

We are IntechOpen, the world's leading publisher of Open Access books Built by scientists, for scientists

6,900

Open access books available

186,000

International authors and editors

200M

Downloads

Our authors are among the

154

Countries delivered to

TOP 1%

most cited scientists

12.2%

Contributors from top 500 universities



WEB OF SCIENCE™

Selection of our books indexed in the Book Citation Index
in Web of Science™ Core Collection (BKCI)

Interested in publishing with us?
Contact book.department@intechopen.com

Numbers displayed above are based on latest data collected.
For more information visit www.intechopen.com



An Animal Model of Sepsis in Appendicitis: Assessment of the Microcirculation

Eduardo Ryoiti Tatebe, Priscila Aikawa, José Jukemura,
Paulina Sannomiya and Naomi Kondo Nakagawa
*University of São Paulo School of Medicine
Brazil*

1. Introduction

Acute appendicitis is one of the most common causes of inflammation in the abdomen. Appendicitis, characterized by inflammation of the appendix, is an urgent clinical illness with significant morbidity, which increases with diagnostic delay. Perforation and peritonitis are associated with increased morbidity and mortality, especially in the very young, the elderly and immune-suppressed patients.

The diagnosis is based on the patient's history by the classic signs and symptoms of appendicitis (abdominal pain in the right iliac fossa, fever, anorexia, nausea, and vomiting) and physical examination. Children and the elderly have fewer signs and symptoms, or cannot adequately describe them. In pregnant women, particularly during the second and third trimester, the diagnosis of acute appendicitis is often delayed because of the nonspecific clinical abdominal presentation. In these conditions, diagnosis often requires imaging methods (ultrasound and/or CT scanning), and the incidence of complications is more frequent. Most patients usually recover well after surgical treatment, but complications can occur if treatment is delayed or if perforation that results in peritonitis or sepsis is present.

Sepsis and septic shock are clinical syndromes that result from complex interactions between the host and infectious agents. These events are characterized by hemodynamic derangements, widespread microcirculatory disturbances and cellular alterations leading to heterogeneous flow distribution, capillary obstruction and, therefore, to an uncoupling between cellular oxygen need and oxygen supply (De Backer et al., 2002; Hinshaw, 1996; Sakr et al., 2004). Despite improvements in treatments for sepsis, there are still gaps in our knowledge of the physiopathology and therapeutic interventions.

1.1 Cecal Ligation and Puncture as an experimental model of appendicitis

Among several experimental animal models, perforated appendicitis by cecal ligation and puncture (CLP), particularly in rodents, has been used to investigate the pathophysiology and assess the effectiveness of therapies in sepsis and septic shock. The CLP model begins with bowel exposure, followed by cecal ligation distal to the ileocecal valve. Thereafter, the cecum is perforated by a needle and the contents squeezed into the peritoneal cavity. The number of punctures, the diameter of the hole and the total amount of squeezed bowel content can introduce several variations of the model that will directly induce lethal or non-

lethal sepsis. Sepsis in the CLP model is caused by contamination of the peritoneal cavity with a mixed flora of microorganisms and by the ischemic/necrotic tissue complications. Without the appropriate clinical (fluid resuscitation and antibiotics) and surgical treatment (necrotic tissue resection and peritoneal lavage), a rapid onset of septic shock can be observed.

1.1.1 Advantages of the CLP model

In this work, we will focus on the experimental model of CLP in rodents that is a simple and reproducible model widely used in research. The CLP model allows for control of the setting and reduction of some of the variables. We have focused on the mechanisms responsible for the altered immunological, cardiovascular, respiratory and metabolic changes as a model for acute perforated appendicitis in humans.

The CLP model can also be used to evaluate cardiac output/total and regional blood flow (Angle et al., 1998; Jarrar et al., 2000; Yang et al., 2002), metabolism (Lang et al., 1990; Wang et al., 1999), immune function (Ayala & Chaudry, 1996; Kato et al., 2004; Schneider et al., 2000), apoptosis (Reddy et al., 2001; Ayala & Chaudry, 1996; Chung et al., 2003; Coopersmith et al., 2002), cytokines (Schneider et al., 2000; Vianna et al., 2004), resuscitation (Esmon, 2004; Marx et al., 2004; Yang et al., 2002), antibiotics (Doerschug et al., 2004; Vianna et al., 2004), and microbial components (Ayala & Chaudry, 1996; Esmon, 2004; Mollitt, 2002; Yang et al., 2001).

1.1.2 Limitations of the CLP model

The CLP model in small mammals, particularly rodents, has some limitations on the translation to humans. One difference that is common between rodents and humans is that the mice or rats can tolerate quite well the cecal ligation alone without puncture. These animals can block the necrotic tissue and survive. Humans, in turn, are not able to overcome by themselves. Another aspect is related to the size of the animal. Several technical and physiological difficulties may appear. Among them, the inability to obtain large quantities of blood and other fluids for tests over long periods of observation (Hubbard et al., 2005), and the technology to obtain accurate physiological measurements in these small animals.

1.2 Other models of acute appendicitis

Rabbits and pigs have also been used as experimental models of acute appendicitis. However, due to anatomical differences, the use of pigs is very limited. Pig does not have an appendix and the occlusion is performed in the uterine horn to study surgical procedures, such as an endoscopic transgastric appendectomy (Sumiyama et al., 2006). In rabbits, investigators use the vascular partial or total clamping method to obtain necrotic tissue mimicking the acute appendicitis (Nunes & Silva, 2005).

2. Intravital microscopy in the assessment of microcirculation

A well-established technique applied in many experimental models of sepsis is the intravital microscopy (Figure 1). This technique allows the *in vivo* and *in situ* observation of the microvascular bed of different tissues, such as the mesentery, ileum, liver, and skeletal muscle of rats, mice, rabbits and felines. Suitable tissues are selected if they can be easily exteriorized and transilluminated, as illustrated in Figure 2. It is important to minimize the preparative surgery and to maintain the physiological conditions: temperature, extracellular

fluid composition, pH, and gas tensions. The introduction of close circuit television has facilitated quantification of many of the variables, such as leukocyte-endothelial interactions, through the possibility to store images on videotape for detailed off-line analysis. More recently, analyses have been performed online by using image-computer software (Nakagawa et al, 2006).



Fig. 1. Equipment for Intravital Microscopy.

Intravital microscopy has been applied to evaluate different pathophysiological aspects of the microcirculation during several challenges. In addition, intravital microscopy has also been used to test novel prophylactic and therapeutic approaches that aim to prevent or attenuate manifestations of sepsis-associated microvascular disorders and cellular dysfunctions. In the mesentery, microcirculatory observations have focused on capillary obstruction, capillary or arteriolar density, microvessel reactivity and leukocyte-endothelial interactions in post-capillary venules (Harris, 2006; Kim & Harris, 2006; Nakagawa et al., 2006, 2007; Schmidt et al., 1997; Smalley et al., 2000; Walther et al., 2004; Woodman et al., 2000). A representative photomicrograph of rat mesenteric microcirculation is shown in Figure 3. In other organs, such as lungs and heart, increased leukocyte-endothelial interactions have been observed mostly induced by physical trapping in pre-capillary microvessels and capillaries (Kubo et al., 1999; Waisman et al., 2006). In liver, blood flow/perfusion regulation is at the arteriolar and sinusoidal level (Baveja et al., 2002; Kamoun et al., 2005).

Microcirculatory dysfunctions, as seen in humans (De Backer & Dubois, 2001; De Backer et al., 2002; Groner et al., 1999; Trzeciak et al., 2007), have been shown to occur in most experimental models of sepsis (Baveja et al., 2002; Kamoun et al., 2005; Kim & Harris, 2006; Kubo et al., 1999; Nakagawa et al., 2006, 2007; Nakajima et al., 2001; Schmidt et al., 1997; Smalley et al., 2000; Waisman et al., 2006; Walther et al., 2004; Woodman et al., 2000). Endotoxin infusion is a widely used experimental model (Kim & Harris, 2006; Schmidt et al., 1997; Smalley et al., 2000; Nakajima et al., 2001). Increased leukocyte-endothelial interactions and protein leakage in mesenteric microvessels have been shown to occur after acute endotoxemia in rats (Schmidt et al., 1997; Woodman et al., 2000) and cats (Walther et al., 2004). However, there are two major concerns regarding this experimental model: 1) clinical sepsis typically evolves over many days, in contrast to studies on the early effects (1 to 6 hours) of endotoxin administration, and 2) rats are generally more resistant to the effects of endotoxin.



Fig. 2. Positioning of the mesentery in the platform for intravital microscopy.

Therefore, many microvascular changes seen in animal models of endotoxemia may not occur in humans (Chaudry, 1999). On the other hand, laparotomy complicated by sepsis is a common clinical presentation of sepsis. This rationale was used in selecting CLP as a model of polymicrobial and normotensive sepsis (Chaudry, 1999; Chaudry et al., 1979; Farquhar et al., 1996; Hersch et al., 1998; Madorin et al., 1999; Nakagawa et al., 2006, 2007).

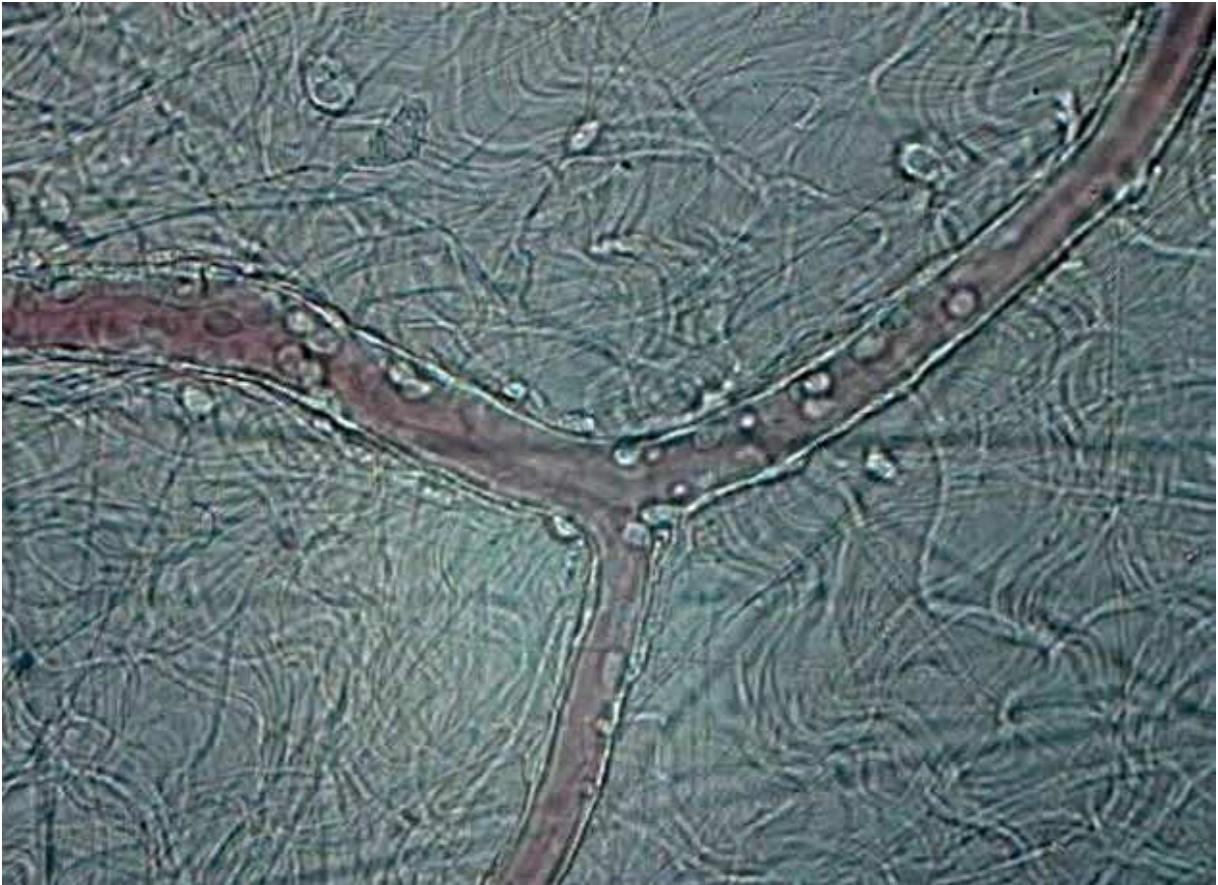


Fig. 3. Photomicrograph of rat mesenteric microcirculation by intravital microscopy showing leukocyte-endothelial interactions under inflammation (425x)

In the CLP model (Figure 4), many microvascular derangements occur such as increased total blood flow to the ileum with preferential redistribution toward the muscularis and away from the mucosa (Farquhar et al., 1996; Hersch et al., 1998; Madorin et al., 1999; Nakajima et al., 2001). The abnormal microvascular blood flow may result in tissue hypoxia and increased permeability (Farquhar et al., 1996). CLP induces an inflammatory response characterized by an increased number of white blood cells, increased leukocyte-endothelial interactions in mesenteric microvessels and lung neutrophil infiltration (Nakagawa et al., 2006).

3. CLP in a double-injury model

In attempting to understand the pathophysiology of septic shock, several investigators have performed double-injury models to study the microcirculation by intravital microscopy in different tissues. Hoffman et al. (1999) observed increased leukocyte adhesion and reduced capillary perfusion in skin microvessels of hamsters submitted to persistent endotoxemia (72 hours) induced by a double-LPS exposure. Swartz et al. (2000) performed CLP followed by the local application of *E. coli* on cremaster muscle. Despite an intra-abdominal infection, there was no increase in leukocyte adhesion in the cremaster muscle. In contrast, Pascual et al. (2003) observed increased leukocyte adhesion to microvessels of cremaster muscle after hemorrhagic shock/reperfusion followed by intratracheal injection of LPS in mice. Smalley

et al. (2000) reported no changes in leukocyte adhesion in the mesentery in an acute model (4 hours) of CLP. However, a topical application of highly diluted fecal matter increased leukocyte adhesion in the mesenteric microcirculation, which was mediated by platelet activation factor. More recently, Nakagawa et al. (2006) observed leukocyte-endothelium interactions in the mesenteric microcirculation after hemorrhagic shock/reperfusion followed by an intra-abdominal sepsis (CLP). Twenty-four hours after the double-injury, rats exhibited an increased number of rolling, adherent and migrated leukocytes accompanied by an increased expression of P-selectin and intercellular adhesion molecule (ICAM)-1 at the mesentery and by leukocyte infiltration and ICAM-1 up-regulation at the lungs.



Fig. 4. Twenty-four hours after cecal ligation and puncture model. Note the impressive intestinal edema around the necrotic tissue.

4. Surgical control of the septic focus

In the model of single-injury (CLP) the surgical removal of the septic focus followed by peritoneal lavage partially controls the inflammatory reaction in these animals. By intravital microscopy, leukocyte-endothelial interactions at the mesentery were normalized by the surgical control. These results support surgical source control as a therapy contributing to resolving the immune dysfunction observed in this specific septic challenge (Nakagawa et al., 2007).

5. Conclusion

Cecal ligation and puncture in rodents is a useful experimental model that mimics appendicitis with pathophysiological alterations enrolled in this process. Surgical removal of the septic focus improves clinical condition and normalizes physiological aspects that are clearly observed in this model. In addition, the study of the microcirculation by intravital microscopy represents a unique research tool to analyse complex biological interactions and disease-related mechanisms.

6. References

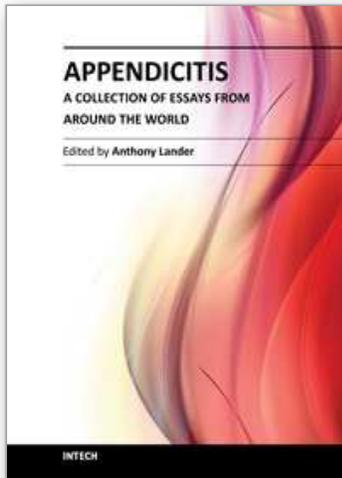
- Angle, N.; Hoyt, D.B.; Coimbra, R.; Liu, F.; Herdon-Remelius, C.; Loomis, W. & Junger, W.G. (1998). Hypertonic saline resuscitation diminishes lung injury by suppressing neutrophil activation after hemorrhagic shock. *Shock*, Vol. 9, No. 3, (March 1998), pp. 164-170, ISSN 1073-2322
- Ayala, A.; Chaudry, I.H. (1996). Immune dysfunction in murine polymicrobial sepsis: mediators, macrophages, lymphocytes and apoptosis. *Shock*, Vol. 6, No. 4, (October 1996), pp. S27-S38, ISSN 1073-2322
- Baveja, R.; Kresge N.; Ashburn, J.H.; Keller, S.; Yokoyama, Y.; Sonin, N.; Zhang, J.X.; Huynh, T. & Clemens, M.G. (2002). Potentiated hepatic microcirculatory response to endothelin-1 during polymicrobial sepsis. *Shock*, Vol.18, No. 5, (November 2002), pp. 415-422, ISSN 1073-2322
- Chaudry, I.H.; Wichterman, K.A. & Baue, A.E. (1979). Effect of sepsis on tissue adenine nucleotide levels. *Surgery*, Vol. 85, No. 2 (February 1979), pp. 205-211, ISSN 0039-6060
- Chaudry, I.H. (1999). Sepsis: lessons learned in the last century and future directions. *Archives of Surgery*, Vol. 134, No. 9, (September 1999), pp. 922-929, ISSN 0004-0010
- Coopersmith, C.M.; Chang, K.C.; Swanson, P.E.; Tinsley, K.W.; Stromberg, P.E.; Buchman, T.G.; Karl, I.E. & Hotchkiss, R.S. (2002). Overexpression of Bcl-2 in the intestinal epithelium improves survival in septic mice. *Critical Care Medicine*, Vol. 30, No. 1, (January 2002), pp. 195-201, ISSN 0090-3493
- De Backer, D. & Dubois, M.J. (2001). Assessment of the microcirculatory flow in patients in the intensive care unit. *Current Opinion in Critical Care*, Vol. 7, No. 3, (June 2001), pp. 200-203, ISSN 1070-5295
- De Backer, D.; Creteur, J.; Preiser, J.C.; Dubois, M.J. & Vincent, J.L. (2002). Microvascular blood flow is altered in patients with sepsis. *American Journal of Respiratory and Critical Care Medicine*, Vol. 166, No. 1, (July 2002), pp. 98-104, ISSN 1535-4970
- Doerschug, K.C.; Powers, L. S.; Monick, M.M.; Thorne, P.S. & Hunninghake, G.W. (2004). Antibiotics delay but do not prevent bacteremia and lung injury in murine sepsis. *Critical Care Medicine*, Vol. 32, No. 2, (February 2004), pp. 489-494, ISSN 0090-3493
- Esmon, C.T. (2004). Why do animal models (sometimes) fail to mimic human sepsis? *Critical Care Medicine*, Vol. 32, No. 5, (May 2004), pp.S202-S208, ISSN 0090-3493
- Farquhar, I.; Martin, C.M.; Lam, C.; Potter, R.; Ellis, C.G. & Sibbald, W.J. (1996). Decreased capillary density in vivo in bowel mucosa of rats with normotensive sepsis. *Journal of Surgical Research*, Vol. 61, No. 1, (February 1996), pp. 190-196, ISSN 0022-4804
- Groner, W.; Winkelman, J.W.; Harris, A.G.; Ince, C.; Bouma, G.J.; Messmer, K. & Nadeau, R.G. (1999). Orthogonal polarization spectral imaging: a new method for study of

- the microcirculation. *Nature Medicine*, Vol. 5, No. 10, (October 1999), pp. 1209-1213, ISSN 1078-8956
- Hersch, M.; Madorin, W.S.; Sibbald, W.J. & Martin, C.M. (1998). Selective gut microcirculatory control (SGMC) in septic rats: a novel approach with a locally applied vasoactive drug. *Shock*, Vol. 10, No. 4, (October 1998), pp. 292-297, ISSN 1073-2322
- Hinshaw, L.B. (1996). Sepsis/septic shock: participation of the microcirculation: an abbreviated review. *Critical Care Medicine*, Vol. 24, No.6, (June 1996), pp. 1072-1078. ISSN 0090-3493
- Hoffman, J.N.; Vollmar, B.; Inthorn, D.; Schildberg, F.W. & Menger, M.D. (1999). A chronic model for intravital microscopic study of microcirculatory disorders and leukocyte/endothelial cell interactions during normotensive endotoxemia. *Shock*, Vol. 12, No.5, (November 1999), pp. 355-364, ISSN 1073-2322
- Hubbard, W.J.; Choudhry, M.; Schwacha, M.G.; Kerby, J.D.; Rue III, L.W.; Bland, K.I. & Chaudry, I.H. (2005). Cecal Ligation and Puncture. *Shock*, Vol. 24, No. 1, (December 2005), pp. 52-57, ISSN 1073-2322
- Jarrar, D.; Wang, P.; Song, G.Y.; Cioffi, W.G.; Bland, K.I. & Chaudry, I.H. (2000). Inhibition of Tyrosine Kinase Signaling After Trauma-Hemorrhage: A Novel Approach for Improving Organ Function and Decreasing Susceptibility to Subsequent Sepsis. *Annals of Surgery*, Vol. 231, No. 3, (March 2000), pp. 399-407, ISSN 0003-4932
- Kamoun, W.S.; Shin, M.C.; Keller, S.; Karaa, A.; Huynh, T. & Clemens, M.G. (2005). Induction of biphasic changes in perfusion heterogeneity of rat liver after sequential stress in vivo. *Shock*, Vol. 24, No. 4, (October 2005), pp. 324-331, ISSN 1073-2322
- Kato, T.; Hussein, M.H.; Sugiura, T.; Suzuki, S.; Fukuda, S.; Tanaka, T.; Kato, I. & Togari, H. (2004). Development and characterization of a novel porcine model of neonatal sepsis. *Shock*; Vol. 21, No. 4, (April 2004), pp. 329-335, ISSN 1073-2322
- Kim, M.H. & Harris, N.R. (2006). Leukocyte adherence inhibits adenosine-dependent venular control of arteriolar diameter and nitric oxide. *American Journal of Physiology-Heart and Circulatory Physiology*, Vol. 291, No. 2, (August 2006), pp. H724-731, ISSN 0363-6135
- Kubo, H.; Doyle, N.A.; Graham, L.; Bragwan, S.D.; Quinlan, W.M. & Doerschuk, C.M. (1999). L- and P-selectin and CD11/CD18 in intracapillary neutrophil sequestration in rabbit lungs. *Am J Respir Crit Care Med* 1999;159:267-274.
- Lang, C.H.; Dobrescu, C. & Mészáros, K. (1990). Insulin-mediated glucose uptake by individual tissues during sepsis. *Metabolism*, Vol. 39, No.10, (October 1990), pp. 1096-1107, ISSN 0026-0495
- Madorin, W.S.; Martin, C.M. & Sibbald, W.J. (1999). Dopexamine attenuates flow motion in ileal mucosal arterioles in normotensive sepsis. *Critical Care Medicine*, Vol. 27, No. 2 (February 1999), pp. 394-400, ISSN 0090-3493
- Marx, G.; Pedder, S.; Smith, L.; Swaraj, S.; Grime, S.; Stockdale, H. & Leuwer, M. (2004). Resuscitation from septic shock with capillary leakage: hydroxyethyl starch (130 Kd), but not Ringer's solution maintains plasma volume and systemic oxygenation. *Shock*, Vol. 21, No. 4, (April 2004), pp. 336-341, ISSN 1073-2322
- Mollitt, D.L. (2002). Infection control: avoiding the inevitable. *Surgical Clinics of North America*, Vol. 82, No. 2, (April 2002), pp 365-378, ISSN 0039-6109

- Nakagawa, N.K.; Nogueira, R.F.; Correia, C.J.; Shiwa, S.R.; Costa Cruz, J.W.M.; Poli de Figueiredo, L.F.; Rocha e Silva, M. & Sannomiya, P. (2006). Leukocyte-endothelium interactions after hemorrhagic shock/reperfusion and cecal ligation/puncture: an intravital microscopic study in rat mesentery. *Shock*, Vol. 26, No. 2, (August 2006), pp. 180-186, ISSN 1073-2322
- Nakagawa, N.K.; Jukemura, J.; Aikawa, P.; Nogueira, R.A.; Poli de Figueiredo, L.F. & Sannomiya, P. (2007). In vivo observation of mesenteric leukocyte-endothelial interactions after cecal ligation/puncture and surgical source control. *Clinics*, Vol. 62, No. 3, (May-June 2007), pp. 321-326, ISSN 1807-5932
- Nakajima, Y.; Baudry, N.; Duranteau, J. & Vicaut, E. (2001). Microcirculation in intestinal villi. *American Journal of Respiratory and Critical Care Medicine*, Vol. 164, No. 8, (October 2001), pp. 1526-1530, ISSN 1535-4970
- Nunes, F.C. & Silva, A.L.(2005). Acute ischaemic Appendicitis in rabbits: new model with histopathological study. *Acta Cirurgica Brasileira*, Vol. 20, No. 5, (September-October 2005), pp. 399-404, ISSN 1678-2674
- Pascual, J.L.; Khwaja, K.A.; Ferri, L.E.; Giannias, B.; Evans, D.C.; Razek, T.; Michel, R.P. & Christou, N.V. (2003). Hypertonic saline resuscitation attenuates neutrophil lung sequestration and transmigration by diminishing leukocyte-endothelial interactions in a two hit model of hemorrhagic shock and infection. *Journal of Trauma* Vol. 54, No. 1, (January 2003), pp. 121-132, ISSN 0022-5282
- Sakr, Y.; Dubois, M.J.; De Backer, D.; Creteur, J. & Vincent, J.L. (2004). Persistent microcirculatory alterations are associated with organ failure and death in patients with septic shock. *Critical Care Medicine*, Vol. 32, No. 9, (September 2004), pp. 1825-1831, ISSN 0090-3493
- Schmidt, H.; Schmidt, W.; Muller, T.; Bohrer, H.; Gebhard, M.M. & Martin, E. (1997). N-acetylcysteine attenuates endotoxin-induced leukocyte-endothelial cell adhesion and macromolecular leakage in vivo. *Critical Care Medicine*, Vol. 25, No. 5, (May 1997), pp. 858-863, ISSN 0090-3493
- Schneider, C.P.; Nickel, E.A.; Samy, A.T.S.; Schwacha, M.G.; Cioffi, W.G.; Bland, K.I.; Chaudry, I.H.(2000). The aromatase inhibitor, 4-hydroxyandrostenedione, restore immune responses following trauma-hemorrhages in males and decreases mortality from subsequent sepsis. *Shock*; Vol. 14, No. 3, (September 2000), pp. 347-353, ISSN 1073-2322
- Smalley, D.M.; Childs, E.W. & Cheung, L.Y. (2000). The local effect of PAF on leukocyte adherence to small bowel mesenteric venules following intra-abdominal contamination. *Inflammation*, Vol. 24, No. 5, (October 2000), pp. 399-410, ISSN 0360-3997
- Sumiyama, K.; Gostout, C.J.; Rajan, E.; Bakken, T.A.; Deters, J.L.; Knipschild, M.A.; Hawes, R.H.; Kalloo, A.N.; Pasricha, P.J.; Chung, S.; Kansevov, S.V. & Cotton, P.B. (2006). Pilot Study of the porcine uterine horn as an in vivo appendicitis model for development of endoscopic transgastric appendectomy. *Gastrointestinal Endoscopy*, Vol. 64, No. 5, (November 2006), pp. 808-812, ISSN 0016-5107
- Swartz, D.E.; Seely, A.J.E.; Ferri, L.; Giannias, B. & Christou, N.V. (2000). Decreased systemic polymorphonuclear neutrophil (PMN) rolling without increased PMN adhesion in peritonitis at remote sites. *Archives of Surgery*, Vol. 135, No. 8, (August 2000), pp. 959-966, ISSN 0004-0010

- Trzeciak, S.; Dellinger, R.P.; Parrillo, J.E.; Guglielmi, M.; Bajaj, J.; Abate, N.L.; Arnold, R.C.; Colilla, S.; Zanutti, S.; & Hollenberg, S.M. (2007). Early microcirculatory perfusion derangements in patients with severe sepsis and septic shock: relationship to hemodynamics, oxygen transport, and survival. *Annals of Emergency Medicine*, Vol.49, No. 1 (January 2007), pp. 88-98, ISSN 0196-0644
- Vianna, Rosa C.S.; Gomes, R.N.; Bozza, F.A.; Amancio, R.T.; Bozza, P.T.; David, C.M.N. & Castro-Faria-Neto, H.C. (2004). Antibiotic Treatment in a Murine Model of Sepsis: Impact on Cytokines and Endotoxin Release. *Shock*, Vol. 21, No. 2, (February 2004), pp. 115-120, ISSN 1073-2322
- Waisman, D.; Abramovich, A.; Brod, V.; Lavon, O.; Nurkin, S.; Popovski, F.; Rotschild, A. & Bitterman, H. (2006). Subpleural microvascular flow velocities and shear rates in normal and septic mechanically ventilated rats. *Shock*, Vol. 26, No. 1, (July 2006), pp. 87-94, ISSN 1073-2322
- Walther, A.; Czabanka, M.; Gebhard, M.M. & Martin, E. (2004). Glycoprotein IIB/IIIa-inhibition and microcirculatory alterations during experimental endotoxemia - an intravital microscopic study in the rat. *Microcirculation*, Vol. 11, No. 1, (January-February 2004), pp. 79-88, ISSN 1073-9688
- Wang, P.; Ba, Z.F.; Cioffi, W.G.; Bland, K.I. & Chaudry, I.H.(1999). Salutary effects of ATP-MgCl₂ on the depressed endothelium-dependent relaxation during hyperdynamic sepsis. *ritical Care Medicine*, Vol. 27, No. 5,(May 1999), pp. 959-964, ISSN 0090-3493
- Woodman, R.C.; Teoh, D.; Payne, D. & Kubes, P. (2000). Thrombin and leukocyte recruitment in endotoxemia. *American Journal of Physiology-Heart and Circulatory Physiology*, Vol. 279, No. 3, (September 2000), pp. H1338-1345, ISSN 0363-6135
- Yang, S.; Zhou, M.; Chaudry, I.H. & Wang, P. (2001). The role of lipopolysaccharide in stimulating adrenomedullin production during polymicrobial sepsis. *Biochimica et Biophysica Acta-Molecular Basis of Disease*, Vol. 1537, No. 2, (September 2001), pp 167-174, ISSN 0925-4439
- Yang, S.; Zhou, M.; Chaudry, I.H. & Wang, P. (2002). Novel Approach to prevent the transition from the hyperdynamic phase to the hypodynamic phase of sepsis: role of adrenomedullin and adrenomedullin-binding protein-1. *Annals of Surgery*, Vol. 236, No 5, (November 2002), pp. 625-633, ISSN 0003-4932

IntechOpen



Appendicitis - A Collection of Essays from Around the World

Edited by Dr. Anthony Lander

ISBN 978-953-307-814-4

Hard cover, 226 pages

Publisher InTech

Published online 11, January, 2012

Published in print edition January, 2012

This book is a collection of essays and papers from around the world, written by surgeons who look after patients of all ages with abdominal pain, many of whom have appendicitis. All general surgeons maintain a fascination with this important condition because it is so common and yet so easy to miss. All surgeons have a view on the literature and any gathering of surgeons embraces a spectrum of opinion on management options. Many aspects of the disease and its presentation and management remain controversial. This book does not answer those controversies, but should prove food for thought. The reflections of these surgeons are presented in many cases with novel data. The chapters encourage us to consider new epidemiological views and explore clinical scoring systems and the literature on imaging. Appendicitis is discussed in patients of all ages and in all manner of presentations.

How to reference

In order to correctly reference this scholarly work, feel free to copy and paste the following:

Eduardo Ryoiti Tatebe, Priscila Aikawa, José Jukemura, Paulina Sannomiya and Naomi Kondo Nakagawa (2012). An Animal Model of Sepsis in Appendicitis: Assessment of the Microcirculation, *Appendicitis - A Collection of Essays from Around the World*, Dr. Anthony Lander (Ed.), ISBN: 978-953-307-814-4, InTech, Available from: <http://www.intechopen.com/books/appendicitis-a-collection-of-essays-from-around-the-world/an-animal-model-of-sepsis-in-appendicitis-assessment-of-the-microcirculation>

INTECH
open science | open minds

InTech Europe

University Campus STeP Ri
Slavka Krautzeka 83/A
51000 Rijeka, Croatia
Phone: +385 (51) 770 447
Fax: +385 (51) 686 166
www.intechopen.com

InTech China

Unit 405, Office Block, Hotel Equatorial Shanghai
No.65, Yan An Road (West), Shanghai, 200040, China
中国上海市延安西路65号上海国际贵都大饭店办公楼405单元
Phone: +86-21-62489820
Fax: +86-21-62489821

© 2012 The Author(s). Licensee IntechOpen. This is an open access article distributed under the terms of the [Creative Commons Attribution 3.0 License](#), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

IntechOpen

IntechOpen