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Comparison of Seminal Superoxide Dismutase (SOD) Activity Between Elite Athletes, Active and Non Active Men

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1. Introduction

It is increasingly recognized that reactive oxygen species (ROS) originating from the spermatozoa as well as from the leukocytes are of significant pathophysiological importance in the etiology of male infertility (Iwasaki and Gagnon, 1992; Zini *et al.*, 1993; Ochsendorf *et al.*, 1994; Shekarriz *et al.*, 1995; Smith *et al.*, 1996; Agarwal *et al.*, 2006; Tremellen, 2008). ROS, defined as including oxygen ions, free radicals and peroxides, may cause infertility by two principal mechanisms. First, ROS damage the sperm membrane which in turn reduces the sperm's motility and ability to fuse with the oocyte. Secondly, ROS directly damage sperm DNA, compromising the paternal genomic contribution to the embryo (Tremellen, 2008). Due to their high content of polyunsaturated fatty acids and their capacity to generate ROS, human spermatozoa are very sensitive to oxidative stress (Aitken and Clarkson, 1987; Aitken *et al.*, 1989; Smith *et al.*, 1996). To protect spermatozoa from oxidative damage, seminal plasma is endowed with numerous enzymatic antioxidants (AOs) such as superoxide dismutase (SOD), catalase and glutathione peroxidase (Fujii *et al.*, 2003; Garrido *et al.*, 2004; Murawski *et al.*, 2007). It has recently been reported, that superoxide anion (O_2^-) may be involved in fatty-acid peroxidation (Niess and Simon, 2007). Superoxide dismutase (SOD) and catalase inactivate the superoxide anion (O_2^-) and peroxide (H_2O_2) radicals by converting them into water and oxygen. SOD as an important element of seminal plasma superoxide anion scavenging capacity plays an essential role in maintaining the balance between ROS generation and degradation. Decrease of its capacity can result in abnormal sperm motility determined as sperm hyperactivation, and hence infertility (De Lamirande and Gagnon, 1993). The addition of SOD to sperm in culture has been confirmed to protect

them from oxidative attack (Kobayashi *et al.*, 1991). Although some investigators have shown no association between SOD activity and male fertility (Miesel *et al.*, 1997; Zini *et al.*, 2000; Hsieh *et al.*, 2002), others have reported a reduction in seminal plasma SOD activity in infertile males (Alkan *et al.*, 1997; Sanocka *et al.*, 1997; Siciliano *et al.*, 2001). Recently Murawski *et al.* (2007) reported a significantly lower semen SOD activity in infertile males, as compared with normospermic men. They showed a positive correlation between SOD activity in seminal plasma and semen quality parameters - sperm concentration and overall motility, which are regarded as most important for normal fertilizing ability of the spermatozoa (Murawski *et al.*, 2007).

Physical exercise has been shown to increase ROS and oxidative stress causing disruptions of homeostasis. Given that generation of oxidative stress is a natural part of physically active people may be susceptible to ROS-induced damage in sperm motility and male infertility, depending on the exercise mode, intensity, and duration as well as antioxidant capacity. Exercise training, on the other hand, has been shown to have modifying effects on oxidative stress, depending on training load, training specificity and the basal level of training. Growing evidence suggests that aerobic exercise training can result in an augmented SOD activity and a reduction in lipid peroxidation (Mena *et al.*, 1991; Ortenblad *et al.*, 1997; Ji, 1998; Suzuki and Ohno, 2000; Niess *et al.*, 2007). It is well documented that due to intensive training programs, trained and athletic people have developed total antioxidant capacity, and in particular high levels of SOD, in several tissues (Banfi *et al.*, 2006; Dekany *et al.*, 2006; Tayler *et al.*, 2006). However, our knowledge about antioxidant capacity in seminal plasma of athletic and trained men is less than meager, and it was up to now unclear if and under which conditions exercise training may influence seminal antioxidant capacity. Nonetheless, the amounts of enzymatic antioxidants in human semen have been well measured in normal and infertile men (Nissen and Kreysel, 1983; Jeulin *et al.*, 1989; Kobayashi *et al.*, 1991; Alkan *et al.*, 1997; Miesel *et al.*, 1997; Sanocka *et al.*, 1997; Zini *et al.*, 1993).

Taken together, considering the probable positive correlation and beneficial impact of SOD activity on human semen quality parameters and male fertility on the one hand, and the fact that the SOD and antioxidant capacity of various tissues are highly different between individuals with different level of physical fitness on the other hand, we wanted to find an answer to the question if the SOD activity of seminal plasma is different in individuals with different levels of physical activity. Thus, the purpose of this study was to evaluate the SOD activity in elite athletes, recreationally active and non active men.

2. Materials and methods

2.1 Subjects

A total of 40 semen samples were obtained from investigated groups in this study (Table 1). Of these, 15 samples were obtained from competitive elite athletes (e.g. including wrestlers, runners, football players, and swimmers) who were regularly training 4-5 days per week (Elite group). Thirteen samples were provided by non-obese and physically active males (active group) who participated in educational or recreational physical activities for 4-5 h per week. And 12 samples were obtained from healthy males who had a sedentary lifestyle without practicing in any sport for at least 6 months prior to study (control group). Physical activity of subjects was assessed by a questionnaire and subjects were matched based on training status and training quantity.

To be eligible to participate in the study, subjects were required to meet the following criteria: 1) unmarried men 18–28 year of age; 2) in good health, as determined by a normal physical examination and routine laboratory tests within the previous year; 3) no history of chronic disease, including reproductive disorders; 4) no history of use of medications that could alter the H-P-G axis, such as anabolic steroids; 5) regular eating patterns and no history of depressive illness; 6) normal physical and sexual development; 7) not working in professions where the activity might influence reproductive capacity; 8) no relevance previous surgery (eg, vasectomy reversal or varicocele removal) ; and 9) appropriate history of physical activity for the different groups described above (Lucía *et al.*, 1996; Di Luigi *et al.*, 2001). Informed consent was obtained from each subject. Before the initiation of the study protocol, each of them was introduced to the methods of this investigation. The Human Subject Internal Review Board committee of the Urmia University of IRAN approved the study (approval number 03/686).

Variable	Group	Elite	Active	Control	Sig*
Age (<i>year</i>)		23.1±1.3	23.4±0.9	22.0 ±2.0	0.053
Height (<i>cm</i>)		176.6±6.3	175.8±6.3	175.5±6.3	0.884
Weight (<i>kg</i>)		74.1±12.3	72.1±6.6	75.8±10.9	0.515
BMI (<i>kg/m²</i>)		20.9±2.5	22.1±1.4	24.3±2.6	*0.001
Fat (%)		8.5±3.7	11.2±3.1	15.5±5.1	*0.014

BMI = Body Mass Index.

*: P<0.05, significant difference between groups.

Table 1. Individual physical characteristics of subjects.

All subjects were given clear instructions on how to collect their semen at site. Each subject collected one semen sample by masturbation into a sterile container after at least 3-4 days of abstinence from ejaculations (Chia *et al.*, 1998; Nikoobakht *et al.*, 2005; Kao *et al.*, 2008). The majorities of samples were provided on site or were delivered to the laboratory within 30 min of collection. Each subject also completed a questionnaire concerning the duration (days) of the abstinence before collecting each sample (Jeulin *et al.*, 1989).

2.2 Seminal SOD activity measurement

Semen analyses were performed according to WHO guidelines (Caballero *et al.*, 1992; Nikoobakht *et al.*, 2005). Semen evaluations were performed on each sample by the same experienced technician throughout the study, for the assessment of SOD activity. All of the semen samples were then cryopreserved using the Test Yolk Buffer with Glycerol as a freezing medium, according to the protocol described in WHO guidelines (4-th edition, 1999).

SOD activity was measured by colorimetric assay (Zini *et al.*, 2000 and 2002). The commercially available colorimetric method was used (Randox Laboratories Ltd, UK). This method employs xanthine and xanthine oxidase to generate superoxide radicals which react with 2-(4-iodophenyl)-3-(4-nitrophenol) - 5-phenyltetrazoliumchloride (I.N.T) to form red formazan dye. The SOD activity is then measured by the degree of inhibition of this reaction. One unit of SOD inhibits reduction of INT by 50% under the conditions of the assay. After thawing, the seminal plasma was diluted 30-fold with 10 mM phosphate buffer, pH 7.0. The assay was performed at 37°C. Phosphate buffer was used as blank. Mixed

substrate and xanthine oxidase were added into standards and sample tubes and vortexed well. With spectrophotometer adjusted at a wavelength of 505 nm, the initial absorbance (A1) was read. Final absorbance (A2) was read exactly after 3 minutes, and percentages of inhibition of standards and samples were calculated. The SOD activity was measured using calibration curve of percentage inhibition for each standard against Log10 of standards and SOD activity was expressed as IU/ml.

2.3 Statistical analysis

Data are expressed as means \pm SD. Differences among groups were determined by analysis of variance (ANOVA) for continuous variables. If the *F*-ratio was significant, differences among groups were subsequently identified using a Bonferroni *post-hoc* analysis. The statistical software program SPSS for windows, version 17.0 was used for all data analyses. All statistical tests were performed at a significance level of 0.05.

3. Results

Semen parameters (mean \pm SD) were compared between study groups and the results are shown in Table 2. No significant differences were observed between groups in semen parameters ($P > 0.05$) except of normal morphology ($P \leq 0.05$). The result of Bonferroni test showed that the observed difference is more pronounced between elite group with active and control groups. However, there was no significant difference between active and control groups.

At this study the seminal SOD activity (IU/ml) of 3 groups was investigated. One-way ANOVA analysis showed that there is a significant difference between three groups (Table 2). The result of Bonferroni analysis revealed significantly higher SOD activity in seminal plasma of elite group than those of active and control groups ($P \leq 0.05$). However, no statistically significant difference was observed between active and control groups ($P > 0.05$).

Variable	Group	Elite	Active	Control	Sig*
Total sperm count (millions)		89.6 \pm 55.1	86.2 \pm 52.9	87.4 \pm 54.5	0.089
Volume (ml)		3.3 \pm 1.4	2.7 \pm 1.2	2.3 \pm 1.3	0.071
Concentration ($\times 10^6/ml$)		56.6 \pm 2.9	51.7 \pm 2.6	53.5 \pm 2.8	0.123
Motility (%)		72.8 \pm 15.6	66.2 \pm 16.1	69.7 \pm 14.9	0.068
Viability (%)		80.5 \pm 13.5	77.9 \pm 12.9	74.7 \pm 13.8	0.059
Normal morphology (%)		31.4 \pm 10.6	19.1 \pm 11.4	23.7 \pm 10.1	* 0.034
SOD (IU/ml)		34.3 \pm 9.4	17.9 \pm 5.6	24.2 \pm 6.6	* 0.001

*: $P < 0.05$, significant difference between groups.

Table 2. Comparison of Semen parameters and SOD activity between elite athletes, recreationally active and sedentary men.

4. Discussion

The results emerging from this study show that semen from elite athletes have higher SOD content than those of the active and sedentary (control) men. However, no significant difference was observed between samples of active and control groups. Therefore, these results represent that elite athletic men have developed seminal antioxidant capacity at least

in the case of SOD, suggesting that spermatozoa from elite athletes may be less susceptible to ROS-induced peroxidative damage, and hence, infertility. ROS have been shown to cause infertility by directly damaging the sperm DNA (Tremellen, 2008). Spermatozoa are susceptible to oxidative damage because their plasma membranes are rich in polyunsaturated fatty acids and have low concentrations of scavenging enzymes. SOD as one of the important elements of seminal plasma superoxide anion scavenging capacity plays an essential role in maintaining the balance between ROS generation and degradation through preventing increases in ROS concentration. Due to the protective effects against peroxidative damage and oxidative stress, seminal SOD has been shown to preserve sperm motility and viability. Sperm motility has been found to associate with men fertility (Jones *et al.*, 1979; Smith *et al.*, 1996; Murawski *et al.*, 2007; Agarwal *et al.*, 2008). The essential role of SOD as antioxidative defense enzyme is inferred from the observation that complete loss of motility of a sperm sample is directly proportional to the SOD activity of that sample (Storey, 1997). In particular, low SOD activity has been shown to be responsible for male infertility (Alkan *et al.*, 1997). Several investigators have reported reductions in SOD activity in semen of infertile men (Alkan *et al.*, 1997; Sanocka *et al.*, 1997; Siciliano *et al.*, 2001), although some have not (Zini *et al.*, 1993; Miesel *et al.*, 1997; Hsieh *et al.*, 2002). Recently Murawski *et al.* (2007) reported a significantly lower semen SOD activity in infertile men, as compared with normospermic men. They showed a positive correlation between SOD activity in seminal plasma and semen quality parameters - sperm concentration and overall motility, which are regarded as the most important for normal fertilizing ability of the spermatozoa (Murawski *et al.*, 2007).

Although the protective effect of seminal plasma has been well recognized, no study has investigated its antioxidative properties in individuals with different fitness level, in particular in athletic men. To the best of our knowledge, the present study provides the first evidence that elite athletes have an augmented SOD capacity than recreationally active and sedentary control men. Although there is no definitive explanation for this discrepancy, but the notion that high maximal oxygen uptake (VO_{2max}), which is a consequence of systemic endurance training (Tanaka and Swensen, 1998), are correlated to elevated antioxidant enzyme activity in other tissues (Jenkins *et al.*, 1984), can explain some of the disparities in our study. These observations suggest that the fitness level of subject as well as the type and amount of exercise training could be taken into consideration when comparing antioxidant capacity of active people. As the active group in our study were exercising just 4-5h per week, it seems possible that this amount of exercise is not enough to enhance antioxidant capacity. Dekany *et al.* (2006) have also referred to fitness level of athletes and type of exercise as important factors determining the blood level of antioxidant enzymes (Dekany *et al.*, 2006). In the review by Clarkson (1995) it has been documented that the "weekend athlete" may not have the augmented antioxidant defense system produced through continued training (Clarkson, 1995). This may make them more susceptible to oxidative stress.

The fact that rigorous exercise training programs may be required to promote antioxidant enzyme activity in skeletal muscle (Powers and Leeuwenburgh, 1999), can support our results. Intensive endurance training has been postulated as a potential muscle's antioxidant defense system up-regulator (Fatouros *et al.*, 2004). Powers *et al.* (1994) experimentally analyzed the relationship between the magnitude of the training stimulus (i.e., exercise intensity and daily duration) and the activity of SOD in locomotor skeletal muscles. Nine groups of rats ran at three different daily durations (i.e., 30, 60, 90 min.d) and three different exercise intensities (i.e., 55, 65, and 75% of VO_{2max}) (Powers *et al.*, 1994). Furthermore, previous studies have reported

that high-intensity running training can elevate antioxidant enzyme activities in erythrocytes and decrease neutrophil superoxide anion production both at rest and in response to exhaustive acute exercise (Miyazaki *et al.*, 2001). These data clearly show that high intensity long-term exercise training is superior to recreationally exercise in the up-regulation of seminal plasma SOD activity, representing adaptation to regular training. In fact, the training-stressed cells provide high antioxidant enzyme content. It has been generally believed that training induction of antioxidant enzymes is a cellular adaptation to oxidative stress caused by free radical generation during heavy exercises (Jenkins, 1988; Ji, 1998).

Prior to this work, numerous studies have been published focusing on the adaptation of different tissue's antioxidant capacity to exercise training (Mena *et al.*, 1991; Ortenblad *et al.*, 1997; Ji, 1998; Suzuki *et al.*, 2000; Banfi *et al.*, 2006; Dekany *et al.*, 2006; Tayler *et al.*, 2006; Garcia-Lopez *et al.*, 2007; Niess *et al.*, 2007). Recently Garcia Lopez *et al.* (2007) showed an increase in MnSOD mRNA levels of peripheral mononuclear cell (PBMC) in response to endurance training in middle-aged men (Garcia-Lopez *et al.*, 2007). Tauler *et al.* (2006) reported an increase in erythrocyte superoxide dismutase activity after the training/competition period for amateur trained male athletes (Tayler *et al.*, 2006). Banfi *et al.* (2006) showed a significantly higher plasma Glutathione reductase (GR) activity in trained elite soccer players, as compared with sedentary controls (Banfi *et al.*, 2006). Investigating antioxidant status of interval-trained athletes, Dekany *et al.* (2006) reported high level of blood enzymatic antioxidants in highly trained athletes (Dekany *et al.*, 2006). Ravi *et al.* (2004) showed a significantly increase in myocardium superoxide dismutase after 4 weeks low-intensity swim training in rats (Ravi *et al.*, 2004). Also, Yamamoto *et al.* (2002) reported higher superoxide dismutase activity and antioxidant capacity for physically active rats in compared to sedentary rats (Yamamoto *et al.*, 2002). However, both in treadmill-trained rats (Leeuwenburgh *et al.*, 1994) and running trained humans (Tiidus *et al.*, 1996) endurance training did not affect antioxidant defense.

Some limitations of the present study should be taken into consideration. First, we measured the SOD activity just in seminal plasma and did not measure in spermatozoa. Secondly, five subjects (active group n=2, and control group n=3) could not provide the semen sample in time and were excluded from the study. This might influence the compared results. Thirdly, this study comprised a relatively small sample thus limiting statistical power to detect differential effects. However, this is, to our knowledge, the first study analyzed the seminal antioxidant activity in individuals with different level of physical fitness.

In conclusion, the results of present study demonstrate that in compared to recreationally active and sedentary men, the elite athletes have developed SOD capacity. The mechanisms responsible for this increase are unknown and remain an active area of research. However, this raises the question that does the training-induced increase in seminal plasma SOD capacity provide increased protection against ROS-induced sperm dysfunction and infertility in high level athletes? If it does, we are able to provide novel insights into athletic health and male fertility. Further studies are warranted to deal the antioxidant capacity of seminal plasma and spermatozoa of individuals with different level of physical fitness as well as the effect of various exercise programs on seminal antioxidant capacity.

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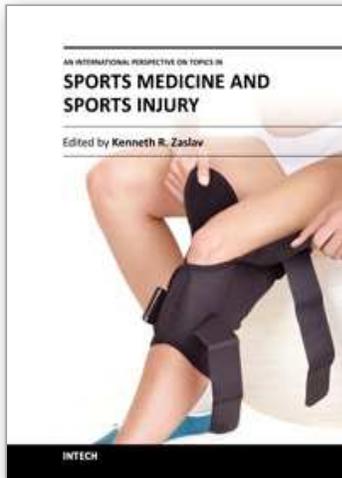
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