

We are IntechOpen, the world's leading publisher of Open Access books Built by scientists, for scientists

6,900

Open access books available

186,000

International authors and editors

200M

Downloads

Our authors are among the

154

Countries delivered to

TOP 1%

most cited scientists

12.2%

Contributors from top 500 universities



WEB OF SCIENCE™

Selection of our books indexed in the Book Citation Index
in Web of Science™ Core Collection (BKCI)

Interested in publishing with us?
Contact book.department@intechopen.com

Numbers displayed above are based on latest data collected.
For more information visit www.intechopen.com



The Power of an Evolutionary Perspective in Studies of Endocrinology

Jing He¹, David M. Irwin^{1,2} and Ya-Ping Zhang^{1,3}

¹*State Key Laboratory of Genetic Resource and Evolution, Kunming Institute of Zoology, Chinese Academy of Sciences*

²*Department of Laboratory Medicine and Pathobiology and Banting and Best Diabetes Centre, University of Toronto*

³*Laboratory for Conservation and Utilization of Bioresource, Yunnan University*
^{1,3}*China*
²*Canada*

1. Introduction

Much of our understanding of the molecular basis of endocrinology has been the product of highly productive studies that have focused on specific molecules (e.g., hormones) and their specific immediate interacting partners. However, biomolecules are not isolated particles, but instead they are elements of highly integrated interaction networks, and specific interactions among them drive virtually all cellular functions and underlie phenotypic complexity and diversity. Many hormones, and their specific receptors and other interacting proteins, are known to be evolutionarily related, which raises intriguing questions concerning how specificity originated within these systems.

Our previous studies have illustrated that biochemical entities are developmentally and evolutionarily fluid, with capabilities to be altered both in composition and behavior. Gene birth and death are widespread phenomena in genome evolution and accounts for the great diversity of gene families involved in endocrinology. While concordance of evolutionary histories both in pattern and process of hormones, receptors and interacting proteins might be expected for integrated systems, studies have shown that the evolutionary history of receptors need not mirror that of their ligands. Simultaneous emergence, or loss, of multiple interacting partners by multiple gene duplication or gene loss is unlikely in evolution. Gene duplication is essential in the development of complex endocrinology. It is creative in producing elements that allow evolutionary tinkering and thus plays a major role in gene co-option (i.e., recruitment for novel functions) facilitating the evolution of greater biological complexity. Alternatively, if an interacting partner is lost, the retained partner may either be subsequently lost or, more interestingly, serve as raw material in evolution and become recruited into a new interaction yielding a new function. Thus, a stepwise process of elaboration through mutation and optimization ensues, adapting genes (and their encoded proteins) into the physiology of an organism.

Here we review several recent advances in our understanding of the evolution of hormone signalling pathways that illustrate the power of an evolutionary perspective. Among our

examples are the motilin and ghrelin signalling pathways where we have demonstrated that both the hormones (motilin and ghrelin) and their specific receptors descended from common ancestors through independent gene duplications. The motilin receptor originated well before the evolution of the hormone, and the motilin-motilin receptor specificity has arisen, as the result of ligand-receptor coevolution, after the hormone gene duplicated. Similarly, motilin and its specific receptor gene specifically lost on the rodent lineage followed a stepwise process. Once one of the interacting partners is lost, the retained partner may subsequently be lost, or serve as raw material in evolution and become recruited into a new function.

Given the evolutionary dynamics of the genome and the plasticity of biomolecule networks, an evolutionary perspective is necessary to understand many aspects of the molecular basis of endocrinology. An integrated evolutionary comparative strategy helps enhance our understanding of the assemblage of the complex endocrine systems, provide important clues in interaction capacity exploration, and identify the main diversification events of the endocrine systems and potential cross-talk between them through evolutionary related interacting proteins. In addition, knowledge of how elements of the endocrine systems that underlie cellular functions are evolutionarily and developmentally interact, help not only in choosing appropriate species to examine function, but also provide genetic makers to probe the emergence of specific traits and characteristics, uncovering the genetic basis that underlie the morphological and behavior changes, and thus enhance our understanding of how organisms adapt to changing environments.

2. The molecular basis of endocrine systems

Endocrinology is the branch of biology that focuses on regulatory systems involving a group of specialized chemical substances called hormones that travel through blood. New developments in endocrinology have been dominated by progress in molecular genetics. Although these investigations often relate to rare single gene disorders, they have resulted in major advances in our understanding of the cellular mechanisms of hormone action (Cegla, Tan and Bloom 2010; Hodson et al. 2010; Peter and Vallo 1998). Most endocrinology studies are medically orientated, and thus anthropocentric, with little or no comparative or evolutionary perspective, and therefore provide few insights into our comprehension of the enigmatic origin and diversification of these systems (Markov et al. 2008).

Much of our understanding of the molecular basis of endocrinology has come from highly productive studies that have focused on specific molecules and their immediate interacting partners, in fact, hormonal systems are a central part. Indeed the specificity of each hormone ligand and receptor pair is maintained in divergent species (Moyle et al. 1994), while biochemical entities are developmentally and evolutionarily fluid, with a much wider range of capabilities for alteration in both composition and behavior (Avisé and Ayala 2007; Wilkins 2007). Thus intriguing questions concerning how the diversity and specificity occur within these systems remain to be answered.

Genomes are documents of life history, and their structures continually change throughout evolution. Humans represent only a leaf on the tree of life. The anthropocentrism view of evolution, where humans are the pinnacle of gradually developed complexity, is a one-sided and incorrect view (Markov et al. 2008). In the light of evolution, an approach that considers each taxa equally, adopts a comparative strategy, integrates information from diverse organisms and various scientific approaches, helps enhance our understanding of

the assemblage of the complex endocrine systems and the main events that have prompted their diversification, and yields better insight into their biological functions and potential cross-talk between them through evolutionary related interacting proteins.

3. Evolutionary mechanisms for the diversification of endocrine systems

Genomes are the entirety of organisms' hereditary information, and encode all the information necessary to give rise to biomolecular products. The structure and content of genomes do not remain static, and continually change through evolutionary time. Major evolutionary mechanisms that have been instrumental in shaping the genome include gene duplication, gene loss and evolutionary shift.

3.1 Gene duplication

As genome size increases, gene content tends to increase, although at a disproportionately lower rate in eukaryotes compared to non-eukaryotes (Gregory 2005; Hou and Lin 2009; Konstantinidis and Tiedje 2004; Lynch and Conery 2003). The gene number increases with genome size, and morphological complexity is mostly generated by expanding the sizes of gene families rather than due to a growth of the number of unique gene types, hence multicellular organisms employ large sets of similar gene products while exhibiting extraordinary biodiversity. The elements of endocrine systems often group into families (e.g., hormones and their specific receptors), whose members have diverged to various extents in regulation and function (Danks et al. 2011; Hoffmann and Opazo 2011; Irwin 2010; Kim et al. 2011; Sundström, Dreborg and Larhammar 2010).

Gene duplication is the most important mechanism for generating new genes and new biochemical processes and has facilitated the evolution of biodiversity and complexity (Li 2006; Ohno 1970). The most obvious contribution is supplying the raw genetic material for the various evolutionary forces (e.g., mutation, genetic drift and natural selection) to act upon, which lead to specialized or new gene functions (Zhang 2003). Without gene duplication, the capability and plasticity of organisms in adapting to changing environments would be severely limited. Gene duplication can also contribute to species divergence (Ting et al. 2004) along with the origin of species-specific features (Zhang et al. 2002). Duplication may involve part of gene, a single gene, part of a chromosome, an entire chromosome, or the whole genome. The first four scenarios are also known as regional duplications, because they do not result in a doubling of the entire genome. Hereinafter, the role of whole genome duplication and regional duplication will be discussed respectively.

3.1.1 Whole genome duplication

Genome duplication is often considered to be more important than regional duplication in evolution, as it allows for the duplication of the entire regulatory systems. Regional duplications, on the other hand, generally allow for part of a regulatory system to be duplicated, which may lead to imbalances that may disrupt normal function. The significance of whole genome duplication has been highlighted by the studies of invertebrate chordates and the base of vertebrate evolution (Garcia-Fernández and Holland 1994), as well as fish diversification (Dehal and Boore 2005; Jaillon et al. 2004). Genome duplication facilitated the appearance and diversification of complex features, such as the endocrine system (Holland et al. 2008). The amphioxus genome exhibits considerable

synteny with the human genome, but lacks the whole-genome duplications characteristic of vertebrates (Dehal and Boore 2005). Holland et al. (2008) examined the existence of endocrine components based on the amphioxus genome, and reasoned that ancestral chordate only possess a basic set of endocrine functions. Hereby a fully functional endocrine system must have arisen after the divergence of cephalochordate, which was driven in all probability by subsequent genome duplications.

3.1.2 Regional duplication

While whole genome duplication events are not uncommon, nevertheless they only infrequently contribute to the evolution of well-developed bisexual organisms, as they likely disrupt the mechanism of sex determination and would be quickly eliminated (Li 1997). Regional duplications make up the gap, providing new genetic material for local elaboration and optimization, and fuel the evolution of lineage-specific variability (Li 1997; Ohno 1970). Whole genome duplications and small-scale duplications have very different consequences. Selective retention of different duplicates, and enrichment of signaling proteins and transcription factors, have been observed in yeast, plants, early vertebrate and fish following whole genome duplications (Conant 2010; Gout, Duret and Kahn 2009; Huminiecki and Heldin 2010; Kassahn et al. 2009; Manning and Scheeff 2010). This indicates that the individual duplication of signaling proteins and transcriptional regulators may be deleterious, since interactions between them are relatively transient and subtle, requiring a dosage balance from the whole genome duplication to survive. It is reasonable to expect that, simultaneous duplications of ligands and downstream signaling genes are required to allow the expansion of the complex endocrine systems. As it is, only part of the regulatory system has been duplicated in regional duplication, then, what are the evolutionary consequences of such an imbalanced outcome?

3.2 Gene loss

Studies on genome evolution have focused on the creation of new genes, including changes in regulatory mechanisms, and often neglect the role of selective gene loss in shaping these genomes. Gene loss or pseudogenization is a widespread phenomenon in genome evolution (Wang, Grus and Zhang 2006). The differential fixation of mutational gene loss after genome duplication illustrates the power of these types of events (Semyonov et al. 2008; van Hoek and Hogeweg 2009). Within gene families gene turnover, caused by differential gene gain and loss, leads to diverse patterns of gene distributions on different lineages, and contributes significantly to the evolution of biodiversity and may be the basis for reproductive isolation and speciation in geographically isolated populations (Gagneux and Varki 2001; Gout, Duret and Kahn 2009; Hahn, Demuth and Han 2007; Kettler et al. 2007; Powell et al. 2008). Sometimes, the ubiquitous, and near-stochastic, gene loss process can lead to the loss of single copy genes. By taking advantages of the availability of large amounts of vertebrate genomic information, it has been shown that many important human endocrine genes have been found to be missing or inactivated in other vertebrates, and vice versa (He et al. 2010; Irwin 2010; Pitel et al. 2010). In the same way, evolutionary comparisons among different taxa can help identify novel elements of endocrine systems that are not possessed by model animals. Endocrine entities are not isolated particles, but are elements of highly integrated interaction networks, and play their role through specific interactions (Carroll, Bridgham and Thornton 2008). As a random process, the simultaneous

loss of multiple interacting partners is unlikely, despite the intimate association between them. Gene loss, a dramatic genetic event, leads to the immediate loss of specific interactions, and probably affects interaction turnover as greatly as gene duplication. If a signaling protein is lost, then what are the evolutionary consequences for the retained partners?

3.3 Evolutionary shift

Genes are not only duplicated and lost, there are many genes which have been conserved and are unambiguous orthologues in a wide variety of taxonomic species, but evolutionary shifts occur frequently and orthologues genes can have distinct functions in different taxa (Macqueen et al. 2010; Zhou et al. 2008). Species adapt to diverse ecosystems and environments, and have differing genetic backgrounds. As selective constraints vary, it impacts the pattern of gene evolution, and changes in selection can yield changes in function (Irwin 2001). In some cases, positive selection appears to underlie the evolutionary shift (Wallis 2001; Liu et al. 2001); while in others, inefficient purifying selection and increased genetic drift, associated with a reduction in effective population size, are the cause (Macqueen et al. 2010).

The genetic network of the endocrine systems are developmentally and evolutionarily fictile, the elemental composition is prone to be altered via gene gain and loss, and its physiological properties frequently change through mutations in endocrine gene coding sequences and/or regulatory systems, and turnover of interacting biomolecules. Using a comparative strategy, integrating information on species phylogenetic relationships, gene evolutionary history, gene sequences and functional properties such as expression, interaction and physiological data, should enhance our understanding of how they evolved and yield better insight into their biological functions. The large amounts of accumulating genetic information is a powerful resource for addressing these questions.

4. Case studies for evolutionary endocrinology: Lessons from the motilin/ghrelin hormone family and their receptors

4.1 Gene duplication plays a major role in gene co-option

4.1.1 Ghrelin and motilin

Ghrelin and motilin represent a novel gastrointestinal hormone family in mammals (Inui 2001). Not only are ghrelin and motilin structurally related, but, the sequence and overall structure of their precursor genes show considerable similarity (Fig. 1).

Ghrelin is derived by posttranslational cleavage from its precursor preproghrelin (GHRL), and is a circulating peptide hormone that is secreted mainly by the stomach and acts upon the hypothalamus and hindbrain (Nakazato et al. 2001; Kojima and Kangawa 2005). Growth hormone secretagogue receptor (GHSR) is the specific receptor for ghrelin and a G protein-coupled receptor, and upon stimulation releases growth hormone (GH) from the pituitary (Howard et al. 1996; Kojima et al. 1999; Sun, Ahmed and Smith 2003). Evidence from mammals suggests that ghrelin also acts to stimulate gastric motility, increase appetite and food intake, and induce a positive energy balance leading to body weight gain (Murray et al. 2003; Peeters 2005). Prepromotilin (MLN) is posttranslationally processed to yield a secreted peptide that is then cleaved at a paired basic amino acid site and gives rise to motilin (Poitras 1993). Motilin primarily acts to increase gastrointestinal motility by activating neural pathways or via the direct stimulation of smooth muscles. In human and dog it has been suggested that motilin has a physiological role in the regulation of a motor pattern

typical for the fasted state (Poitras 1993). It is of interest to note that motilin also has a weak GH-releasing effect (Samson et al. 1984). GPR38, an orphan G protein coupled receptor, was identified as the motilin receptor, MLNR, through a remarkable process of reverse pharmacology (Feighner et al 1999). GHSR and MLNR, whose sequences are very similar, are members of the β -group of rhodopsin-like receptor family (Holst et al. 2004).

Despite the very close resemblance of the hormones and receptors, to date there is no evidence for any cross-reactivity between the ligands, which corresponds to the fact that the pharmacophore of the peptides are quite different (Peeters 2005). Octanoylation of serine³ is a unique and crucially important feature of ghrelin and studies have demonstrated that without the octanoyl group the potency of GHRL is dramatically decreased (Peeters 2005; Kaiya et al. 2001).

4.1.2 Evolution of the motilin/ghrelin hormone gene family

Genes for ghrelin have been cloned from a number of vertebrate species. Using bioinformatic methods, we have identified additional ghrelin gene sequences from diverse mammalian species and a frog *Xenopus tropicalis* (Table 1). Motilin genes have only been identified and characterized in mammals and birds, even after the use of bioinformatic approaches (Table 2). Ghrelin and motilin genes are both single copy genes in all of the species studied, and reside in conserved gene neighborhoods respectively, strongly supporting their orthology. The amino acid sequences of ghrelin are well conserved among species, especially in the N terminal region, and the same principle holds for motilin (Table 1-2). Interestingly, when the comparative genomic analysis was conducted between human and other vertebrates (chicken, *X. tropicalis*, medaka, tetraodon, and zebrafish) aimed at the GHRL and MLN neighborhood regions, it was revealed that homologs of the human GHRL and MLN flanking genes, which are located on different chromosomes in amniotes, were found to reside on the same chromosome near the GHRL locus in medaka, tetraodon, zebrafish, and *X. tropicalis*. This observation suggests that there was a duplication of the GHRL gene yielding MLN on the amniote lineage however there was no overlap in the genomic neighborhoods for GHRL and MLN. We could not identify any sequences similar to ghrelin and/or motilin genes in the recently released lamprey and deuterostome draft genomes. GHRL sequences from fish and amphibians possess only a single putative endoprotease recognition site C-terminal to the signal peptidase cleavage site, thus can produce only a single posttranslational-processed peptide, ghrelin. In contrast, GHRL of amniotes (reptiles, birds, and mammals) possess three putative endoprotease recognition sites, potentially giving rise to a second posttranslational-processed peptide, a 24-residue ghrelin-associated peptide (Fig. 1). The second peptide has recently been identified to be obestatin in mammals (Zhang et al. 2005). All MLNs, which are only found in reptiles, birds and mammals, possess three putative endoprotease recognition sites, thus potentially give rise to two posttranslational-processed peptides, motilin and a 17-residue peptide in a position homologous to obestatin (Fig 1). Phylogenetic analysis revealed that bullfrog GHRL groups with amniote MLN rather than amniote GHRL, although the bootstrap support for this conclusion is low (Fig. 2).

Based on the distribution of GHRL and MLN genes in the species studied, comparative genomics analysis between human and other vertebrates, and endoprotease cleavage sites distribution in GHRL/MLN genes, we surmise that MLN was generated by a gene duplication on the early amniote lineage as illustrated in figure 2 (He et al. 2011). Other potential evolutionary scenarios (e.g., duplication prior to the fish-tetrapod divergence) require a larger number of gene deletion events along with parallel gain or loss of endopeptidase cleavage sites, and thus are less parsimonious.

Species	Ghrelin sequence	Obestatin homolog sequence
<i>Homo sapiens</i>	GSSFLSPEHQRVQQRKESKKPP AKLQP	FNAPFDVGIKLSGVQYQQHS QALG
<i>Pan troglodytes</i>	GSSFLSPEHQRVQQRKESKKPP AKLQP	FNAPFDVGIKLSGVQYQQHS QALG
<i>Pongo pygmaeus</i>	GSSFLSPEHQRVQQRKESKKPP AKLQP	FNAPFDVGIKLSGVQYQQHS QALG
<i>Hylobates lar</i>	GSSFLSPEHQRVQQRKESKKPP AKLQP	FNAPFDVGIKLSGVQYQQHS QALG
<i>Macaca fuscata</i>	GSSFLSPEHQRAQQRKESKKPP AKLQP	FNAPFDVGIKLSGVQYQQHS QALG
<i>Papio hamadryas</i>	GSSFLSPEHQRAQQRKESKKPP AKLQP	FNAPFDVGIKLSGVQYQQHS QALG
<i>Saimiri sciureus</i>	GSSFLSPEHQRIQQRKESKKPPA KLQP	FNAPFDVGIKLSGVQYQQHS QALG
<i>Macaca mulatta</i>	GSSFLSPEHQRAQQRKESKKPP AKLQP	FNAPFDVGIKLSGVQYQQHS QALG
<i>Otolemur garnettii</i>	GSSFLSPDHQKIQQRKESKKPPA KLQP	FNSPLDVGIKLSGAQYQQHS QALG
<i>Cebus paella</i>	GSSFLSPEHQRMQQRKESKKPP AKLQS	FNVPDFVGIKLSGVQYQQHS QALG
<i>Aotus trivirgatus</i>	GSSFLSPEHQRIQQRKESKKPPA KLQP	FNAPFDVGIKLSGIQYQQHSQ ALG
<i>Mesocricetus auratus</i>	GSSFLSPEHQKAQQRKESKKPQ AKLQP	FNAPFDVGIKLSGAQYQQHG RALG
<i>Mus musculus</i>	GSSFLSPEHQKAQQRKESKKPP AKLQP	FNAPFDVGIKLSGAQYQQHG RALG
<i>Rattus norvegicus</i>	GSSFLSPEHQKAQQRKESKKPP AKLQP	FNAPFDVGIKLSGAQYQQHG RALG
<i>Meriones unguiculatus</i>	GSSFLSPEHQKTQQRKESKKPP AKLQP	FNAPFDVGIKLSGAQYQQHG RALG
<i>Oryctolagus cuniculus</i>	GSSFLSPEHQKAQQRKDAKKPP ARLQP	
<i>Felis catus</i>	GSSFLSPEHQKVQQRKESKKPP AKLQP	FNAPFDVGIKLSGAQYHQHG QALG
<i>Canis familiaris</i>	GSSFLSPEHQKLQQRKESKKPP AKLQP	FNAPFDVGIKLSGPQYHQHG QALG
<i>Equus caballus</i>	GSSFLSPEHHKVQHRKESKKPP AKLKP	FNAPFDVGIKLSGAQYHQHS QALG
<i>Rangifer tarandus</i>	GSSFLSPEHQKLQRKEPKKPSGR LKP	FNAPFDIGIKLSGAQSLQHGQ TLG
<i>Capra hircus</i>	GSSFLSPEHQKLQRKEPKKPSGR LKP	FNAPFNIGIKLSGAQSLQHGQ TLG
<i>Ovis aries</i>	GSSFLSPEHQKLQRKEPKKPSGR LKP	FNAPFNIGIKLSGAQSLQHGQ TLG
<i>Bos taurus</i>	GSSFLSPEHQKLQRKEAKKPSG	FNAPFNIGIKLAGAQSLQHG

Species	Ghrelin sequence	Obestatin homolog sequence
	RLKP	QTLG
<i>Bubalus bubalis</i>	GSSFLSPEHQKLQRKEPKKPSGR LKP	FNAPFNIGIKLSGAQSLQHGQ TLG
<i>Ailuropoda melanoleuca</i>	GSSFLSPEHQKVQRKESKKPPA KLQP	FNAPFDVGIKLSGAQYQEHG QALG
<i>Sus scrofa</i>	GSSFLSPEHQKVQQRKESKKPA AKLKP	FNAPCDVGIKLSGAQSDQHG QPLG
<i>Kogia breviceps</i>	GSSFLSPEHQKLQRKEAKKPSG RLKP	
<i>Myotis lucifugus</i>	GSSFLSPEHQKAQQRKESKKPP AKLQP	FNAPFDVGIKLSGAQSHWHG QALG
<i>Erinaceus europaeus</i>	GSSFLSPEHQKGQQRKEPKKPP GKVQP	FSAPFDVGLRLSGAQYEQHG EALR
<i>Dasypus novemcinctus</i>	GSSFLSPEHQKTQLRKEFKKPA KLQP	FNAPFDVGIKLSGAQYQQHG RSLG
<i>Echinops telfairi</i>	GSSFLSPGHPKVQPQRKESKTPA GKLQA	FNVPFDIGIKVSVAQYGEHGR ALD
<i>Loxodonta africana</i>	GSSFLSPKNQKLQQRKESKKPP AKLQP	
<i>Monodelphis domestica</i>	GSSFLSPEHPKTQRKETKKPSVK LQP	FNAPFDIGIKVAEAQYQQYG HALE
<i>Gallus gallus</i>	GSSFLSPTYKNIQQQKDTRKPTA RLH	FNVPFEIGVKITEREYQEYGG ALE
<i>Meleagris gallopavo</i>	GSSFLSPAYKNIQQQKDTRKPT ARLHP	FNVPFEIGVKITEREYQEYGG ALE
<i>Anas platyrhynchos</i>	GSSFLSPEFKKIQQQNDPTKTTA KIH	FHVPFEIGVKITEEEYQEYGG TLE
<i>Anser sp.</i>	GSSFLSPEFKKIQQQNDPAKAT AKIH	FNVPFEIGVKITEEEYQEYGG TLE
<i>Dromaius novaehollandiae</i>	GSSFLSPDYKKIQQRKDPRKPTT KLH	FNVPFEIGVKITEEQYQEYGG MLE
<i>Trachemys scripta elegans</i>	GSSFLSPEYQNTQQRKDPKKHT KLN	LNVPFEIGVKITEDQYQEYGG VLE
<i>Rana catesbeiana</i>	GLTFLSPADMQKIAERQSQNKL RHGNMN	
<i>Rana esculenta</i>	GLTFLSPADMRKIAERQSQNKL RHGNMN	
<i>Danio rerio</i>	GTSFLSPTQKPQGRRPPRVG	
<i>Carassius auratus</i>	GTSFLSPAQKPQGRRPPRMG	
<i>Ictalurus punctatus</i>	GSSFLSPTQKPQNRGDRKPPRV G	
<i>Oreochromis mossambicus</i>	GSSFLSPSQKPQNKVKSSRIG	

Species	Ghrelin sequence	Obestatin homolog sequence
<i>Oreochromis niloticus</i>	GSSFLSPSQKPQNKVKSSRIG	
<i>Oncorhynchus mykiss</i>	GSSFLSPSQKPQGKGKPPRVG	
<i>Acanthopagrus schlegelii</i>	GSSFLSPSQKPQNRGKSSRVG	
<i>Anguilla japonica</i>	GSSFLSPSQRPQGKDKKPPRVG	

Table 1. Bioactive peptide sequences from diverse vertebrata ghrelin gene.

Species	Motilin sequence
<i>Homo sapiens</i>	FVPIFTYGELQRMQEKERNKGQ
<i>Pan troglodytes</i>	FVPIFTYGELQRMQEKERNKGQ
<i>Macaca mulatta</i>	FVPIFTYGELQRMQEKERSKGQ
<i>Cavia porcellus</i>	FVPIFTYSELRRTQEREQNKRL
<i>Oryctolagus cuniculus</i>	FVPIFTYSELQRMQERERNRGH
<i>Felis catus</i>	FVPIFTHSELQRIREKERNKGQ
<i>Canis familiaris</i>	FVPIFTHSELQKIREKERNKGQ
<i>Ovis aries</i>	FVPIFTYGEVQRMQEKERYKGQ
<i>Bos taurus</i>	FVPIFTYGEVRRMQEKERYKGQ
<i>Sus scrofa</i>	FVPSFTYGELQRMQEKERNKGQ
<i>Equus caballus</i>	FVPIFTYSELQRMQEKERNRGQ
<i>Myotis lucifugus</i>	FVPIFTHSELQRMQEKERNKEQ
<i>Dasypus novemcinctus</i>	FVPIFTYSELQRMQEKEWNKGQ
<i>Loxodonta africana</i>	FVPIFTYSEIRRMQERERNNGQ
<i>Monodelphis domestica</i>	FVPIFTYSDVQRMQEKERNKGQ
<i>Ornithorhynchus anatinus</i>	FIPIFTHSDVQRMQERERNKGQ
<i>Gallus gallus</i>	FVPFFTQSDIQKMQEKERNKGQ

Table 2. Bioactive peptide sequences from diverse vertebrata motlin gene.

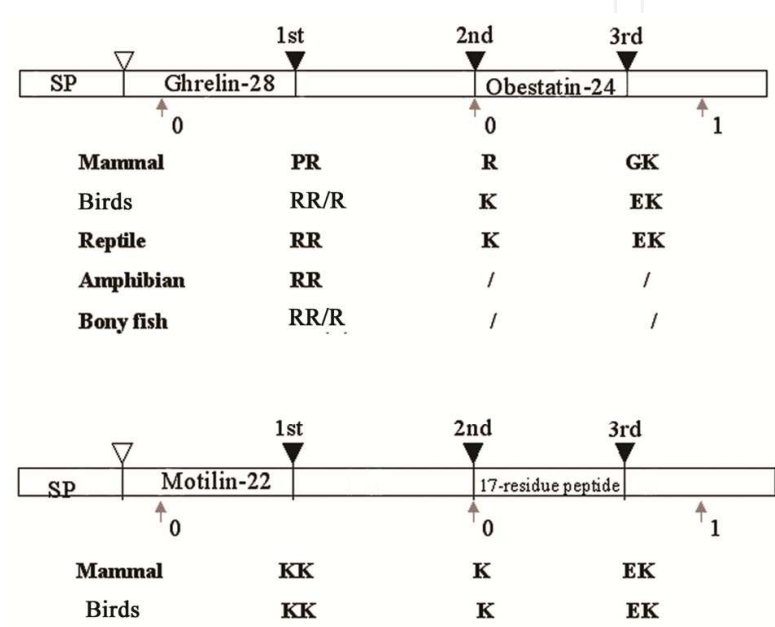


Fig. 1. Schematic representation of ghrelin and motilin preproteins. The preproteins of ghrelin and motilin are represented by boxes divided into protein domains, proportional to their length. The open and filled triangles indicate the locations of cleavage sites used by signal peptidase and proprotein convertase, respectively. The sequences of the putative endoproteinase cleavage sites of various vertebrate classes are shown below. Alternative processing sites in birds and bony fish are indicated, “↑” denotes intron position with intron phase shown beside the arrow. SP, signal peptide. “/”, lack of putative endoproteinase recognition sites.

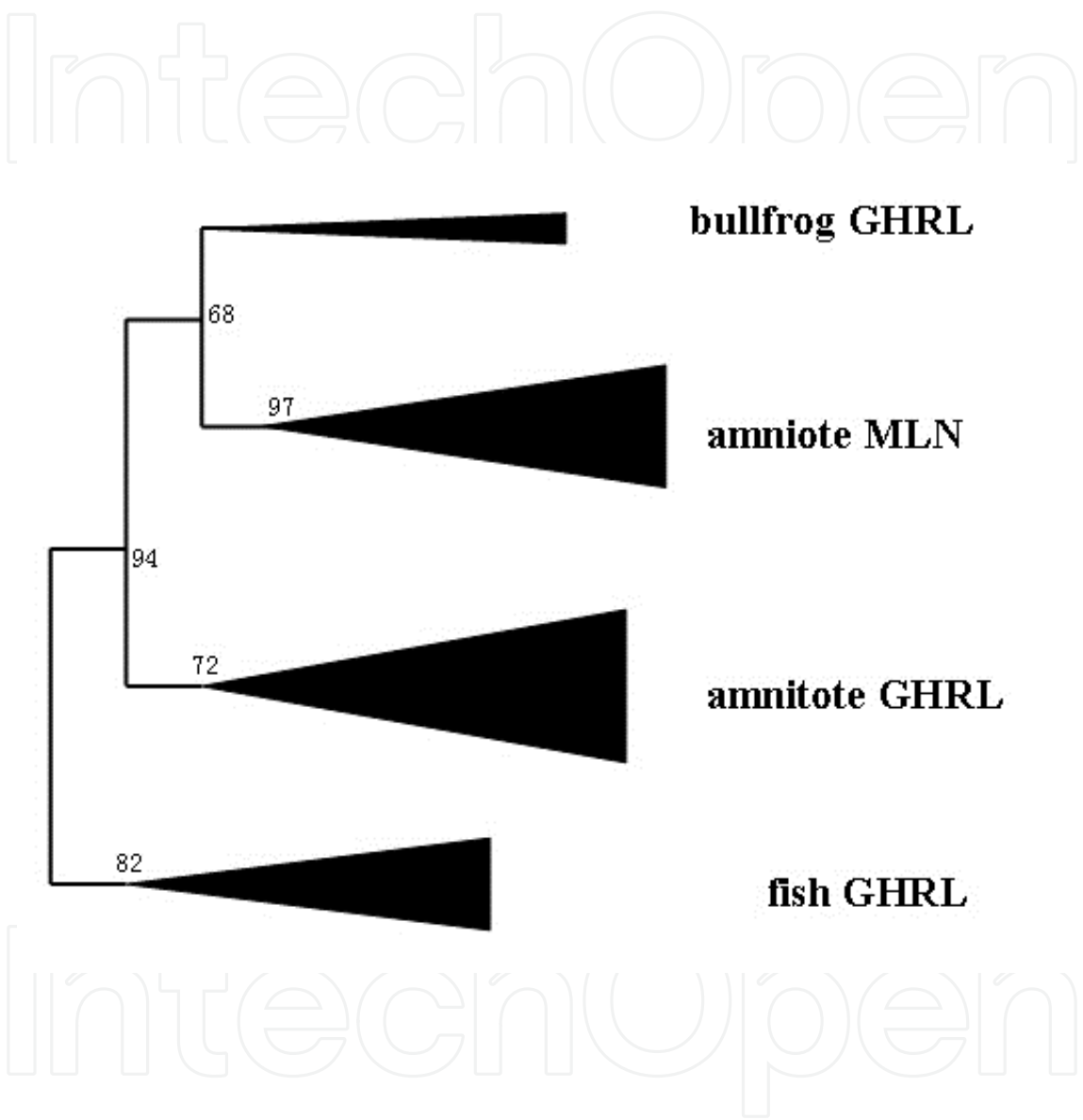


Fig. 2. Schematic representation of the motilin/ ghrelin gene family phylogenetic relationships. Bootstrap percentages are shown on interior branches. GHRL, preproghrelin. MLN, prepromotilin.

4.1.3 Evolution of ghrelin and motilin receptors

Only a small number of Ghrelin and motilin receptor genes are known and most of these are from mammals, with ortholog from only two species of fish having previously being characterized (Palyha et al. 2000; Chan and Cheng 2004). Our bioinformatic searches of diverse vertebrate genomes resulted in the identification of a great number of potential receptors. The orthology of the different receptors was established using analysis of synteny of the genes. In combination with phylogenetic reconstruction, the monophyly of each receptor type was established, and no more than one copy of each type of receptor was identified in any of the studied species. GHSR and MLNR are more closely related to each other than they are to any other characterized receptor, and the gene duplication that generated them happened more than 450 million years ago, before the divergence of ray-finned fish and tetrapods (He et al. 2011). Through comparative and evolutionary analyses, we found a new type of receptor in fish, which does not have an ortholog in any non-fish vertebrate. The function of this new receptor is unknown. The sequence of this new receptor has some peculiarities, such as possessing long extracellular loops 2 and 3, which are about 100 residues longer than the analogous loops in GHSR and MLNR (He et al. 2011). Residues at both ends of these loops have been shown to be functionally important for hormone binding and action in homologous receptors; however, the function of these residues in the loops of these novel receptors is not clear (Matsuura, Dong and Miller 2002).

Studies have shown that the GHSR orthologs in fish can be activated by growth hormone secretagogues (GHSs), while MLNR orthologs in fish failed to be activated by GHSs or mammalian motilin (Palyha et al. 2000). The identification of the ancient binding state of MLNR prior to the emergence of motilin has proven challenging, however it is reasonable to speculate that GHSR and MLNR experienced functional diversification shortly after their duplication, and the ancient MLNR did not have either ghrelin or motilin binding properties. Evolutionary studies suggest MLNR experienced an episode of rapid evolution on the branch leading to amniotes, which was driven by positive selection, and accumulated amino acid changes in ligand binding cleft. This time period of rapid evolution coincides with the date of the GHRL/ MLN gene duplication event, thus it is reasonable to speculate, that the burst of rapid evolution in MLNR was a consequence of coevolution with its new ligand, and that motilin binding specificity of MLNR only evolved as a result of ligand-receptor coevolution after the motilin gene diverged from the ghrelin gene on the amniote lineage. In contrast, GHSR has evolved under a constant selective constraint throughout vertebrates, with the ghrelin/GHSR system being maintained and functionally conserved from fish to mammals (He et al. 2011).

4.1.4 Gene duplication and gene co-option

The motilin/ghrelin hormone and their receptors were produced by independent duplication events that occurred at different points in time. The discordance of the evolutionary histories for the hormones and receptors indicate that the intimate interacting partners of an endocrine system can be produced by individual duplications, the composition and functions of each part of the endocrine network do not remain static, and that parts of a system can be co-opted for novelties, and that these processes often involve gene duplication and subsequent divergence. Structural and evolutionary relatedness allows promiscuous interaction properties, which can serve as the starting point for accommodation. Divergence of function in different species is accomplished by hormone/receptor coevolution to improve binding affinity and/or specificity. A major role

for gene co-option, involving gene duplication and divergence, should be recognized that creates potential elements which selection can act upon within a biological network to evolve new functions (He et al. 2011; True and Carroll 2002). The growth of gene families allows for more flexible gene expression and/or the evolution of new biochemical specificities (Rubenstein 1990; Sharman and Holland 1996), thus facilitating the evolution of greater biological complexity (Duboule and Wilkins 1998).

4.2 Loss of the motilin and its specific receptor genes in rodents

The evolution of motilin and its specific receptor in rodents provides an illustration of the consequences of gene loss. Motilin is a 22-amino acid peptide synthesized by endocrine cells of the duodeno-jejunal mucosa and has a profound stimulatory effect on gastrointestinal contractility (Poitras and Peeters 2008), indicating that motilin and its specific receptor serve as potent active prokinetic drug target candidates. However, the clinical development of potential therapies is limited as both the mouse and rat, the most frequently used laboratory animals, are natural knockouts for the motilin and its specific receptor, that is these animals lack these genes and functional targets (He et al. 2010). These observations raise a number of intriguing questions – how can these animals survive without motilin? How were the genes lost? Did any other endocrine system compensate? What does this mean for our understanding of the human hormones? While we can't answer all of these questions, our studies revealed that the motilin receptor was pseudogenized specifically on the rodent lineage, while the motilin gene exhibited diverse evolutionary consequences in different rodent species (He et al. 2010). Once an interacting partner is lost, retained partners may be lost, as demonstrated by the independent loss of MLN in mice, rats and in the guinea pig, or serve as raw material in evolution, as suggested by the retention of MLN in the kangaroo rat. Genomic sequence information suggest, that in the monophyletic Dipodomys subfamily, the MLN gene is intact and is under sustained evolutionary constraint, suggesting it has been recruited into a novel function, a function distinct from traditional motilin signaling (our unpublished observations). Intriguingly, studies have suggested that, after the break down of the MLN signaling pathway, the ghrelin signaling pathway was recruited to compensate for this loss in the rat (Dass et al. 2003; Depoortere et al. 2003). Given the ubiquity and its stochastic nature, the simultaneous loss of a hormone and its specific receptor is unlikely. As a dramatic genetic change, a gene loss leads to an immediate loss of specific interactions. The functional redundancy among gene family members could allow a compromise for the deleterious gene loss. Existing genes can be modified, or recruited, into new interactions that yield new functions through mutation and optimization (Jacob 1977; Khersonsky, Roodveldt and Tawfik 2006; Tokuriki and Tawfik 2009). Motilin is not a unique case. As similar events have occurred to leptin, an important adipose derived hormone (Brennan and Mantzoros 2006; Zhang et al. 1994), which does not exist in the chicken, and likely other birds, while a functional leptin receptor has conserved in these species (Horev et al. 2000; Ohkubo, Tanaka and Nakashima 2000; Pitel et al. 2010). It is possible that the lineage specific losses of motilin and leptin during evolution contributed to the evolution of novel metabolic regulatory mechanisms in these species.

4.3 Evolutionary shifts in existing genes

The proglucagon gene illustrates some of these issues. The vertebrate proglucagon gene encodes three glucagonlike sequences (glucagon, glucagon-like peptide-1 [GLP-1], and glucagon-like peptide-2 [GLP-2]) that play distinct roles in mammalian metabolic regulation

(Drucker 2001; Drucker 2002; Jiang and Zhang 2003; Kieffer and Habener 1999). Glucagon, produced by the A cells of the pancreatic islets, counteracts insulin's effect on blood glucose level depression (Jiang and Zhang 2003). GLP-1 functions as an incretin hormone in mammals, potentiating insulin release, and thus regulating glucose metabolism (Drucker 2001, 2002). In contrast, glucagon and GLP-1 have similar physiological functions in fish, and resemble that of mammalian glucagon (Duguay and Mommsen 1994; Plisetskaya and Mommsen 1996). The receptors for glucagon, GLP-1, and GLP-2 have emerged before the divergence of fish and mammals; however, the GLP-1 class of receptors has specifically been lost in fish, and accordingly the incretin action of GLP-1. A fish specific duplication produced a second glucagon receptor-like gene on the ancestral fish lineage. The new glucagon receptor-like gene shifted its binding specificity from glucagon to GLP-1 ensues, meanwhile maintained the ancestral downstream signaling. Thus through receptor loss and gain, existing hormone was recruited into new roles, and undoubtedly enabled evolutionary divergence (Irwin and Wong 2005).

While ghrelin and its specific receptor (GHSR) genes has been maintained and functionally conserved from fish to mammals, there are some significant differences in the function of the ghrelin/GHSR system in birds compared to other vertebrates (Richards 2010). Some of the actions of ghrelin are conserved in birds (e.g., GH release), while others, such as the effect of ghrelin on food intake, are opposite to those found in mammals and other vertebrate species (Hiroyuki et al. 2007; Kaiya et al. 2009; Kaiya et al. 2008). Besides ghrelin, the ghrelin gene has the potential to encode another peptide hormone – obestatin (Zhang et al. 2005). We observed episodic evolution for both the ghrelin and motilin genes during primitive placental mammal evolution, the period when a functional obestatin hormone might have originated (He, Irwin and Zhang 2010). It is possible that some of the lineage-specific physiological adaptations are due to the episodic evolution of the motilin and ghrelin genes.

Gene duplication, pseudonization, and the gain and loss of interactions through mutations in existing genes are major evolutionary processes shaping the specific interaction among biomolecules (Berg, Lässig and Wagner 2004; Wagner 2001; Wagner 2003; He et al. 2010). Thus, once a mutation arises, a stepwise process of elaboration and optimization ensues, which gradually integrates and orders mutations into a coherent pattern. Given the evolutionary dynamics of the genome and the plasticity of biomolecular networks, an evolutionary perspective is necessary to understand many aspects of the molecular basis of endocrinology.

5. Conclusion

Biological evolution is the process of generating biodiversity. Different phenotype corresponds to a given genomic control. New genes, new interactions, and new biochemical processes are essential for the molecular basis of the evolution of biodiversity and complexity. Genetic networks of endocrine systems are developmentally and evolutionarily fictile, elemental compositions within them are prone to be altered through gene gain and loss, and its physiological properties frequently change with mutations in gene coding sequences and/or regulatory systems, and turnover of interacting biomolecules.

The endocrine system consists of several glands in different parts of the body, which secrete hormones directly into the blood. Hormones usually have many different functions and

modes of action; one hormone may play roles in different target organs, and conversely, target organs are affected by more than one hormone. Although quite irregular, there are still some formulas that can be followed. Structural and evolutionary relatedness generate promiscuous interaction properties, and provide important clues to interaction capacity exploration. Tracing the origin and studying the molecular evolution of endocrine systems should help us comprehend the main events that have prompted the diversification of these systems. In the light of evolution, through a comparative strategy, integrating information from diverse species helps to enhance our understanding of the assemblage of complex endocrine systems, identifying novel components of endocrine systems, and potential cross-talk between them through evolutionarily related interacting proteins. In addition, knowledge of how elements that underlie cellular functions are evolutionarily and developmentally interact, not only helps in choosing appropriate species to examine function, but also provide genetic makers to probe the evolution of specific traits and characteristics, disclosing the genetic basis that underlie the morphological and behavior changes, and thus helping enhance our understanding of how changing environments led to biochemical adjustments.

6. Acknowledgments

This work was supported by grants from the National Basic Research Program of China (973 Program, 2007CB411600), the National Natural Science Foundation of China (30621092, 30623007), and Bureau of Science and Technology of Yunnan Province (O803481101).

7. References

- Avise JC, Ayala FJ. From the Academy: Colloquium Perspective: In the light of evolution I: Adaptation and complex design. *Proc Natl Acad Sci USA*. 2007 104 Suppl 1:8563-8566.
- Berg J, Lässig M, Wagner A. Structure and evolution of protein interaction networks: a statistical model for link dynamics and gene duplications. *BMC Evol Biol*. 2004 4:51.
- Brennan AM, Mantzoros CS. Drug Insight: the role of leptin in human physiology and pathophysiology--emerging clinical applications. *Nat Clin Pract Endocrinol Metab*. 2006 2:318-327.
- Carroll SM, Bridgham JT, Thornton JW Evolution of hormone signaling in elasmobranchs by exploitation of promiscuous receptors. *Mol Biol Evol*. 2008 25:2643-2652.
- Cegla J, Tan TM, Bloom SR. Gut-brain cross-talk in appetite regulation. *Curr Opin Clin Nutr Metab Care*. 2010 13:588-593.
- Chan CB, Cheng CH. Identification and functional characterization of two alternatively spliced growth hormone secretagogue receptor transcripts from the pituitary of black seabream *Acanthopagrus schlegelii*. *Mol Cell Endocrinol*. 2004 214:81-95.
- Conant GC. Rapid reorganization of the transcriptional regulatory network after genome duplication in yeast. *Proc Biol Sci*. 2010 277:869-876.

- Danks JA, D'Souza DG, Gunn HJ, Milley KM, Richardson SJ. Evolution of the parathyroid hormone family and skeletal formation pathways. *Gen Comp Endocrinol*. 2011 170:79-91.
- Dass NB, Munonyara M, Bassil AK, Hervieu GJ, Osbourne S, Corcoran S, Morgan M, Sanger GJ. Growth hormone secretagogue receptors in rat and human gastrointestinal tract and the effects of ghrelin. *Neuroscience* 120:443-453.
- Dehal P, Boore JL. Two rounds of whole genome duplication in the ancestral vertebrate. *PLoS Biol*. 2005 3:e314.
- Depoortere I, De Winter B, Thijs T, De Man J, Pelckman P, Peeters T 2003 Comparison of the prokinetic effects of ghrelin, GHRP-6 and motilin in rats in vivo and in vitro. *Gastroenterology* 124:580.
- Drucker DJ. Biological actions and therapeutic potential of the glucagon-like peptides. *Gastroenterology* 2002 122:531-544.
- Drucker DJ. The glucagon-like peptides. *Endocrinology* 2001 142:521-527
- Duboule D and Wilkins A S. The evolution of 'bricolage'. *Trends Genet*. 1998 14:54-59.
- Duguay SJ, Mommsen TP. Molecular aspects of pancreatic peptides. In: *Fish physiology*, vol. 13 (Hoar WS and Randall DJ, eds). Academic Press, San Diego. 1994 225-271.
- Feighner SD, Tan CP, McKee KK, et al. Receptor for motilin identified in the human gastrointestinal system. *Science* 1999 284:2184-2188.
- Gagneux P, Varki A. Genetic differences between humans and great apes. *Mol Phylogenet Evol*. 2001 18:2-13.
- Garcia-Fernández J, Holland PW. Archetypal organization of the amphioxus Hox gene cluster. *Nature* 1994 370:563-566.
- Gout JF, Duret L, Kahn D. Differential Retention of Metabolic Genes Following Whole-Genome Duplication. *Mol Biol Evol*. 2009 265:1067-1072.
- Gregory TR. Synergy between sequence and size in large-scale genomics. *Nature Rev Genet*. 2005 6:699-708.
- Hahn MW, Demuth JP, Han SG. Accelerated rate of gene gain and loss in primates. *Genetics* 2007 177:1941-1949.
- He J, Irwin DM, Chen R, Zhang YP. Stepwise loss of motilin and its specific receptor genes in rodents. *J Mol Endocrinol*. 2010 44:37-44.
- He J, Irwin DM, Zhang YP. Motilin and ghrelin gene experienced episodic evolution during primitive placental mammal evolution. *Sci China Life Sci*. 2010 53:677-682.
- He J, Irwin DM, Zhang YP. Gene duplication plays a major role in gene co-option: Studies into the evolution of the motilin/ghrelin family and their receptors. *Chinese Science Bulletin*. 2011 56: 2690-2697.
- Hiroyuki Kaiya, Veerle M Darras, Kenji Kangawa. Ghrelin in Birds: Its Structure, Distribution and Function. *Journal of Poultry Science* 2007 44:1-18.
- Hodson DJ, Molino F, Fontanaud P, Bonnefont X, Mollard P. Investigating and modelling pituitary endocrine network function. *J Neuroendocrinol*. 2010 22:1217-1225.
- Hoffmann FG, Opazo JC. Evolution of the relaxin/insulin-like gene family in placental mammals: implications for its early evolution. *J Mol Evol*. 2011 72:72-79.

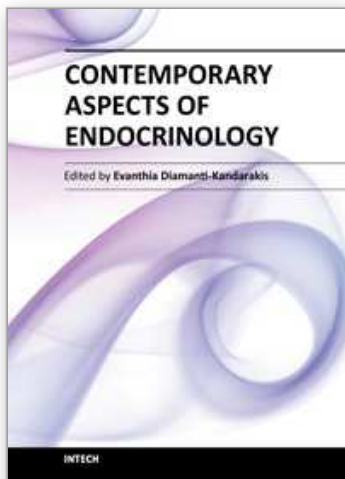
- Holland LZ, Albalat R, Azumi K, et al. The amphioxus genome illuminates vertebrate origins and cephalochordate biology. *Genome Res.* 2008 18:1100-1111.
- Holst B, Holliday ND, Bach A, Elling CE, Cox HM, Schwartz TW. Common structural basis for constitutive activity of the ghrelin receptor family. *J Biol Chem.* 2004 279:53806-53817.
- Horev G, Einat P, Aharoni T, Eshdat Y, Friedman-Einat M. Molecular cloning and properties of the chicken leptin-receptor (CLEPR) gene. *Mol Cell Endocrinol.* 2000 162:95-106.
- Hou Y, Lin S. Distinct gene number-genome size relationships for eukaryotes and non-eukaryotes: gene content estimation for dinoflagellate genomes. *PLoS One.* 2009 4:e6978.
- Howard AD, Feighner SD, Cully DF, et al. A Receptor in Pituitary and Hypothalamus That Functions in Growth Hormone Release. *Science* 1996 273:974-977.
- Huminięcki L, Heldin C-H: 2R and modeling of vertebrate signal transduction engine. *BMC Biol* 2010 8:146.
- Inui A. Ghrelin: an orexigenic and somatotrophic signal from stomach. *Nat. Rev. Neurosci.* 2001 2:551-560.
- Irwin DM, Wong K. Evolution of new hormone function: loss and gain of a receptor. *J Hered.* 2005 96:205-11.
- Irwin DM. Evolution of genes for incretin hormones and their receptors. *Vitam Horm.* 2010 84:1-20.
- Irwin DM. Molecular evolution of proglucagon. *Regul Pept.* 2001 98:1-12
- Jacob F. Evolution and tinkering. *Science* 1977, 196: 1161-1166.
- Jaillon O, Aury J, Brunet F, et al., Genome duplication in the teleost fish *Tetraodon nigroviridis* reveals the early vertebrate proto-karyotype. *Nature* 2004 431:946-957.
- Jiang G, Zhang BB. Glucagon and regulation of glucose metabolism. *Am J Physiol.* 2003 284:E671-E678.
- Kaiya H, Furuse M, Miyazato M, Kangawa K. Current knowledge of the roles of ghrelin in regulating food intake and energy balance in birds. *Gen Comp Endocrinol.* 2009 163:33-38.
- Kaiya H, Kojima M, Hosoda H, et al. Bullfrog ghrelin is modified by n-octanoic acid at its third threonine residue. *J Biol Chem.* 2001 276:40441-40448.
- Kaiya H, Miyazato M, Kangawa K, Peter RE, Unniappan S. Ghrelin: a multifunctional hormone in non-mammalian vertebrates. *Comp Biochem Physiol A Mol Integr Physiol.* 2008 149:109-128.
- Kassahn KS, Dang VT, Wilkins SJ, Perkins AC, Ragan MA. Evolution of gene function and regulatory control after whole-genome duplication: comparative analyses in vertebrates. *Genome Res.* 2009 19:1404-1418.
- Kettler GC, Martiny AC, Huang K, et al. Patterns and implications of gene gain and loss in the evolution of *Prochlorococcus*. *PLoS Genet.* 2007 312:e231.
- Khersonsky O, Roodveldt C, Tawfik DS. Enzyme promiscuity: evolutionary and mechanistic aspects. *Curr Opin Chem Biol.* 2006 10:498-508.
- Kieffer TJ, Habener JF. The glucagon-like peptides. *Endocr Rev.* 1999 20:876-913.

- Kim DK, Cho EB, Moon MJ, Park S, Hwang JI, Kah O, Sower SA, Vaudry H, Seong JY. Revisiting the evolution of gonadotropin-releasing hormones and their receptors in vertebrates: secrets hidden in genomes. *Gen Comp Endocrinol*. 2011 170:68-78.
- Kojima M, Hosoda H, Date Y, Nakazato M, Matsuo H, Kangawa K. Ghrelin is a growth-hormone-releasing acylated peptide from stomach. *Nature* 1999 402:656-60.
- Kojima M, Kangawa K. Ghrelin: structure and function. *Physiol Rev*. 2005 85:495-52.
- Konstantinidis KT, Tiedje JM. Trends between gene content and genome size in prokaryotic species with larger genomes. *Proc Natl Acad Sci USA*. 2004 101:3160-3165.
- Li WH. *Molecular Evolution*. Sinauer Associates, Sunderland, 1997.
- Liu JC, Makova KD, Adkins RM, Gibson S, Li WH. Episodic evolution of growth hormone in primates and emergence of the species specificity of human growth hormone receptor. *Mol Biol Evol*. 2001 18:945-953.
- Lynch M, Conery JS. The origins of genome complexity. *Science* 2003 302:1401-1404.
- Macqueen DJ, Delbridge ML, Manthri S, Johnston IA. A newly classified vertebrate calpain protease, directly ancestral to CAPN1 and 2, episodically evolved a restricted physiological function in placental mammals. *Mol Biol Evol*. 2010 27:1886-1902.
- Manning G, Scheeff E. How the vertebrates were made: selective pruning of a double-duplicated genome. *BMC Biol*. 2010 8:144.
- Markov GV, Paris M, Bertrand S, Laudet V. The evolution of the ligand/receptor couple: a long road from comparative endocrinology to comparative genomics. *Mol Cell Endocrinol*. 2008 293:5-16.
- Matsuura B, Dong M and Miller LJ. Differential determinants for peptide and non-peptidyl ligand binding to the motilin receptor. Critical role of second extracellular loop for peptide binding and action. *J Biol Chem*. 2002 277:9834-9839.
- Moyle WR, Campbell RK, Myers RV, Bernard MP, Han Y, Wang X. Co-evolution of ligand-receptor pairs. *Nature* 1994 368:251-255.
- Murray C D, Kamm M A, Bloom S R, et al. Ghrelin for the gastroenterologist: history and potential. *Gastroenterology* 2003 125:1492-1502.
- Nakazato M, Murakami N, Date Y, Kojima M, Matsuo H, Kangawa K, Matsukura S. A role for ghrelin in the central regulation of feeding. *Nature* 2001 409:194-198.
- Ohkubo T, Tanaka M, Nakashima K. Structure and tissue distribution of chicken leptin receptor (cOb-R) mRNA. *Biochim Biophys Acta*. 2000 1491:303-308.
- Ohno S. *Evolution by gene duplication*. Springer-Verlag, Berlin, 1970.
- Palyha OC, Feighner SD, Tan CP, et al. Ligand activation domain of human orphan growth hormone (GH) secretagogue receptor (GHS-R) conserved from Pufferfish to humans. *Mol Endocrinol*. 2000 14:160-169.
- Peeters T L. Ghrelin: a new player in the control of gastrointestinal functions. *Gut* 2005 54:1638-649.
- Peter EC, Vallo T. Recent advances Advances in endocrinology. *Arch Dis Child*. 1998 78:278-284.

- Pitel F, Faraut T, Bruneau G, Monget P. Is there a leptin gene in the chicken genome? Lessons from phylogenetics, bioinformatics and genomics. *Gen Comp Endocrinol.* 2010 167:1-5.
- Plisetskaya EM, Mommsen TP. Glucagon and glucagon-like peptides in fishes. *Int Rev Cytol*, 1996 168:187-257.
- Poitras P, Peeters TL. Motilin. *Current Opinion in Endocrinology, Diabetes and Obesity* 2008 15:54-57.
- Poitras P: Motilin. In: *Gut Peptides: Biochemistry and Physiology*. Edited by Walsh JH, Dockaray GJ. Raven Press. New York, 1993. 261-303.
- Powell AJ, Conant GC, Brown DE, Carbone I, Dean RA. Altered patterns of gene duplication and differential gene gain and loss in fungal pathogens. *BMC Genomics.* 2008 9:147.
- Richards MP, McMurtry JP. The avian proghrelin system. *Int J Pept.* 2010. pii: 749401.
- Rubenstein PA. The functional importance of multiple actin isoforms. *Bioessays* 1990 12:309-315.
- Samson WK, Lumpkin MD, Nilaver G, McCann SM. Motilin: a novel growth hormone releasing agent. *Brain Res Bull.* 1984 12:57-62.
- Semyonov J, Park JI, Chang CL, Hsu SY. GPCR genes are preferentially retained after whole genome duplication. *PLoS One.* 2008 3:e1903.
- Sharman A C and Holland P W H. Conservation duplication and divergence of developmental genes during chordate evolution. *Netherlands Journal of Zoology* 1996 46:47-67.
- Sun Y, Ahmed S, Smith RG. Deletion of ghrelin impairs neither growth nor appetite. *Mol Cell Biol.* 2003 23:7973-7981.
- Sundström G, Dreborg S, Larhammar D. Concomitant duplications of opioid peptide and receptor genes before the origin of jawed vertebrates. *PLoS One.* 2010 5:e10512.
- Ting CT, Tsaur SC, Sun S, Browne WE, Chen YC, Patel NH, Wu CI. Gene duplication and speciation in *Drosophila*: evidence from the *Odysseus* locus. *Proc Natl Acad Sci USA.* 2004 101:12232-12235.
- Tokuriki N, Tawfik DS. Protein dynamism and evolvability. *Science* 2009 324:203-207.
- True JR, Carroll SB. Gene co-option in physiological and morphological evolution. *Annu Rev Cell Dev Biol.* 2002 18:53-80.
- van Hoek MJ, Hogeweg P. Metabolic adaptation after whole genome duplication. *Mol Biol Evol.* 2009 26:2441-2453.
- Wagner A. How the global structure of protein interaction networks evolves. *Proc Biol Sci.* 2003 270:457-466.
- Wagner A. The yeast protein interaction network evolves rapidly and contains few redundant duplicate genes. *Mol Biol Evol.* 2001 18:1283-1292.
- Wallis M. Episodic evolution of protein hormones in mammals. *J Mol Evol.* 2001 53:10-18.
- Wang X, Grus WE, Zhang J. Gene losses during human origins. *PLoS Biology.* 2006 4:e52.
- Wilkins AS. Between "design" and "bricolage": genetic networks, levels of selection, and adaptive evolution. *Proc Natl Acad Sci USA.* 2007 104 Suppl 1:8590-8596.

- Zhang J, Zhang YP, Rosenberg HF. Adaptive evolution of a duplicated pancreatic ribonuclease gene in a leaf-eating monkey. *Nat Genet.* 2002 30:411–415.
- Zhang J. Evolution by gene duplication: an update. *Trends in Ecology & Evolution* 2003 18:292-298.
- Zhang JV, Ren PG, Avsian-Kretchmer O, et al. Obestatin, a peptide encoded by the ghrelin gene, opposes ghrelin's effects on food intake. *Science* 2005 310:996-999.
- Zhang Y, Proenca R, Maffei M, Barone M, Leopold L, Friedman JM. Positional cloning of the mouse obese gene and its human homologue. *Nature* 1994 372:425-32.
- Zhou XR, Wang YZ, Smith JF, Chen R. Altered expression patterns of TCP and MYB genes relating to the floral developmental transition from initial zygomorphy to actinomorphy in *Bournea* (Gesneriaceae). *New Phytol.* 2008 178:532-543.

IntechOpen



Contemporary Aspects of Endocrinology

Edited by Dr. Evanthia Diamanti-Kandarakis

ISBN 978-953-307-357-6

Hard cover, 454 pages

Publisher InTech

Published online 30, November, 2011

Published in print edition November, 2011

This book aims to provide readers with a general as well as an advanced overview of the key trends in endocrine disorders. While covering a variety of topics ranging from thyroid carcinogenesis and pituitary adenomas to adrenal tumors and metabolic bone disease, this book also focuses on more specific issues not yet fully elucidated (e.g. the molecular pathways involved in thyrotropin beta gene regulation or monogenic phosphate balance disorders). Readers of different fields and background will have the opportunity to update their knowledge and more importantly to clarify areas of uncertainty and controversies in several topics of endocrine disorders.

How to reference

In order to correctly reference this scholarly work, feel free to copy and paste the following:

Jing He, David M. Irwin and Ya-Ping Zhang (2011). The Power of an Evolutionary Perspective in Studies of Endocrinology, Contemporary Aspects of Endocrinology, Dr. Evanthia Diamanti-Kandarakis (Ed.), ISBN: 978-953-307-357-6, InTech, Available from: <http://www.intechopen.com/books/contemporary-aspects-of-endocrinology/the-power-of-an-evolutionary-perspective-in-studies-of-endocrinology>

INTECH
open science | open minds

InTech Europe

University Campus STeP Ri
Slavka Krautzeka 83/A
51000 Rijeka, Croatia
Phone: +385 (51) 770 447
Fax: +385 (51) 686 166
www.intechopen.com

InTech China

Unit 405, Office Block, Hotel Equatorial Shanghai
No.65, Yan An Road (West), Shanghai, 200040, China
中国上海市延安西路65号上海国际贵都大饭店办公楼405单元
Phone: +86-21-62489820
Fax: +86-21-62489821

© 2011 The Author(s). Licensee IntechOpen. This is an open access article distributed under the terms of the [Creative Commons Attribution 3.0 License](https://creativecommons.org/licenses/by/3.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

IntechOpen

IntechOpen