We are IntechOpen, the world's leading publisher of Open Access books Built by scientists, for scientists

6,300

Open access books available

172,000

International authors and editors

190M

Downloads

Our authors are among the

154

Countries delivered to

TOP 1%

most cited scientists

12.2%

Contributors from top 500 universities

WEB OF SCIENCE™

Selection of our books indexed in the Book Citation Index
in Web of Science™ Core Collection (BKCI)

Interested in publishing with us?
Contact book.department@intechopen.com

Numbers displayed above are based on latest data collected.
For more information visit www.intechopen.com

Resolution of Colitis-Associated Inflammation

Hennebert Olivier, Pelissier Marie-Agnès and Morfin Robert
Biotechnologie, CNAM, Paris, France

1. Introduction

Ulcerative colitis (UC) is an inflammatory bowel disease related to chronic relapsing inflammation of the intestinal tract of unknown aetiology (Podolsky, 2003). Maintenance of colonic inflammation involves a network of inflammatory mediators resulting in a dramatic increase in the production of reactive oxygen species that contribute to the functional change characteristic of UC (Pavlick et al, 2002). Animal models provide evidence that altered cytokine (IL-1 β) and eicosanoid secretion patterns may play a role in UC (Blumberg et al, 1999; Stenson, 2007). Eicosanoids, including leukotrienes generated by the 5-lipoxygenase pathway (Murthy et al, 1997) and prostaglandins (PGs) produced by the cyclooxygenases COX-1 or COX-2, are lipid mediators implicated in the pathophysiology of UC (Funk, 2001).

Inflammation in experimentally induced colitis is characterized by increased PGs such as PGE₂ and PGD₂. PGD₂ (Ajuebor et al, 2000; Melgar et al, 2006) is further converted by dehydration to 15-deoxy- $\Delta^{12,14}$ -PGJ₂ (15d-PGJ₂) (Monneret et al, 2002), a stable PG and putative endogenous PPAR γ ligand with cytoprotective and anti-inflammatory properties, and hence a new potential therapeutic target in inflammatory bowel diseases (Dubuquoy et al, 2006).

7 α -Hydroxy-DHEA, the innate metabolite of dehydroepiandrosterone (DHEA), is normally produced in the colon and many other tissues (Doostzadeh & Morfin, 1996; Morfin & Courchay, 1994) but overproduced by IL-1 β during the inflammatory process as shown in mice and humans (Dulos et al, 2004; Dulos et al, 2005). We have previously shown that pre-treatment with DHEA and 7 α -hydroxy-DHEA resulted in protective anti-oxidant effects in the colon of healthy and colitis rats (Pelissier et al, 2004; Pelissier et al, 2006). 7 α -Hydroxy-DHEA is converted by NADP(H)-dependent 11 β -hydroxysteroid dehydrogenase type 1 (11 β -HSD1) to 7 β -hydroxy-DHEA through an oxido-reductive process via 7-oxo-DHEA (Muller et al, 2006b). Through this mechanism, 7 α -hydroxy-DHEA may inhibit 11 β -HSD1-dependent production of inactive cortisone from active cortisol (Hennebert et al, 2007a).

Epiandrosterone (EpiA), which is derived from DHEA, is also converted into 7-hydroxylated metabolites (Kim et al, 2004). We have recently shown that 11 β -HSD1 is responsible for converting 7 α -hydroxy-EpiA into 7 β -hydroxy-EpiA (Chalbot & Morfin, 2005; Hennebert et al, 2007b), which is readily produced in small quantities in human tissues (Jacolot et al, 1981; Kim et al, 2004).

7β-Hydroxy-EpiA has been shown to exert neuroprotective effects *in vivo* at remarkably low doses in animal models of cerebral ischemia (Pringle et al, 2003) and Alzheimer's disease (Dudas et al, 2004). However, dose levels needed to demonstrate these effects are far below the concentrations necessary to inhibit the 11β-HSD1-dependent-cortisol transformation indicating that other mechanisms may be involved.

Finally, studies in rheumatoid arthritis and brain injury models have shown that DHEA can modulate the expression of COX-2 mRNA or PGE₂ synthesis (Malik et al, 2003; Sun et al, 2006).

Taken together, these finding suggest that certain 7-hydroxysteroids may play a role in the modulation of PGs production in inflammation. This role may permit to resolve inflammation in inflammatory bowel diseases.

2. The resolution of inflammation

Inflammatory resolution occurs after the sharp PGE₂-driven inflammation process and is thought to result from cytoprotective PGD₂ and 15d-PGJ₂ action. A shift from arachidonic acid-derived PGE₂ to PGD₂ production occurs to these ends (Haworth & Buckley, 2007; Rajakariar et al, 2007) and is illustrated in Figure 1.

Figure 1 outlines the native inflammation process and its relations to immunity onset and circulating steroid metabolism. Inflammation-triggered increase in cellular cytochrome P450-7B1 which results in augmented 7α-hydroxy-steroid production was shown both in a

Fig. 1. At the onset of inflammatory response, arachidonic acid (AA) is converted to PGH₂ via COX. In turn, PGE₂, PGI₂ and PGD₂ are produced from PGH₂ by synthases (PGE₂S, PGI₂S, PGD₂S). PGD₂ is transformed non-enzymatically (Sp) to 15d-PGJ₂ which interacts with PPAR-γ. Non PPAR-γ-mediated action of 15d-PGJ₂ are not shown.

mouse model of rheumatoid arthritis and in humans with rheumatoid arthritis (Dulos et al, 2004; Dulos et al, 2005). Due to this increase, competitive inhibition of the cellular 11 β -HSD1 may occur (Muller et al, 2006a) through use of the enzyme for transformation of 7 α -hydroxysteroid into 7 β -hydroxysteroids (Hennebert et al, 2007a; Muller et al, 2006b). Thus, the circulating inactive cortisone made available to the inflamed cells is not activated into cortisol which would quench the onset of immune processes. Our finding of a 7 α -hydroxy-dehydroepiandrosterone-triggered increase of immune response in mice supports this relation to immunity onset (Morfin & Courchay, 1994).

These basic facts led to questions relative to the effect of 7 β -hydroxysteroids on inflammation, PG metabolism, cell protection and related mechanisms of action. Answers were found through investigations using several models of inflammation in rats and humans.

3. Choice in the 7 β -Hydroxysteroid used in treatments

Among 7 β -hydroxysteroids, 7 β -hydroxy-epiandrosterone (7 β -hydroxy-EpiA) was selected because of its reported effects as a neuroprotector (Pringle et al, 2003). 7 β -Hydroxy-EpiA is a native steroid which derives from testosterone and dehydroepiandrosterone as reported (Niro et al, 2010) and illustrated in Figure 2. The chemical synthesis of 7 β -hydroxy-EpiA was carried out (Ricco et al, 2011) and provided 400 mg of the steroid which is available now for investigations in our group and abroad. Several doses of 7 β -hydroxy-EpiA (0.01, 0.1 and 1 mg/kg) were used for the treatments.

Fig. 2. Epiandrosterone derives from testosterone and dehydroepiandrosterone (DHEA) and is a substrate for cytochrome P450-7B1 (2 EC 1.14.13.100) producing 7 α -hydroxy-epiandrosterone which is inter-converted then into 7 β -hydroxy-epiandrosterone by 11 β -hydroxysteroid dehydrogenase type 1 (3 EC 1.1.1.148). Other enzymes are 1. 3 β -hydroxy-5-ene steroid dehydrogenase (EC 1.1.1.145, EC 5.3.3.1) 4. 17 β -hydroxysteroid dehydrogenase (EC 1.1.1.51) 5. Steroid Δ 4-5 α -reductase (EC 1.3.99.5) 6. 3 β -hydroxysteroid dehydrogenase (EC 1.1.1.51).

4. Investigations in rats with dextran sodium sulphate-induced colitis

4.1 Colitis induction and experimental design

Male Wistar rats (180-200g) were acclimated for 7 days and were divided into 2 controls groups (one sham-treated and one treated) and 2 colitis groups (one sham-treated and one treated). Treatments with 7β-hydroxy-EpiA were carried out for 7 days prior to colitis induction (Figure 3). Doses (0.01, 0.1 and 1 mg/kg) were dissolved in dimethyl sulfoxide and injected *i.p.* daily for 7 days. Dimethyl sulfoxide alone was injected to the sham-treated animals. Colitis was then induced by addition of 5% dextran sodium sulphate (DSS) to drinking water for 7 days. Plain water was given to the control groups. Clinical and histological signs of colitis occurred in all the DSS-treated rats. Typically, all the rats exhibited symptoms of colitis and severe diarrhea 5 days after the onset of DSS treatment followed by rectal bleeding and shortening of the colon after 7 days. Increase in colonic myeloperoxidase activity indicated an invasion of the colonic mucosa by neutrophils (Hennebert et al, 2008).

Fig. 3. Experimental design. DSS: 5% DSS in drinking water; DMSO: dimethyl sufoxide; 7β-OH-EpiA: 7β-hydroxy-EpiA in DMSO.

4.2 Histological examinations

A portion (1 cm) of the proximal colon was fixed in 4% formaldehyde and embedded in paraffin. Tissue sections (5 μm) were stained either with hematoxylin/eosin or Alcian blue for evaluation of colonic damage and mucus goblet cells content, respectively.

Major hallmarks of colonic inflammation, namely cryptic distorsion, neutrophil infiltration in the mucosal tissue (Figure 4B1), and loss of goblet cells which contained less mucins

(Figure 4B2) were apparent in the colitis group at day 13 and were more pronounced at day 14 (Figure 4D1-4D2) when compared with sham-control group (Figure 4A1,A2 and C1,C2).

The two low doses of 7 β -hydroxy-EpiA (0.01, 0.1 mg/kg) prevented the DSS-induced colonic damages as indicated by the suppression of diarrhea and rectal bleeding. The colon length reduction was less pronounced in the groups treated by the low steroid doses (0.01, 0.1 mg/kg) than in the colitis group. All 7 β -hydroxy-EpiA doses (0.01, 0.1 and 1 mg/kg) antagonized mucus depletion in goblet cells (Figure 4E2,F2,G2) and improved histological changes such as the abnormality of crypts and neutrophil infiltration (Figure 4E1,F1,G1) (Hennebert et al, 2008).

Fig. 4. Acute colitis induced by DSS: effect of 7 β -hydroxy-EpiA on colon injury. Histological appearance of rat colonic mucosa after hematoxylin/eosin (A1-G1) or alcian blue stain (A2-G2) at days 13 and 14: sham (A1,A2) and DSS-treated (B1,B2) at day 13; sham (C1,C2) and DSS-treated (D1,D2) at day 14; and 0.01 mg/kg 7 β -hydroxy-EpiA (E1,E2), 0.1 mg/kg (F1,F2) and 1 mg/kg (G1,G2) at day 14. No histological modification was present in sham animals (A and C) at days 13 and 14. Mucosal injury in DSS-induced colitis rats starting at day 13 and being more pronounced at day 14 (D) was characterized by necrosis of the epithelium, focal ulceration, infiltration of inflammatory cells and mucin goblet cell depletion. Treatment with 7 β -hydroxy-EpiA (E-G) reduced the morphological alteration associated with DSS administration protecting the mucosal architecture. Magnification was 40x for all slices.

4.3 7 β -Hydroxy-EpiA-induced changes in PG colonic tissue levels

7 β -Hydroxy-EpiA treatments did not alter PGE₂ and PGD₂ colonic tissue levels in control rats without colitis (data not shown). DSS administration resulted into a marked increase of PGE₂, PGD₂ and 15d-PGJ₂ colonic synthesis at day 13 (Figure 5). At day 14, PGE₂ levels whereas PGD₂ synthesis was increased and 15d-PGJ₂ levels were reduced (Figure 5).

7 β -Hydroxy-EpiA treatment in rats with DSS-induced colitis significantly decreased the colonic PGE₂ synthesis at days 13 and 14 when compared with untreated colitis rats. The greatest decrease was observed with the lowest dose (0.01 mg/kg) which led to a return towards basal values (Figure 5A). PGD₂ levels remained unchanged with the highest steroid-doses whereas the low doses (0.01 and 0.1mg/kg) caused a small but significant decrease (Figure 5B). Following the treatment with 7 β -hydroxy-EpiA and throughout colitis

induction, 15d-PGJ₂ levels were maintained above control levels ranging from 5 to 14-fold increase (Figure 5C). Since 15d-PGJ₂ results from the produced PGD₂, addition of both PG levels may reflect the total production of PGD₂. This leads to observations identical with those derived from 15d-PGJ₂ measurements alone (Figure 4D). The decrease in 15d-PGJ₂ observed at the end of colitis induction underlies that 7β-hydroxy-EpiA treatment produced large amounts of 15d-PGJ₂ prior to colitis induction. This was confirmed after measurement of colonic 15d-PGJ₂ during 7β-hydroxy-EpiA treatment (Figure 5). Thus, a 51-fold increase was obtained with the 0.1 mg/kg dose at day 2 (D2) and decreased progressively from D4 to D6 (increments ranging from of 44-fold to 5-fold) (Figures 5C and 6). Conclusion of these findings is that 7β-hydroxy-EpiA treatments induced a shift from PGE₂ to 15d-PGJ₂ production (Hennebert et al, 2008).

Fig. 5. Effect of 7β-hydroxy-EpiA on colonic synthesis of PGE₂ (A), PGD₂ (B), 15d-PGJ₂ (C) and PGD₂+15d-PGJ₂ (D). Measurements were carried out in triplicate during DSS administration at days D11, D13 and D14. Data are expressed as mean ± S.E.M. with n=3-15 (*) Dextran sodium sulphate-induced colitis group versus Sham-control group; (α) 7β-hydroxy-EpiA treated group versus Dextran sodium sulphate-induced colitis group (*p* < 0.05).

4.4 7β-Hydroxy-EpiA-induced changes in PG-producing enzymes

Because significant modifications in prostaglandin levels were observed with DSS administration and 7β-hydroxy-EpiA treatments, we tested whether the expression of COX-2, mPGES-1 and H-PGDS was altered by quantifying specific mRNA by real-time RT-PCR. Transcription of these genes was examined and related to that of the HPRT1 house-keeping gene (Hennebert et al, 2008).

Fig. 6. Colonic 15d-PGJ₂ levels at various times during and after 7 β -hydroxy-EpiA treatment (D2-D9). Values are mean $\pm$ S.E.M. (n= 3-15) . (*) comparison to the sham-control group ($p<0.05$).

In control rats without colitis, the daily 7 β -hydroxy-EpiA treatment (0.1 mg/kg) induced a significant 1.5 fold increase in COX-2 mRNA levels at 15h followed by a decrease up to day 4 and thereafter returned to basal values (Figure 7A). mPGES-1 mRNA expression increased transiently between 6h to 15h returning towards basal levels at day 2. H-PGDS mRNA expression remained similar to that in sham-control rats throughout the time course.

After colitis induction, a 2.5 fold increase in COX-2 mRNA expression was observed at days 13 and 14 of DSS administration (Figure 7B) while mPGES-1 mRNA was significantly augmented at day 13 only (Figure 7C). H-PGDS was not altered by colitis (Figure 7D). In conclusion, 7 β -hydroxy-EpiA treatment (all doses) suppressed the increase in both COX-2 and mPGES-1 mRNA synthesis in the colitis group from day 13 to day 14 (Figure 7).

4.5 7 β -Hydroxy-EpiA overall effects on colitis

Colitis was fully induced in untreated rats after DSS administration for 7 days. Treatment for 7 days with several doses of 7 β -hydroxy-EpiA prior to colitis induction led to a prevention of colitis onset. Examination of myeloperoxidase activity and oxidative stress markers such as carbonylated proteins and Tbars provided evidence of their decrease after steroid treatment (Hennebert et al, 2008). The preventive effects of 7 β -hydroxy-EpiA could result in part from the early increase of the prostaglandin 15d-PGJ₂ that might contribute to down regulate the inflammatory response. 7 β -Hydroxy-EpiA triggered an increase in COX-2 and m-PGES1 expression but depressed the elevation of colonic PGE₂ synthesis in dextran sodium sulphate-treated rats. Thus, 7 β -hydroxy-EpiA triggered a shift from the PGE pathway towards the PGD pathway, causing increased production of 15d-PGJ₂, the dehydrated stable metabolite of PGD₂ (Scher & Pillinger, 2005). Our finding that small doses of 7 β -hydroxy-EpiA, an endogenous steroid produced in colon, can prevent colonic damage through a shift from PGE₂ to PGD₂ production via changes in COX-2 expression, followed by an increase of 15d-PGJ₂ levels in the colon result from investigations carried out on a rat

Fig. 7. Colonic expression of COX-2, m-PGES1 and H-PGDS mRNA after 0.1 mg/kg 7β-hydroxy-EpiA treatment and before colitis induction (A). During colitis induction, mRNA levels for COX-2 (B), m-PGES-1 (C) and H-PGDS (C) were measured at days 11, 13 and 14. mRNA levels are expressed relative to that of HPRT1 standing at a basal level of 1 shown as a continuous line. (*) DSS-colitis versus basal level ($p < 0.05$). (**) DSS + 0.01, 0.1 and 1 mg/kg 7β-hydroxy-EpiA-treated group versus DSS-colitis group ($p < 0.05$).

model where inflammation and colitis were obtained by dextran sodium sulphate administration. Two immediate questions arise from these findings: i) does this process, leading to the resolution of inflammation, occur in any inflammation? ii) is 7β-hydroxy-EpiA as efficient in humans as in rats when faced with inflammation? Answers to these questions were seek in human cell cultures.

5. A model for inflammation studies in humans

Addition of TNF-α to cultures of human peripheral blood monocytes (PBMC) results into the cellular stress found in inflammatory conditions through production of mediators released from the eicosanoid pathway. These mediators include PGE₂ that is associated with inflammation, and PGD₂ and 15d-PGJ₂ which are associated with the resolution of inflammation (Haworth & Buckley, 2007). Therefore, we used human PBMC cultured in the presence or absence of 0.01 μg/mL TNF-α in order to test the effects of 7β-hydroxy-EpiA on PG production (Le Mee et al, 2008). PBMC were cultivated either for 4 or 24 hours. RIA measurements were carried out for the PG levels released in the medium, and mRNA of the eicosanoid pathway were measured in cell extracts by quantitative PCR. All mRNA measurements were made relative to the expression of house-keeping gene HPRT1 (Le Mee et al, 2008).

5.1 7β-Hydroxy-EpiA-induced a shift in PG production

7β-Hydroxy-EpiA addition to the medium induced no significant change in PG levels in PBMC cultured for 4 h. In marked contrast, significantly increased 15d-PGJ₂ levels were obtained after cultivation for 24 h (Figure 8). This increment occurred both in the presence and in the absence of TNF-α. A significant decrease in PGE₂ levels was also obtained in the presence of TNF-α only (Figure 8B). These findings indicated that 7β-hydroxy-EpiA treatment of TNF-α-associated inflammation patterns were causative of a shift from PGE₂ to 15d-PGJ₂ production. Since 15d-PGJ₂ is a spontaneous dehydrated derivative of PGD₂, production of PGD₂ must have been increased along with the inflammation-related conditions for its conversion to 15d-PGJ₂ (Le Mee et al, 2008). 7β-Hydroxy-EpiA-triggered changes in the expression of the enzymes producing PGE₂ and PGD₂ had to be examined then.

Fig. 8. PG levels released in the medium of human PBMC cultured for 24 h fortified with 1-100 nM 7β-hydroxy-EpiA. Cultures were carried out in the absence (A) or in the presence of TNF-α (B). (*) significantly different from controls (*p* < 0.05).

5.2 7β-Hydroxy-EpiA-induced changes in eicosanoid pathway enzyme expression

In addition to COX-2 and m-PGES1 which are responsible for arachidonic acid conversion to PGH₂ and PGE₂, respectively, we measured the mRNA levels of the known two enzymes

responsible for PGD2 production, namely H-PGDS and L-PGDS. No significant change in enzyme mRNA levels was found in cells culture with 7β-hydroxy-EpiA for 4 h. In marked contrast, cultivation for 24 h led to significant differences in enzyme expression. Cultures without addition of TNF-α showed that 7β-hydroxy-EpiA significantly increased m-PGES1 and decreased H-PGDS mRNA levels (Figure 9A). The findings were opposite in the presence of TNF-α, with significant decrease in m-PGES1 and no change in H-PGDS mRNA levels (Figure 9B). No significant effect of 7β-hydroxy-EpiA was found for COX-2 and L-PGDS. These findings parallel well with PG levels measurements, and imply that the 7β-hydroxy-EpiA-triggered shift from PGE₂ to 15d-PGJ₂ production is mainly due to a lower expression of the m-PGES1 enzyme responsible for PGE₂ production (Le Mee et al, 2008).

Fig. 9. Measurement of COX-2, m-PGES1, H-PGDS and L-PGDS mRNA levels relative to the expression of house-keeping gene HPRT1. The human PBMC were cultured for 24 h without or with 1-100 nM 7β-hydroxy-EpiA. Cultures were carried out in the absence (A) or the presence of TNF-α. (*) significantly different from control group (*p* < 0.05).

5.3 A putative specific receptor for 7β-Hydroxy-EpiA

Our findings that a 10⁻⁹ M dose of 7β-hydroxy-EpiA was a trigger for the resolution of inflammation and for cellular protection led us to consider this steroid as a possible candidate for interference with a putative nuclear receptor. A precursor for 7β-hydroxy-

EpiA, namely 5 α -androstane-3 β ,17 β -diol, was shown to bind preferentially to the estrogen receptor- β (ER- β) with agonistic activity and a K_d of 2 nM (20 times larger than for estradiol) (Kuiper et al, 1997). Several reports inferred that ER- β was involved in the protection against breast and prostate cancer (Dondi et al, 2010; Nilsson et al, 2001), and this led us to test this hypothesis on three breast cancer cell lines, namely, MCF-7, MDA-231 and SKBR-3 (Niro et al, 2011).

6. Tamoxifen-like effect of 7 β -Hydroxy-EpiA on breast cancer cell proliferation

Among ERs, MCF-7 cells are known to express ER- α , ER- β and GPR30. MCF-7 cells proliferate in the presence of 10⁻⁸ M estradiol, and their proliferation is markedly inhibited by 10⁻⁶ M tamoxifen (TAM), even in the presence of estradiol. Cultivations in the presence of 10⁻⁷-10⁻⁹ M 7 β -hydroxy-EpiA without TAM or estradiol inhibited MCF-7 cell proliferation to TAM levels, whereas use of both TAM and 7 β -hydroxy-EpiA, or estradiol + 7 β -hydroxy-EpiA, decreased the proliferation below TAM levels (Figure 10A). These results indicated that 7 β -hydroxy-EpiA doses 10-10³ lower than the TAM dose produced TAM-like effects on MCF-7 growth. In contrast, the MDA-231 cell receptor status is ER- β +, GPR30+. Their proliferation was not increased by estradiol, and TAM effects were not as strikingly important as in MCF-7 cells (Niro et al, 2011).

Cultivations of the MDA-231 cells in the presence of 10⁻⁷-10⁻⁹ M 7 β -hydroxy-EpiA without TAM or estradiol did not significantly change the proliferation patterns, whereas use of either estradiol + TAM or estradiol + 7 β -hydroxy-EpiA, decreased the proliferation much below TAM or estradiol levels (Figure 10B). In the case of SKBR-3 cells, the receptor status is GPR30+. SKBR-3 cell line proliferation was not significantly changed by estradiol, TAM or 10⁻⁷-10⁻⁹ M 7 β -hydroxy-EpiA. Cultivations of SKBR-3 cells in the presence of estradiol + 7 β -hydroxy-EpiA significantly decreased cell proliferation (Figure 10C) (Niro et al, 2011).

The conclusions drawn from these studies imply the native 7 β -hydroxy-EpiA in the regulation of ER-mediated estrogen effects. As for TAM, the responsiveness of MCF-7 cells to 7 β -hydroxy-EpiA was better than for MDA-231 and SKBR-3 cells, and this implied that the antiproliferative and cytoprotective effects can be mediated by a receptor linked to one or several ERs. In any case, the receptor responsible for 7 β -hydroxy-EpiA effects needs to be identified and whether and how ERs are involved in the mechanism leading to cell protection require further examination.

7. Pathophysiological significance of 7 β -Hydroxy-EpiA levels

The first requirement for the production of native 7 β -hydroxy-EpiA is the presence of the steroid hormone substrate testosterone for generation of epiandrosterone by the classical steroid metabolism (Figure 2). Scarce studies were addressed to the levels of circulating epiandrosterone in humans. This may be due to ignorance of its fate in hormonal and physiological processes. Two key enzymes are required then for 7 β -hydroxy-EpiA production. One is CYP7B1 which provides a hydroxyl at the 7 α - position on epiandrosterone, and the second is 11 β -HSD1 which converts 7 α -hydroxy-EpiA into 7 β -hydroxy-EpiA. Both enzymes

Fig. 10. Tamoxifen-like effects of 7β-hydroxy-EpiA on the proliferation of MCF-7 (A), MDA-231 (B) and SKBR-3 cell lines. Cells (10⁵) were seeded in triplicate in 24-well plates, and adherent cells were cultivated in 1 mL growth medium at 37°C, 95% humidity and in the presence of 5% CO₂ for 72h. Estradiol (E2), 7β-hydroxy-EpiA (7β-EpiA) and tamoxifen (TAM) were administered in 5μL ethanol at the beginning of assays. Controls contained 5μL ethanol only. * Indicates a significant difference with controls (*p* < 0.05).

are ubiquitous in human tissues, and any deficit in their expression may lead to related pathologies. Their outcome may also be related to aging since the production and levels of steroid precursors such as DHEA and testosterone are known to be significantly decreased after the fifth decade in humans. These considerations would be strongly supported by precise measurements of the 7 β -hydroxy-EpiA circulating levels in humans.

Finally, variations in the susceptibility for the development of diseases may depend on 7 β -hydroxy-EpiA receptor availability and function (Nilsson & Gustafsson, 2002). The precise mechanism of action and the ERs involvement are to be deciphered in order to give a full support to 7 β -hydroxy-EpiA beneficial effects.

8. Concluding remarks

DHEA and testosterone-derived metabolites, namely epiandrosterone and 5 α -androstane-3 β ,17 β -diol, are substrates for the CYP7B1 and their 7 α -hydroxylated products are also converted into 7 β epimer by the 11 β -HSD1. When assayed in rat colitis and inflamed human PBMC at doses 10⁴ lower than circulating free DHEA or testosterone, 7 β -hydroxy-EpiA was shown to shift the PG metabolism from PGE₂ to 15d-PGJ₂ production, thus triggering the resolution of inflammation. In addition, 7 β -hydroxy-EpiA (1 nM) exerted the same effects as TAM (1 μ M) on the proliferation of MCF-7, MDA-231 and SKBR-3 human breast cancer cell lines. These findings suggest that the observed effects of 7 β -hydroxy-EpiA could be mediated in part by ERs.

This overview of recent studies implies that DHEA and/or testosterone are precursors for 7 β -hydroxy-EpiA acting on inflamed tissues for the resolution of inflammation through a putative receptor. CYP7B1 and 11 β -HSD1 are two key enzymes involved in 7 β -hydroxy-EpiA production, and their expression in colitis and other inflammatory diseases could be a rewarding target in further studies.

9. References

- Ajuebor, Singh, & Wallace (2000). Cyclooxygenase-2-derived prostaglandin D(2) is an early anti-inflammatory signal in experimental colitis. *Am. J. Physiol Gastrointest. Liver Physiol.* 279, G238-G244.
- Blumberg, Saubermann, & Strober (1999). Animal models of mucosal inflammation and their relation to human inflammatory bowel disease. *Curr. Opin. Immunol.* 11, 648-656.
- Chalbot & Morfin (2005). Neurosteroids: metabolism in human intestine microsomes. *Steroids* 70, 319-326.
- Dondi, Piccolella, Biserni, Della, Ramachandran, Locatelli, Rusmini, Sau, Caruso, Maggi, Ciana, & Poletti (2010). Estrogen receptor beta and the progression of prostate cancer: role of 5 α -androstane-3 β ,17 β -diol. *Endocr. Relat Cancer* 17, 731-742.
- Doostzadeh & Morfin (1996). Studies of the enzyme complex responsible for pregnenolone and dehydroepiandrosterone 7 α -hydroxylation in mouse tissues. *Steroids* 61, 613-620.

- Dubuquoy, Rousseaux, Thuru, Peyrin-Biroulet, Romano, Chavatte, Chamaillard, & Desreumaux (2006). PPARgamma as a new therapeutic target in inflammatory bowel diseases. *Gut* 55, 1341-1349.
- Dudas, Hanin, Rose, & Wulfert (2004). Protection against inflammatory neurodegeneration and glial cell death by 7beta-hydroxy epiandrosterone, a novel neurosteroid. *Neurobiol. Dis.* 15, 262-268.
- Dulos, van der Vleuten, Kavelaars, Heijnen, & Boots (2005). CYP7B expression and activity in fibroblast-like synoviocytes from patients with rheumatoid arthritis: regulation by proinflammatory cytokines. *Arthritis Rheum.* 52, 770-778.
- Dulos, Verbraak, Bagchus, Boots, & Kaptein (2004). Severity of murine collagen-induced arthritis correlates with increased CYP7B activity: enhancement of dehydroepiandrosterone metabolism by interleukin-1beta. *Arthritis Rheum.* 50, 3346-3353.
- Funk (2001). Prostaglandins and leukotrienes: advances in eicosanoid biology. *Science* 294, 1871-1875.
- Haworth & Buckley (2007). Resolving the problem of persistence in the switch from acute to chronic inflammation. *Proc. Natl. Acad. Sci. U. S. A.* 104, 20647-20648.
- Hennebert, Chalbot, Alran, & Morfin (2007a). Dehydroepiandrosterone 7alpha-hydroxylation in human tissues: possible interference with type 1 11beta-hydroxysteroid dehydrogenase-mediated processes. *J. Steroid Biochem. Mol. Biol.* 104, 326-333.
- Hennebert, Pelissier, Le Mee, Wulfert, & Morfin (2008). Anti-inflammatory effects and changes in prostaglandin patterns induced by 7beta-hydroxy-epiandrosterone in rats with colitis. *J. Steroid Biochem. Mol. Biol.* 110, 255-262.
- Hennebert, Pernelle, Ferroud, & Morfin (2007b). 7alpha- and 7beta-hydroxy-epiandrosterone as substrates and inhibitors for the human 11beta-hydroxysteroid dehydrogenase type 1. *J. Steroid Biochem. Mol. Biol.* 105, 159-165.
- Jacolot, Berthou, Dreano, Bercovici, & Floch (1981). In vivo metabolism of ¹⁴C-labelled 5alpha-androstane-3β,17β-diol. *J. Steroid Biochem.* 14, 663-669.
- Kim, Chalbot, Pompon, Jo, & Morfin (2004). The human cytochrome P4507B1: catalytic activity studies. *J. Steroid Biochem. Mol. Biol.* 92, 383-389.
- Kuiper, Carlsson, Grandien, Enmark, Haggblad, Nilsson, & Gustafsson (1997). Comparison of the ligand binding specificity and transcript tissue distribution of estrogen receptors alpha and beta. *Endocrinology* 138, 863-870.
- Le Mee, Hennebert, Ferrec, Wulfert, & Morfin (2008). 7beta-Hydroxy-epiandrosterone-mediated regulation of the prostaglandin synthesis pathway in human peripheral blood monocytes. *Steroids* 73, 1148-1159.
- Malik, Narayan, Wendling, Cole, Pashko, Schwartz, & Strauss (2003). A novel dehydroepiandrosterone analog improves functional recovery in a rat traumatic brain injury model. *J. Neurotrauma* 20, 463-476.
- Melgar, Drmotova, Rehnstrom, Jansson, & Michaelsson (2006). Local production of chemokines and prostaglandin E2 in the acute, chronic and recovery phase of murine experimental colitis. *Cytokine* 35, 275-283.

- Monneret, Li, Vasilescu, Rokach, & Powell (2002). 15-Deoxy-delta 12,14-prostaglandins D2 and J2 are potent activators of human eosinophils. *J. Immunol.* 168, 3563-3569.
- Morfin & Courchay (1994). Pregnenolone and dehydroepiandrosterone as precursors of native 7-hydroxylated metabolites which increase the immune response in mice. *J. Steroid Biochem. Mol. Biol.* 50, 91-100.
- Muller, Hennebert, & Morfin (2006a). The native antigluocorticoid paradigm. *J. Steroid Biochem. Mol. Biol.* 100, 95-105.
- Muller, Pompon, Urban, & Morfin (2006b). Inter-conversion of 7alpha- and 7beta-hydroxy-dehydroepiandrosterone by the human 11beta-hydroxysteroid dehydrogenase type 1. *J. Steroid Biochem. Mol. Biol.* 99, 215-222.
- Murthy, Murthy, Coppola, & Wood (1997). The efficacy of BAY y 1015 in dextran sulfate model of mouse colitis. *Inflamm. Res.* 46, 224-233.
- Nilsson & Gustafsson (2002). Biological role of estrogen and estrogen receptors. *Crit. Rev. Biochem. Molec. Biol.* 37, 1-28.
- Nilsson, Makela, Treuter, Tujague, Thomsen, Andersson, Enmark, Pettersson, Warner, & Gustafsson (2001). Mechanisms of estrogen action. *Physiol. Rev.* 81, 1535-1565.
- Niro, Hennebert, & Morfin (2010). A native steroid hormone derivative triggers the resolution of inflammation. *Hormone Molecular Biology and Clinical Investigation* 1, 11-19.
- Niro, Hennebert, & Morfin (2011). New insights into the protective effects of DHEA. *Hormone Molecular Biology and Clinical Investigation* 4, 489-498.
- Pavlick, Laroux, Fuseler, Wolf, Gray, Hoffman, & Grisham (2002). Role of reactive metabolites of oxygen and nitrogen in inflammatory bowel disease. *Free Radic. Biol. Med.* 33, 311-322.
- Pelissier, Muller, Hill, & Morfin (2006). Protection against dextran sodium sulfate-induced colitis by dehydroepiandrosterone and 7alpha-hydroxy-dehydroepiandrosterone in the rat. *Steroids* 71, 240-248.
- Pelissier, Trap, Malewiak, & Morfin (2004). Antioxidant effects of dehydroepiandrosterone and 7a-hydroxy-dehydroepiandrosterone in the rat colon, intestine and liver. *Steroids* 69, 137-144.
- Podolsky (2003). The future of IBD treatment. *J. Gastroenterol.* 38 Suppl 15, 63-66.
- Pringle, Schmidt, Deans, Wulfert, Reymann, & Sundstrom (2003). 7-Hydroxylated epiandrosterone (7-OH-EPIA) reduces ischaemia-induced neuronal damage both in vivo and in vitro. *Eur. J. Neurosci.* 18, 117-124.
- Rajakariar, Hilliard, Lawrence, Trivedi, Colville-Nash, Bellingan, Fitzgerald, Yaqoob, & Gilroy (2007). Hematopoietic prostaglandin D2 synthase controls the onset and resolution of acute inflammation through PGD2 and 15-deoxyDelta12 14 PGJ2. *Proc. Natl. Acad. Sci. U. S. A.* 104, 20979-20984.
- Ricco, Revial, Ferroud, Hennebert, & Morfin (2011). Synthesis of 7beta-hydroxy-epiandrosterone. *Steroids* 76, 28-30.
- Scher & Pillinger (2005). 15d-PGJ2: the anti-inflammatory prostaglandin? *Clin. Immunol.* 114, 100-109.
- Stenson (2007). Prostaglandins and epithelial response to injury. *Curr. Opin. Gastroenterol.* 23, 107-110.

Sun, Wu, Tsuang, Chen, & Sheu (2006). The in vitro effects of dehydroepiandrosterone on chondrocyte metabolism. *Osteoarthritis. Cartilage*. 14, 238-249.

IntechOpen

IntechOpen

Colitis

Edited by Dr Fukata

ISBN 978-953-307-799-4

Hard cover, 196 pages

Publisher InTech

Published online 05, January, 2012

Published in print edition January, 2012

Inflammation of the colon is collectively called "Colitis". Since a variety of conditions may cause colitis and its manifestations are similar among the causes, selection of the right treatment based on the correct diagnosis is important in the management of this group of illnesses. Over the last few decades, a major shift has been observed in the clinical attention to the pathogenesis of colitis from infectious to idiopathic inflammatory bowel diseases. Colitis cases that are associated with chemical therapeutics and specific pathogens such as amoeba, have become prominent in hospitalized individuals and immune deficient patients, respectively. In addition, a great deal of progress has been made in colitis research triggering the need for updating our knowledge about colitis. This book Colitis provides comprehensive information on the pathogenesis, mechanism of resolution, and treatment strategies of colitis.

How to reference

In order to correctly reference this scholarly work, feel free to copy and paste the following:

Hennebert Olivier, Pelissier Marie-Agnès and Morfin Robert (2012). Resolution of Colitis-Associated Inflammation, Colitis, Dr Fukata (Ed.), ISBN: 978-953-307-799-4, InTech, Available from: <http://www.intechopen.com/books/colitis/resolution-of-colitis-associated-inflammation>

INTech
open science | open minds

InTech Europe

University Campus STeP Ri
Slavka Krautzeka 83/A
51000 Rijeka, Croatia
Phone: +385 (51) 770 447
Fax: +385 (51) 686 166
www.intechopen.com

InTech China

Unit 405, Office Block, Hotel Equatorial Shanghai
No.65, Yan An Road (West), Shanghai, 200040, China
中国上海市延安西路65号上海国际贵都大饭店办公楼405单元
Phone: +86-21-62489820
Fax: +86-21-62489821

© 2012 The Author(s). Licensee IntechOpen. This is an open access article distributed under the terms of the [Creative Commons Attribution 3.0 License](https://creativecommons.org/licenses/by/3.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

IntechOpen

IntechOpen