996; Maddi et al. 1992; Razavi 2011). The broad activity of these compounds has made pharmaceutical use difficult due to the potential for non-target activity. Although allelopathic research has yet to indicate that the broad spectrum activity of coumarins could limit future use of these compounds for weed control, this may require further investigation as research moves forward.



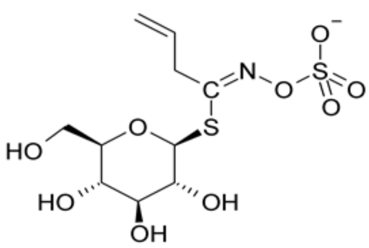
Gluconapin



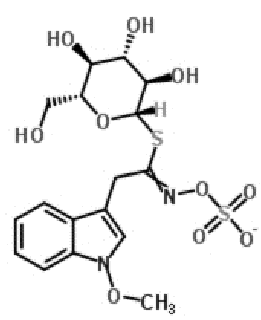
Glucobrassicin



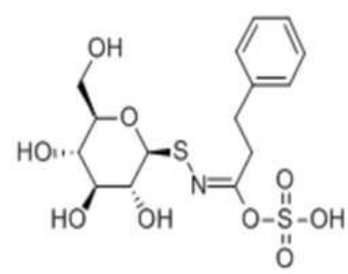
Progoitrin



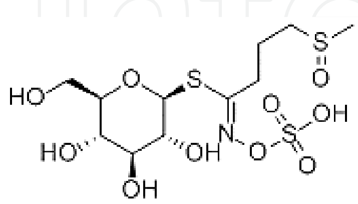
Sinigrin



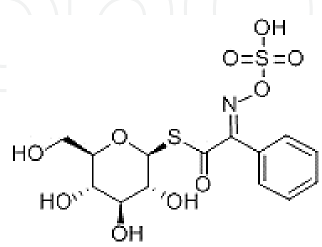
Neoglucobrassicin



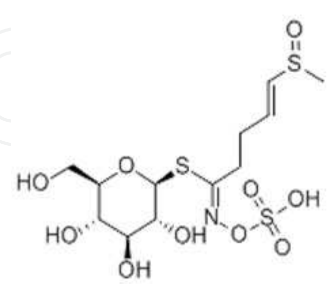
Gluconasturtin



Glucoiberin

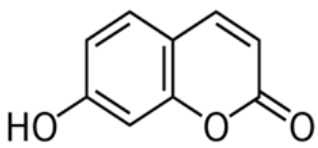


Glucotropaeolin

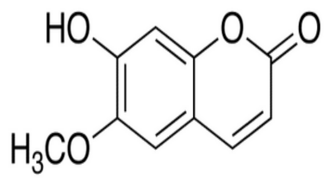


Glucoraphenin

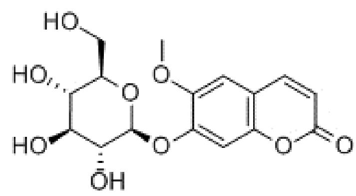
Fig. 2. Glucosinolates, allelopathic compounds known to be produced by plants in the Brassicaceae family as well as other families, are produced in both the root and shoot regions of plants.



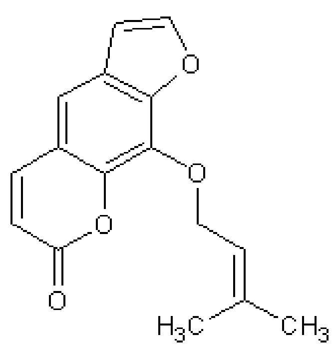
Umbelliferone



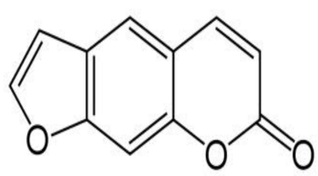
Scopoletin



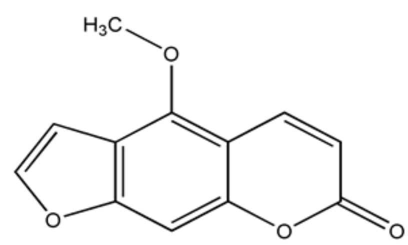
Scopolin



Imperatorin



Psoralen



Bergapten

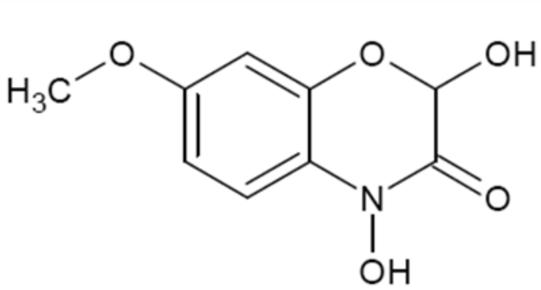
Fig. 3. Coumarins and their subgroups have been identified as allelopathic compounds in several plant families including Apiaceae and Fabaceae.

3.4 Other allelopathic compounds

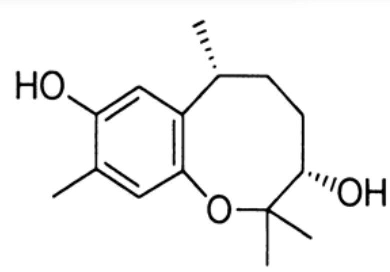
Many other allelochemicals have been detected in a wide range of species; however, a few compounds have been more widely researched. Classes of allelochemicals under thorough investigation, such as the benzoxazinoids, heliannuols, and benzoquinones, offer potential benefits for weed control in agricultural systems (Figure 4) (Macias et al. 2005; Vyvyan 2002). These classes, described briefly below, represent only a few of the many other compounds that may one day provide substantial weed suppression through allelopathy.

Benzoxazinoid compounds, identified in cereal grains such as wheat and rye, include DIBOA [2,4-dihydroxy-(2*H*)-1,4-benzoxazin-3(4*H*)-one] and DIMBOA [2,3-dihydroxy-7-methoxy-(2*H*)-1,4-benzoxazin-3(4*H*)-one] (Burgos and Talbert 2000; Macias et al. 2005). These compounds are easily degraded into other allelopathic forms, BOA (2-benzoxazolinone) and MBOA (7-methoxy-2-benzoxazolinone), within the soil and can diminish plant germination and growth (Barnes et al. 1987; Burgos and Talbert 2000). In light of the allelopathic properties of BOA and MBOA, it is now recognized that continued research efforts are needed to understand the role of breakdown products of initial allelochemicals in inhibiting plant growth (Macias et al. 2005).

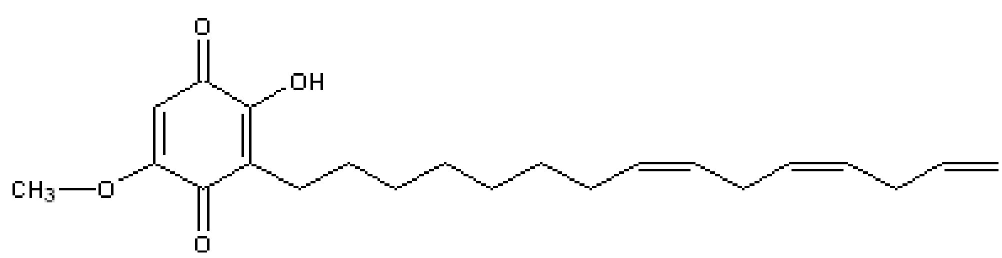
From the sunflower plant (*Helianthus annuus* L.), several compounds have been identified as being allelopathic (Leather 1983; Vyvyan 2002). The heliannuols are classified as phenolic sesquiterpenes and are noted for allelopathic as well as pharmacological activity (Vyvyan 2002). In addition to having been isolated from the sunflower, similarly structured compounds have been detected in animal species as well (Harrison and Crews 1997). Most notable about heliannuolic compounds is their ability to suppress plant growth at relatively low concentrations. Although they have been shown to inhibit growth of many broadleaf weed species, heliannuols appear to have a stimulating effect upon monocotyleden species (Weidenhamer 1996; Vyvyan 2002). This aspect of heliannuol activity may prove difficult when developing weed control applications of these compounds.



DIMBOA



Heliannuol A



Sorgoleone

Fig. 4. Compounds, such as DIMBOA, heliannuol A, and sorgoleone, continue to be studied for their allelopathic properties.

Benzoquinone compounds, primarily sorgoleone, isolated from sorghum [*Sorghum bicolor* (L.) Moench], have also been determined to be highly allelopathic (Netzly et al. 1998). Research with this compound indicates plant growth inhibition is achieved through disruption of photosynthesis as well as reduced chlorophyll development (Einhellig and Souza 1992). Like some other compounds, sorgoleone exhibits selective activity with inhibition of many germinating seedlings but little activity against certain species such as morningglory (*Ipomoea spp*.) (Nimbal et al. 1996). Research conducted with sorghum root exudates compares sorgoleone activity to that of the herbicide, diuron, but has many target sites (Nimbal et al. 1996; Rimando et al. 1998). Thus far, characteristics of sorgoleone show that it is a promising compound for development into a natural herbicide as an alternative to synthetic herbicides.

4. Weed control through allelopathy

Ongoing research into allelopathy seeks to better understand the mechanisms of allelopathy in order to make use of these naturally occurring weed suppressants within agricultural areas. Benefits offered by employing allelopathy as some form of weed control could aid in developing more sustainable agricultural systems for future generations (Einhellig 1994a). Current efforts focus primarily on natural herbicide production and cover crops. Although these concepts are being utilized to some degree, there remains a great deal of research to fully utilize the potential of allelopathy.

The role of naturally derived compounds, or synthetically produced mimics, for use as pesticides has been widely adopted, particularly for insect control. Several plant derived compounds, such as pyrethrum, neem, and nicotine, are important chemicals for insect control in many areas (Isman 2006). Herbicide potentials of isolated plant extracts have been indicated by a number of researchers but to date, few have been marketed. Synthetic compounds, such as cinmethylin, and mesotrione, were developed based upon plant-derived allelochemicals, but release of subsequent plant-based herbicides has lagged (Lee et al. 1997; Macias et al. 2004; Secor 1994; Vyvyan 2002). Slow production and release of herbicides developed in this manner are most likely attributed to limited understanding of the modes of action for many identified allelochemicals. To date, a number of allelochemicals have been isolated and investigated to develop natural herbicides with these compounds. Understanding the mode of action for plant inhibition may aid in the development of new products for the market.

A great deal of research has been devoted to the use of cover crops for weed control. Until recently, however, the allelopathic potential of cover crops has received less attention due, in part, to the lack of knowledge about allelopathy in general. As the functions of allelopathic compounds are beginning to be understood, more focus is being given to the allelochemicals within cover crops. In agricultural settings, cover crops have been in use for a number of years as a ground cover to slow erosion and water runoff as well as to impede germination of weed seed by providing a physical barrier (Kaspar et al. 2001; Price et al. 2008; Sarrantonio and Gallandt 2003). The growing need for sustainable agricultural systems has necessitated increased cover crop research to better utilize these covers for effective weed control. As a result, recent investigations have sought to understand the role of allelopathy for weed suppression within various cover crops (Burgos and Talbert 2000; Khanh et al. 2005; Price et al. 2008; Walters and Young 2008).

5. Allelopathic potential of cover crops

Determining allelopathic potential of exudates of plant species can be difficult and time consuming to complete. Bioassays are generally conducted to identify allelopathic properties of compounds in order to differentiate between allelopathy and mulching effects. Our research has focused on determining the extent of allelopathic effects of available cover crops on weed species as well as crop species. Extract-agar bioassays conducted with radish

(*Raphanus sativus* L.), an indicator species, and cotton (*Gossypium hirsutum* L.) established levels of inhibition for radicle elongation by extracts from cover crops, primarily legumes and cereal grains.

Legume cover crops have the ability to fix atmospheric nitrogen that potentially provides a nitrogen source to the subsequent crop without the need for additional fertilizer applications (Balkcom et al. 2007; Hartwig and Ammon 2002). Legume species such as vetch (*Vicia villosa* Roth), clover (*Trifolium spp.*), black medic (*Medicago lupulina* L.), and winter pea (*Pisum sativum* L.) are typically used as cover crops in agricultural production in the United States (Figure 5) (SARE 2007). Other legume crops beginning to be researched as possible choices for cover crops are sunn hemp (*Crotalaria juncea* L.) and white lupin (*Lupinus albus* L.); however, their availability and use are not as widespread as the previously mentioned legumes. In addition to being a nitrogen source for primary crops, legume covers provide a weed control potential. Due to the rapid degradation of legume residue on the soil surface in comparison to cereal grain residue, weed control through a physical barrier may not last as long into the season as other cover crops.

Fig. 5. Legume cover crops, such as white lupin (in mixture with black oats), provide weed suppression and nitrogen benefits to the subsequent cash crop.

Determining allelopathic effects of legume cover crop extracts concluded that legume covers did inhibit radish and cotton radicle elongation; however, cotton root exhibited less inhibition than that of radish for all included crops (Price et al. 2008) (Figure 6). In our research, hairy vetch had the greatest inhibition while winter pea had the least effect on germinating seedlings. It is important to note that different varieties of cover crops are available for use in agricultural systems and the varieties of one species may differ in level of allelopathy. Although under field conditions, allelopathic performance of these species may fluctuate, it is apparent that these cover crops can provide additional weed control measures over systems that do not include a cover crop.

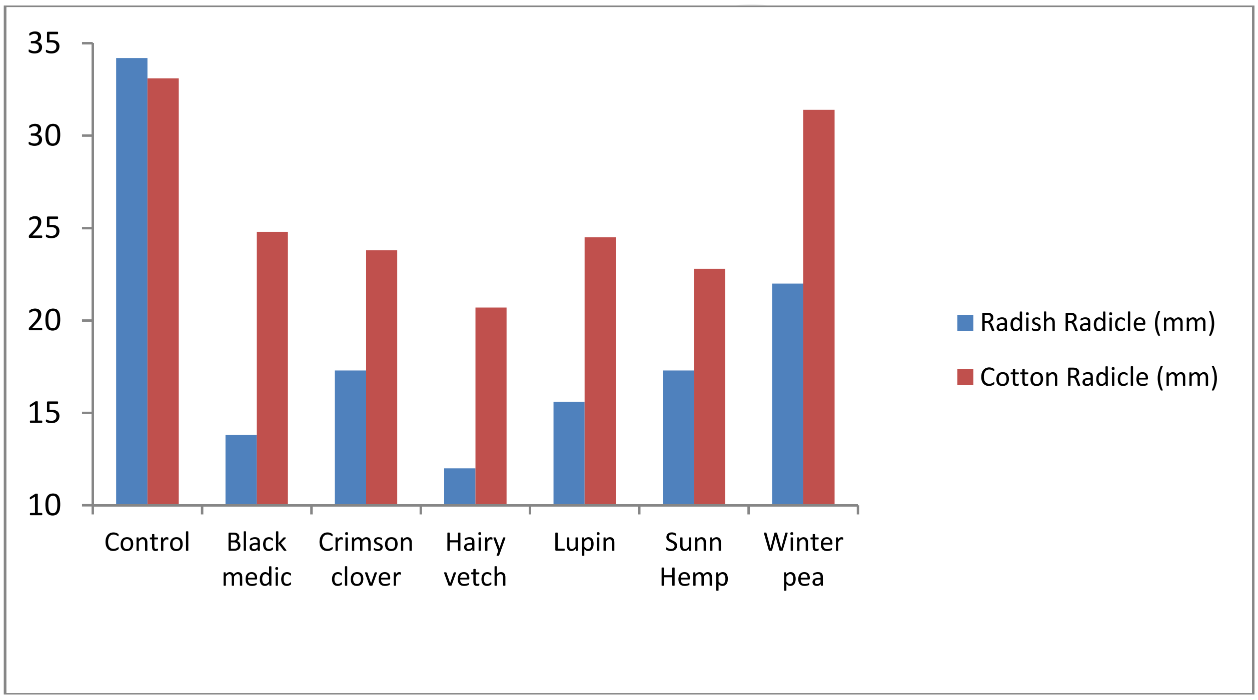


Fig. 6. Legume cover crops affect radicle elongation of different plant species to varying degrees.

Cereal grain crops such as black oat (*Avena strigosa* Schreb), rye, triticale (*X Triticosecale* Wittmack), and wheat, are utilized frequently in conservation systems as cover crops with effective ground cover and weed suppression (Figure 7). Rye is a commonly used cereal cover crop due to its ability to be sown later in the season while maintaining successful growth and its biomass production capability. With increased biomass on the soil surface, weed suppression will be increased as well. Cereal crops will also decay more slowly than more herbaceous plant species and provide some ground cover, and allelochemical release, further into the growing season. Additionally, rye has been noted to be less affected by plant diseases than other cover crops, and aids in reducing insect pests within a system (Wingard 1996).

Like legumes, cereal grain crop exudates in our study were able to significantly inhibit radicle elongation compared to the control (Figure 8). The disparity between radish and cotton radicle inhibition for each cover crop studied suggests that minimized interference with primary crops and increased weed suppression potential could be achieved with the use of cereal grain crops. These allelopathic effects, however, may be amplified or diminished depending on the field environment, plant stress levels, cover crop variety, and a number of other factors involved in determining allelochemical levels. Nevertheless, this research provides a base of allelopathic concentrations and impacts from various cover crops and may be an initial consideration when choosing a cover crop for inclusion in a system.

Fig. 7. Cotton growing in rolled black oat residue. Cereal grain cover crops, like black oat and rye, can be utilized to achieve a large quantity of plant residue on the soil surface.

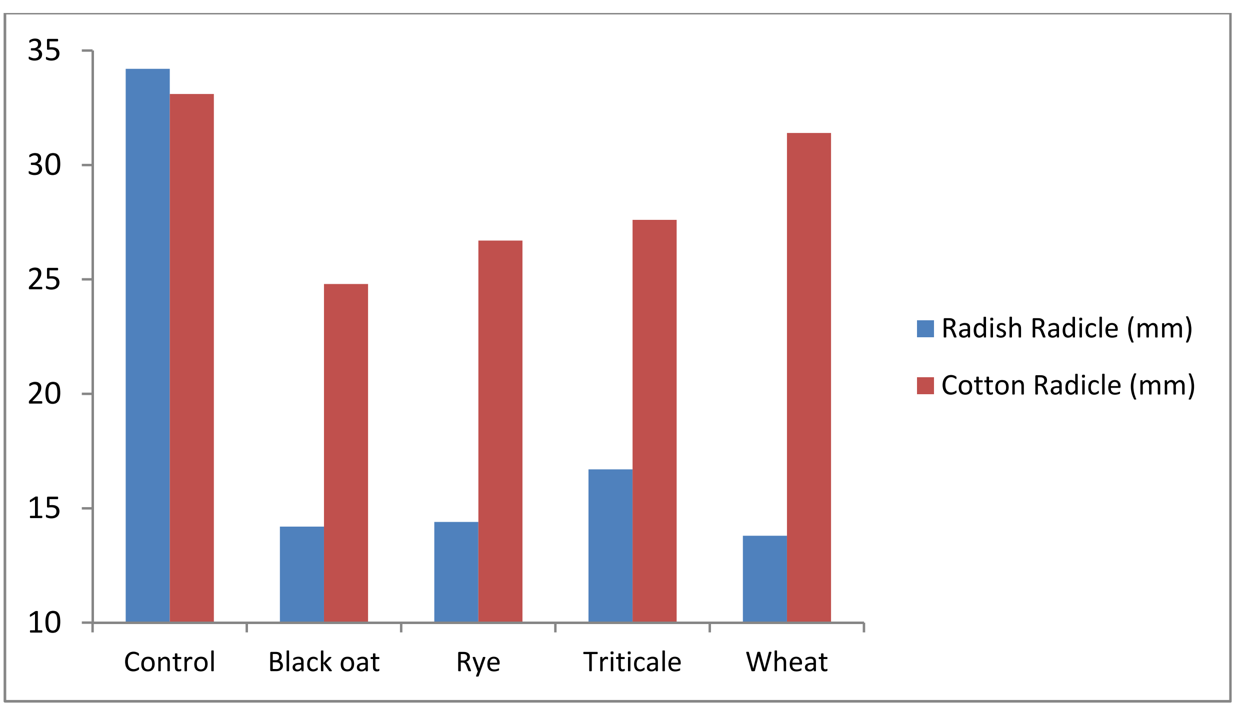


Fig. 8. Radish and cotton radicle elongation is reduced by cereal grain cover crops.

6. Conclusions

The growing demand for sustainable agricultural systems requires that researchers reevaluate current production methods and inputs. To ensure continued productivity and potentially reduce synthetic herbicide requirements, allelopathy has become a focal point for research in the agricultural community. Although, many questions have yet to be resolved, the utilization of allelochemicals for weed suppression remains a promising avenue for reducing herbicide usage. Whether through the development of natural herbicides from isolated allelochemicals or through the application of cover crops with allelopathic properties, allelopathy will most likely be a factor in providing sustainable systems in the future.

7. References

Abenavoli, M.R., C. De Santis, M. Sidari, A. Sorgona, M. Badiani, nad G. Cacco. 2001. Influence of coumarin on the net nitrate uptake in durum wheat (*Triticum durum* Desf. Cv. Simeto). *New Phytologist* 150: 619-627.

Abenavoli, M.R., A. Sorgona, S. Albano, and G. Cacco. 2004. Coumarin differentially affects the morphology of different root types of maize seedlings. *Journal of Chemical Ecology* 30: 1871-1883.

Al-Khatib, K., C. Libbey, and R. Boydston. 1997. Weed suppression with *Brassica* green manure crops in green pea. *Weed Science* 45: 439-445.

Bais, H.P., R. Vepachedu, S. Gilroy, R.M. Callaway, and J.M. Vivanco. 2003. Allelopathy and exotic plant invasion: From molecules and genes to species interactions. *Science* 301: 1377-1380.

Balkcom, K.S., H. Schomberg, W. Reeves, and A. Clark. 2007. Managing cover crops in conservation tillage systems. In *Managing Cover Crops Profitably,* 3rd Edition, ed. A. Clark, 44-65. College Park, MD: Sustainable Agriculture Research and Education.

Barnes, J.P., A.R. Putnam, B.A. Burke, and A.J. Aasen. 1987. Isolation and characterization of allelochemicals in rye herbage. *Phytochemistry* 26: 1385-1390.

Batish, D.R., H.P. Singh, S. Kaur, R.K. Kohli, and S.S. Yadav. 2008. Caffeic acid affects early growth and morphogenetic response of hypocotyl cuttings of mung bean (*Phaseolus aureus*). *Journal of Plant Physiology* 165: 297-305.

Battey, N.H., and H.D. Blackbourn. 1993. The control of exocitosis in plant cells. *New Phytology* 125: 307-308.

Bertin, C., X. Yang, and L.A. Weston. 2003. The role of root exudates and allelochemicals in the rhizosphere. *Plant and Soil* 256: 67-83.

Brown, P.D. and M.J. Morra. 1996. Hydrolysis products of glucosinolates in *Brassica napus* tissues as inhibitors of seed germination. *Plant and Soil* 181: 307-316.

Burgos, N.R. and R.E. Talbert. 2000. Differential activity of allelochemicals from *Secale cereale* in seedling bioassays. *Weed Science* 48: 302-310.

Casley-Smith, J.R. and J.R. Casley-Smith. 1992. Modern treatment of lymphedema II. The benzopyrones. *Australian Journal of Dermatology* 33: 69-74.

Chung, I.M., K.H. Kim, J.K. Ahn, S.C. Chun, C.S. Kim, J.T. Kim and S.H. Kim. 2002. Screening of allelochemicals on barnyardgrass (*Echinochloa crus-galli*) and identification of potentially allelopathic compounds from rice (*Oryza sativa*) variety hull extracts. *Crop Protection* 21: 913-920.

Einhellig, F.A. and I.F. Souza. 1992. Phytotoxicity of sorgoleone found in grain sorghum root exudates. *Journal of Chemical Ecology* 18: 1-11.

Einhellig, F.A. 1994a. Allelopathy: Current status and future goals. In *Allelopathy,* eds. D. Inderjit, K.M.M. Dakshini and F.A. Einhellig, 1-24. Washington, D.C.: American Chemistry Society.

Einhellig, F.A. 1994b. Mechanisms of action of allelochemicals in allelopathy. In *Allelopathy: Organisms, Processes, and Applications*, eds D. Inderjit, K.M.M. Dakshini and F.A. Einhellig, 96. Washington, D.C.: American Chemistry Society.

Einhellig, F.A. 1996. Interactions involving allelopathy in cropping systems. *Agronomy Journal* 88: 886-893.

El-Rokiek, K.G., R. Rafat, N.K. El-Masry, and S.A. Ahmed. 2010. The allelopathic effect of mango leaves on the growth and propagative capacity of purple nutsedge (*Cyperus rotundus* L.). *Journal of American Science* 6: 151-159.

Fan, T.W.M., A.M. Lane, D. Crowley, and R.M. Higashi. 1997.Comprehensive analysis of organic ligands in whole root exudate using nuclear magnetic resonance and gas chromatography-mass spectrometry. *Analytical Biochemistry* 257: 57.

Fitter, A. 2003. Making allelopathy respectable. *Science* 301: 1337-1338.

Glass, A.D.M. and J. Dunlop. 1974. Influence of phenolic acids on ion uptake. *Plant Physiology* 54: 855-858.

Haramoto, E.R. and E.R. Gallandt. 2005. Brassica cover cropping: 1. Effects on weed and crop establishment. *Weed Science* 53: 695-701.

Hartwig, N.L. and H.U. Ammon. 2002. Cover crops and living mulches. *Weed Science* 50:688-699.

Harrison, B. and P. Crews. 1997. The structure and probable biogenesis of helianane, a hetercyclic sesquiterpene, from the Indo-Pacific sponge *Haliclona fascigera. Journal of Organic Chemistry* 62: 2646-2648.

Hoult, J.R.S. and M. Paya. 1996. Pharmacological and biochemical actions of simple coumarins: Natural products with therapeutic potential. *General Pharmacology* 27: 713-722.

Inderjit, D. and L.A. Weston. 2003. Root exudation: an overview. In *Root Ecology*, eds. DeKroon and E.J.W. Visser. Heidelberg, Germany: Springer-Verlag.

Isman, M.B. 2006. Botanical insecticides, deterrents, and repellants in modern agriculture and an increasingly regulated world. *Annual Review of Entomology* 51: 45-66.

Kaspar, T.C., J.K. Radke, and J.M. Laflen. 2001. Small grain cover crops and wheel traffic effects on infiltration, runoff, and erosion. *Journal of Soil and Water Conservation* 56: 160-164.

Khanh, T.D., M.I. Chung, T.D. Xuan, and S. Tawata. 2005. The exploitation of crop allelopathy in sustainable agricultural productions. *Journal of Agronomy and Crop Science* 191: 172-184.

Leather, G.R. 1983. Sunflowers (*Helianthus annuus*) are allelopathic to weeds. *Weed Science* 31: 37-42.

Lee, D.L., M.P. Prisbylla, T.H. Cromartie, D.P. Dagartin, S.W. Howard, W.M. Provan, M.K. Ellis, T. Fraser, and L.C. Mutter. 1997. The discovery and structural requirements of inhibitors of *p*-hydroxyphenylpyrvate dioxygenase. *Weed Science* 45: 601-609.

Li, Z., Q. Wang, X. Ruan, C. Pan, and D. Jiang. 2010. Phenolics and plant allelopathy. *Molecules* 15: 8933-8952.

Macias, F.A., J.M.G. Molinillo, A. Oliveros-Bastidas, D. Marin, and D. Chinchilla. 2004. Allelopathy. A natural strategy for weed control. *Communications in Applied Biological Science* 69: 13-23.

Macias, F.A., A. Oliveros-Bastidas, D. Marin, D. Castellano, A.M. Simonet, and J.M.G. Molinillo. 2005. Degradation studies on benzoxazinoids. Soil degradation dynamics of (2*R*)-2-*O*-β-D-glucopyranosyl-4-hydroxy-(2*H*)-1,4-benzoxazin-3(4*H*)-one (DIBOA-Glc) and its degradation products, phytotoxic allelochemicals from Gramineae. *Journal of Agricultural and Food Chemistry* 53: 554-561.

Maddi, V., K.S. Raghu, and M.N.A. Rao. 1992. Synthesis and anti-inflammatory activity of 3-(benzylideneamino)coumarins in rodents. *Journal of Pharmaceutical Sciences* 81: 964-966.

Malik, M.S., J.K. Norsworthy, A.S. Culpepper, M.B. Riley, and W. Bridges. 2008. Use of wild radish (*Raphanus raphanistrum*) and rye cover crops for weed suppression in sweet corn. *Weed Science* 56: 588-595.

Mithen, R. 2001. Glucosinolates and the degradation products. In *Advances in Botanical Research,* ed. J. Callow, 214-262. New York: Academic Press.

Molisch, H. 1937. *Der Einfluss einer Pflanze auf die andere-Allelopathie.* Fischer, Jena.

Netzly, D.H., J.L. Riople, G. Ejeta, and L.G. Butler. 1988. Germination stimulants of witchweed (*Striga asiatica*) from hydrophobic root exudate of sorghum (*Sorghum bicolor)*.Weed *Science* 36: 441-446.

Nimbal, C.I., J.F. Pedersen, C.N. Yerkes, L.A. Weston, S.C. Weller. 1996. Phytotoxicty and distribution of sorgoleone in grain sorghum germplasm. *Journal of Agriculture and Food Chemistry* 44: 1343-1347.

Norsworthy, J.K. and J.T. Meehan. 2005. Herbicidal activity of eight isothiocyanates onTexas panicum (*Panicum texanum*), large crabgrass (*Digitaria sanguinalis*), and sicklepod *(Senna obtusifolia*). *Weed Science* 53: 515-520.

Norsworthy, J.K., M.S. Malik, P. Jha, and M.B. Riley. 2007. Suppression of *Digitaria sanguinalis* and *Amaranthus palmeri* using autumn-sown glucosinolate-producing cover crops in organically grown bell pepper. *Weed Research* 47: 425-432.

Olofsdotter, M. 2001. Rice-a step toward use of allelopathy. *Agronomy Journal* 93: 3-8.

Patterson, D.T. 1981. Effects of allelopathic chemicals on growth and physiological response of soybean (*Glycine max*). *Weed Science* 29: 53-58.

Petersen, J., R. Belz, F. Walker, and K. Hurle. 2001. Weed suppression by release of isothiocyanates from turnip-rape mulch. *Agronomy Journal* 93: 37-43.

Pramanik, M.H.R., M. Nagal, M. Asao, and Y. Matsui. 2000. Effect of temperature and photoperiod on phytotoxic root exudates of cucumber (*Cucumis sativus*) in hydroponic culture. *Journal of Chemical Ecology* 26: 1953-1967.

Price, A. J., C. S. Charron, and C. E. Sams. 2005. Allyl isothiocyanate and carbon dioxide produced during degradation of *Brassica juncea* tissue in different soil conditions. HortSci. 40:1734-1739.

Price, A.J., D.W. Reeves, and M.G. Patterson. 2006. Evaluation of weed control provided by three winter cereals in conservation-tillage soybean. *Renewable Agriculture and Food Systems* 21: 159-164.

Price, A.J., M.E. Stoll, J.S. Bergtold, F.J. Arriaga, K.S. Balkcom, T.S. Kornecki, and R.L. Raper. 2008. Effect of cover crop extracts on cotton and radish radicle elongation. *Communications in Biometry and Crop Science* 3: 60-66.

Putnam, A.R., and W.B. Duke. 1978. Allelopathy in agroecosystems. *Annual Review of Phytopathology* 16: 431-451.

Putnam, A.R. 1985. Allelopathic research in agriculture: past highlights and potential. In *The Chemistry of Allelopathy,* ed. A.C. Thompson 1-8. Washington, D.C.: American Chemical Society.

Razavi, S.M., G.H. Imanzadeh, and M. Davari. 2010. Coumarins from *Zosima absinthifolia* seeds, with allelopathic effects. *Eurasion Journal of Biosciences* 4: 17-22.

Razavi, S.M. 2011. Plant coumarins as allelopathic agents. *International Journal of Biological Chemistry* 5: 86-90.

Rimando, A.M., F.E. Dayan, M.A. Czarnota, L.A. Weston, and S.O. Duke. 1998. A new photosystem II electron transfer inhibitor from *Sorghum bicolor. Journal of Natural Products* 61: 927-930.

SARE (Sustainable Agriculture Research and Education). 2007. *Managing Cover Crops Profitably,* 3rd edition. Ed. A. Clark. 244 pgs. College Park, MD: Sustainable Agriculture Research and Education.

Sarrantonio, M. and E. Gallandt. 2003. The role of cover crops in North American cropping systems. *Journal of Crop Production* 8: 53-74.

Secor, J. 1994. Inhibition of barnyardgrass 4-hydroxyphenylpyruvate dioxygenase by sulcotrione. *Plant Physiology* 106: 1429-1433.

Seigler, D.S. 1996. Chemistry and mechanisms of allelopathic interactions. *Agronomy Journal* 88: 876-885.

Sterling, T.M., R.L. Houtz, and A.R. Putnam. 1987. Phytotoxic exudates from velvetleaf (*Abutilon theophrasti*) glandular trichomes. *American Journal of Botany* 74: 543-550.

Swain, T. 1977. Secondary compounds as protective agents. *Annual Review of Plant Physiology* 28: 479-501.

Truman, C.C., D.W. Reeves, J.N. Shaw, A.C. Motta, C.H. Burmester, R.L. Raper, and E.B. Schwab. 2003. Tillage impacts on soil property, runoff, and soil loss variations of a Rhodic Paleudult under simulated rainfall. *Journal of Soil and Water Conservation* 58: 258-267.

Uren, N.C. 2000. Types, amounts, and possible functions of compounds released into the rhizosphere by soil-grown plants. In *The Rhizosphere: Biochemistry and Organic Substances at the Soil-Plant Interface*, eds. R. Pinton, Z. Varanini and P. Nannipieri, 19-40. New York: Marcel Dekker, Inc.

Vyvyan, J.R. 2002. Allelochemicals as leads for new herbicides and agrochemicals. *Tetrahedron*. 58: 1631-1646.

Walters, S.A. and B.G. Young. 2008. Utility of winter rye living mulch for weed management in zucchini squash production. *Weed Technology* 22: 724-728.

Weidenhamer, J.D. 1996. Distinguishing resource competition and chemical interference: Overcoming the methodological impasse: Allelopathy in cropping systems. *Agronomy Journal* 88: 866-875.

Weir, T.L., S. Park, and J.M. Vivanco. 2004. Biochemical and physiological mechanisms mediated by allelochemicals. *Plant Biology* 7: 472-479.

Westoby, M., M. Leishman, and J. Lord. 1996. Comparative ecology of seed size and dispersal. *Philosophical Transactions of the Royal Society of London B. Biological Sciences* 351: 1309-1318.

Weston, L.A. 1996. Utilization of allelopathy for weed management in agroecosystems: Allelopathy in cropping systems. *Agronomy Journal* 88: 860-866.

Willis, R.J. 1985. The historical bases of the concept of allelopathy. *Journal of the History of Biology* 18: 71-102.

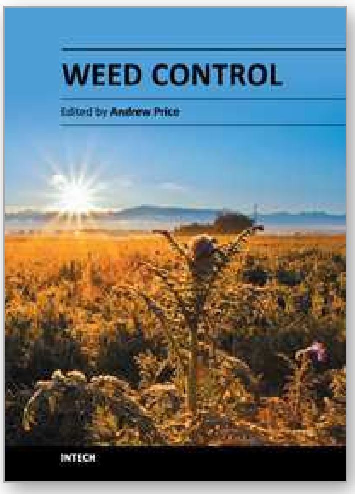
Wingard, C. 1996. Cover crops in integrated vegetable production systems. SARE Project report # PG95-033. Southern Region SARE. Griffin, GA. www.sare.org/projects.

Wolf, R.B., G.F. Spencer, and W.F. Kwolek. 1984. Inhibition of velvetleaf (*Abutilon theophrasti*) germination and growth by benzyl isothiocyanate, a natural toxicant. *Weed Science* 32: 612-615.

Wu, L., X. Guo, M.A. Harivandi. 1998. Allelopathic effects of phenolic acids detected in buffalograss (*Buchloe dactyloides*) clippings on growth of annual bluegrass (*Poa annua*) and buffalograss seedlings. *Environmental and Experimental Botany* 39: 159-167.

Yamamoto, Y. 2008. Movement of allelopathic compound coumarin from plant residue of Sweet vernalgrass (*Anthoxanthum odoratum* L.) to soil. *Japanese Society of Grassland Science* 55: 36-40.

Yu, J.Q., S.F. Ye, M.F. Zhan, and W.H. Hu. 2003. Effects of root exudates and aqueous root extracts of cucumber (*Cucumis sativus*) and allelochemicals, on photosynthesis and antioxidant enzymes in cucumber. *Biochemical Systematics and Ecology* 31: 129-139.



Weed Control

Edited by Dr. Andrew Price

ISBN 978-953-51-0159-8

Hard cover, 276 pages

Publisher InTech

Published online 29, February, 2012

Published in print edition February, 2012

Crop loss due to weeds has challenged agricultural managers since man began to develop the first farming systems. In the past century, however, much progress has been made to reduce weed interference in crop settings through effective yet mostly non-sustainable weed control strategies. With the commercial introduction of herbicides during the mid-1900's, advancements in chemical weed control tactics have provided efficient suppression of a broad range of weed species for most agricultural practices. Currently, with the necessity to design effective sustainable weed management systems, research has been pushing new frontiers on investigating integrated weed management options including chemical, mechanical as well as cultural practices. Author contributions to Weed Science present significant topics of research that examine a number of options that can be utilized to develop successful and sustainable weed management systems for many areas of crop production