We are IntechOpen, the world's leading publisher of Open Access books Built by scientists, for scientists



186,000

200M



Our authors are among the

TOP 1% most cited scientists





WEB OF SCIENCE

Selection of our books indexed in the Book Citation Index in Web of Science™ Core Collection (BKCI)

Interested in publishing with us? Contact book.department@intechopen.com

Numbers displayed above are based on latest data collected. For more information visit www.intechopen.com



Anticancer Properties of Curcumin

Varisa Pongrakhananon^{1,2} and Yon Rojanasakul² ¹Chulalongkorn University, Department of Pharmacology and Physiology Faculty of Pharmaceutical Sciences, Bangkok, ²West Virginia University Department of Basic Pharmaceutical Sciences, Morgantown, West Virginia ¹Thailand ²USA

1. Introduction

Curcumin is a major biological active compound from turmeric or Curcuma longa. This nontoxic natural compound has been reported to possess several biological activities that are therapeutically beneficial to cancer treatment. It has been reported to increase the efficacy of other chemotherapeutic agents and to reduce their toxic side effects which are the major drawback of most chemotherapeutic agents. Curcumin is also well known for its antiinflammatory activity (Amanda and Robert, 2008). Since cancer often develops under chronic inflammatory conditions, curcumin has the potential to be a preventive treatment agent against cancer. Furthermore, unlike most chemotherapeutic agents which act on a specific process of cancer development, i.e., cell growth or apoptosis, curcumin exerts its effect on various stages of cancer development, i.e., oncogene activation (Singh and Singh, 2009), cancer cell proliferation (Simon et al., 1998), apoptosis evasion (Han et al., 1999), anoikis resistance (Pongrakhananon et al., 2010), and metastasis (Chen et al., 2008) (Figure 1). Therefore, curcumin has the potential to overcome chemoresistance which is a major problem in cancer chemotherapy. This chapter will provide an overview of the anticancer activities of curcumin and present pre-clinical and clinical evidence supporting the use of curcumin as an anticancer agent.

Cancer is known to be associated with genetic instability in which *c-myc* serves as a major modifier of many targeted genes (Mai and Mushinski, 2003). Likewise, the mutation of proto-oncogene *ras* has been identified in many types of tumor (Rajalingam et al., 2007). The dysregulation of these oncogenes is well recognized as an initial step in the development of tumorigenesis. Interestingly, curcumin has been reported to have a suppressive effect on the oncogenes and inhibit their downstream effectors such as cell cycle promoting and proapoptotic proteins (Singh and Singh, 2009). Curcumin also exhibits anticancer properties through its ability to inhibit cell proliferation and induce apoptosis. The anti-proliferative effect of curcumin is dependent on its concentration, duration of treatment, and specific cell type. At low doses, curcumin causes cell cycle arrest, while at higher doses it induces apoptosis. Cell proliferation is controlled by several cell cycle regulating proteins, notably the family of cyclin and cyclin-dependent kinases (Kastan and Bartek, 2004) whose expression is tightly associated with tumorigenesis (Diehl, 2002). Curcumin inhibits cell cycle progression by downregulating cyclin D1 and the transition from G1 to S phase in

human head and neck squamous carcinoma cells (Aggarwal et al., 2004). It also inhibits bladder cancer cell proliferation through the downregulation of cyclin A and upregulation of p21 (Park et al., 2006).



Fig. 1. Anticancer properties of curcumin

Curcumin possesses apoptosis-inducing activity causing cancer cell death primarily through the mitochondrial death pathway. It induces an upregulation of the proapoptotic protein Bax and downregulation of the antiapoptotic protein Bcl-2 in breast cancer cells (Chiu and Su, 2009), resulting in the loss of mitochondrial function, release of cytochrome c, and activation of caspase-9 and -3 (Chen et al., 2010). Curcumin also potentiates the cytotoxic effect of chemotherapeutic agents such as cisplatin (Chanvorachote et al., 2009), doxorubicin (Notarbartolo et al., 2005), tamoxifen (Chuang et al., 2002), and placitaxel (Genta and Amiji, 2009). The use of curcumin as a chemo-sensitizing agent in combination therapy has the potential to overcome chemoresistance which is common in advance staged cancers and is a major cause of cancer-related death.

An increasing number of reports have described the inhibitory effect of curcumin on cancer metastasis. Metastasis is a multi-step process involving tumor vascularization, cancer cell detachment, avoidance of anoikis, and increased cell invasion. The vascularization induced by tumor is an essential step providing nutrients, oxygen, and removing waste products for tumor growth and metastasis. A key mechanism that cancer cells utilize during metastasis is the acquisition of anoikis resistance. Anoikis or detachment-induced apoptosis is recognized as an important mechanism preventing cancer cell dissemination and invasion to form

secondary tumors. A recent study by our group has shown that curcumin is able to sensitize lung cancer cells to undergo anoikis through a mechanism that involves post-translational modification of Bcl-2 via the ubiquitin-proteasome pathway (Pongrakhananon et al., 2010). Curcumin also acts as a negative regulator of cancer cell migration and invasion through diverse signaling pathways including MMP-9, MMP-2 and COX-2 (Philip et al., 2004, Lee et al., 2005; Hong et al., 2006; Lin et al., 2009).

Animal and clinical studies of curcumin have been well investigated. In mice, curcumin markedly inhibits DMBA and TPA-induced skin tumor formation (Azuine et al., 1992). In a xenograft model, curcumin administration significantly decreases the incidence of breast cancer metastasis to the lung (Aggarwal et al., 2005). In phase 1 clinical studies, oral administration of curcumin was shown to be well tolerated with no dose-limiting toxicity (Sharma et al., 2004; Lao et al., 2006). In patients with intestinal metaplasia, curcumin treatment showed a significant improvement in the precancerous lesion (Cheng et al., 2001), supporting the clinical use of curcumin as a preventive treatment agent against cancer. Although curcumin has demonstrated promising pharmacological effects and safety both *in vitro* and *in vivo*, poor bioavailability and tissue accumulation have been observed with the compound. Further studies on proper drug delivery, dose optimization, and biodistribution are needed.

2. Chemistry of curcumin

For thousands of years, plants and some parts of animal have been used as dietary agents which have been identified to be biologically active. These natural compounds have gained considerable interest for their potential as treatment and preventive agents for human diseases. Curcumin (diferuloylmethane) is a major biologically active compound extracted from the dried rhizome of turmeric or *Curcuma longa*. It has been widely used for centuries as medicinal plant and food additive due to its yellow color. Its medicinal properties are attributed to curcuminoids, which include curcumin (curcumin I), demethoxycurcumin (curcumin II), and bisdemethoxycurcumin (curcumin III) (Figure 2). Curcumin I (77%) is a

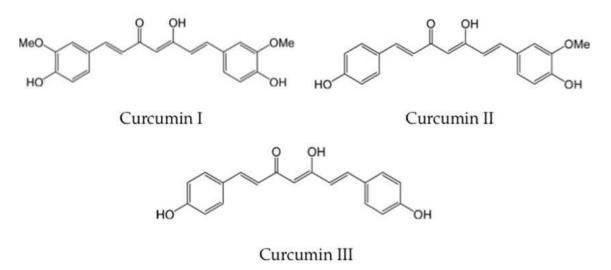


Fig. 2. Chemical structure of curcuminoids

major component found in commercial curcumin, while curcumin II and III constitute approximately 17% and 3% respectively. Cucumin is a water-insoluble compound, but

dissolves well in ethanol, dimethylsulfoxide, and other organic solvents. The molecular weight of curcumin is 368.37 and melting point is 183 °C. It shows a spectrophotometric maximum absorption (λ max) at 450 nm in methanol (Prasad and Sarasija, 1997). Fluorescence of curcumin occurs at 524 nm in acetonitrile and 549 nm in ethanol (Chignell et al., 1994). Curcumin undergoes rapid degradation in phosphate buffer and serum-free media, i.e., 90% within 30 minutes (Wang et al., 1997). In serum-containing (10%) media and human blood, curcumin is more stable with less than 20% degradation in 1 hour, and about 50% after 8 hours. Its degradation products are *trans*-6-(4'-hydroxy-3'-methoxyphenyl)-2,4-dioxo-5hexenal (major) and vanillin, ferulic acid, and feruloyl methane (minor) (Wang et al., 1997).

3. Anticancer properties of curcumin

Curcumin has long been used as a dietary ingredient with known health benefits. Extensive research over the last decade has shown that curcumin possesses anticancer activities and could be used as a preventive or treatment agent against cancers, either as a single or combination therapy with chemotherapeutic agents. Curcumin exhibits biological activities in various stages of carcinogenesis including inhibition of oncogene activation, prevention of cancer-related inflammation, inhibition of cancer cell proliferation, induction of apoptosis and anoikis, prevention of metastasis, and sensitization of cancer cells to chemotherapy.

3.1 Cancer prevention

3.1.1 Chemopreventive properties

Carcinogenesis is a multistep process driven by genetic instability. Continuous exposure to environmental and endogenous genotoxic agents can cause substantial DNA damage. The damaged molecules can be transmitted during cell division producing mutated clones that can give rise to the expansion of premalignant cell population possessing uncontrolled proliferative and invasive properties. Curcumin has been established as a chemopreventive agent that has the ability to suppress or retard the carcinogenic process induced by various chemical carcinogens (Table 1). In animal models of gastric and colon cancer, curcumin inhibits the development of cancerous and precancerous lesions induced by N-methyl-N'nitro-N-nitrosoguanosine (MNNG), a known mutagenic agent causing DNA methylation (Ikesaka et al., 2001). In the study, MNNG was given in drinking water at the concentration of 100 ppm for 8 weeks, and then 0.2% or 0.5% of curcumin was fed to the rats for 55 weeks. The results showed that the number of atypical hyperplasia in curcumin-treated rats was significantly less than that in the control group. Similarly, the curcumin analog bis-1,7-(2hydroxyphenyl)-hepta-1,6-diene-3,5-dione was shown to inhibit the tumorigenic effect of 1,2-dimethylhydrazine in rats (Devasena et al., 2003). Furthermore, natural and synthetic curcuminoids exhibit an inhibitory effect on mutagenesis induced by 2-acetamidofluorene (2-AAF) (Anto et al., 1996). In the study, up to 87% of 2-AAF-induced papilloma was inhibited by bis-(p-hydroxycinnamoyl)methane (curcuminoid III), while 70% and 68% of the papilloma were inhibited by feruloyl-p-hydroxycinnamoylmethane (curcuminiod II) and diferuloylmethane (curcuminoid I), respectively. The most potent curcuminoid was salicylcurcuminoid which completely inhibited the papilloma formation.

3.1.2 Suppression of oncogenes and upregulation of tumor suppressor genes

As mentioned earlier, genetic instability is linked to the initiation of carcinogenesis. This irreversible process involves several molecular events that either promote the activity of

348

oncogenes such as myc and ras or impede the function of tumor suppressor genes such as p53 (Vogelstein and Kinzler, 2004). Curcumin has been shown to suppress oncogenes and activate tumor suppressor genes in various cancer cell types (Table 1). In an *in vivo* study, curcumin suppresses c-fos and c-Ha-ras activation induced by environmental mutagenic agents (Limtrakul et al., 2001). Dietary administration of curcumin (0.1-0.2%) prevents 2-dimethylbenz(α)anthracene (DMBA) and 12-O-tetradecanoylphorbol-13-acetate (TPA)-induced skin tumor in mice. There is an increase in the oncogene (c-fos and ras) expression in this tumor model, which is suppressed by the dietary curcumin. Similarly, c-myc is also a target oncogene attenuated by curcumin in the TPA-induced tumor model (Kakar and Roy, 1994).

Chemopreventive properties					
Cancer	Carcinogen Animal		References		
Stomach cancer	MNNG	MNNG Rat			
Colon cancer	DMH	Rat	Devasena et al., 2003		
Papilloma	2-AAF	-	Anto et al., 1996		
Mammary tumor	DMBA	Rat	Pereira et al., 1996		
Skin tumor	TPA	Mouse	Lu et al., 1993		
Liver cancer	Diethylnitrosamine	Mouse	Chuang et al., 2000		
Oncogene	Oncogene suppression and tumor suppressor gene activation				
Cancer	Mechanism	Mechanism			
Skin tumor	Suppress c-fos and c-Ha-ras- activation by DMBA		Limtrakul et al., 2001		
B cell lymphoma	Suppress c-myc	•			
Mouse skin cancer	Suppress c-myc activat	Suppress c-myc activated by TPA			
Human colon adenocarcinoma	Enhance p53 activity		Song et al., 2005		
Human breast cancer	Enhance p53 activity		Choudhuri et al., 2002		
Human glioma	Enhance p53 activity		Liu et al., 2007		
	Anti-inflammation -related cancer				
Cancer	Mechanism		References		
-	Inhibit NO synthase in macrophages		Brouet & Ohshima, 1995		
Human mantle cell lymphoma	Inhibit constitutive NF-κB activation		Shishodia et al., 2005		
Mouse melanoma	Inhibit constitutive NF-κB activation		Marin et al., 2007		
Human oral cancer	Inhibit constitutive NF-κB activation		Sharma et al., 2006		
Non-small cell lung carcinoma	Inhibit constitutive NF-кВ activation and COX expression		Shishodia et al., 2003		

Table 1. Cancer preventive properties of curcumin

3.1.3 Anti-inflammatory activity

Epidemiological studies have supported the concept that cancers frequently originate at the site of chronic inflammation. This event has increasingly been accepted as the seventh hallmark of cancer (Colotta et al., 2009). Inflammation is a vital physiological process in response to injury, which leads to the mediation of inflammatory cells in the presence of enzymes and cytokines for repairing tissue damage. The linkage between cancer and inflammation is generally characterized as being intrinsic or extrinsic (Mantovani et al., 2008). The extrinsic pathway is induced by inflammation that facilitates cancer development, while the intrinsic pathway is driven by genetic instability causing inflammatory environment-related cancer. Both processes mediate the transcription factors involved in cell proliferative function and persistent activation of this event can lead to cancer. A considerable number of reports have described the linkage between cancer preventive properties and anti-inflammatory action of curcumin (Table 1). Mechanistically, curcumin inhibits the induction of nitric oxide synthase (NOS) by activated macrophages (Brouet and Ohshima, 1995). Nitric oxide (NO) and its derivatives such as peroxynitrite play a critical role in the inflammation process causing fibroblast proliferation and fibrosis (Romanska et al., 2002). Treatment of macrophages with lipopolysaccharide (LPS) and IFN-y induces an inflammatory response that leads to the release of NO, which is inhibited by curcumin. Curcumin also suppresses the transcription of inducible NOS which is activated by LPS and IFN-γ. Since NO is implicated in tumor promotion, the attenuation of NO by curcumin hence suppresses tumor development.

Numerous studies have suggested that NF-KB plays a key role in promoting cancer development during chronic inflammation. NF-kB regulates the expression of several genes involved in the immune and inflammatory responses as well as in cell proliferation and apoptosis (Karin et al., 2002). It is required for the expression of a wide range of inflammatory cytokines and adhesion molecules, and is important in the cellular proliferative function by activating growth factor genes, proto-oncogenes and cell cycle regulators that contribute to carcinogenesis. In a study using human mantle cell lymphoma (MCL), curcumin inhibits constitutive NF-κB activation leading to the suppression of NF-κB regulated genes (Shishodia et al., 2005). MCL expresses a high level of cyclin D1, a cell cycle promoting factor and target transcription of NF-kB, which is a key survival factor of this cancer type. Curcumin treatment causes downregulation of constitutive NF-KB activation, inhibits IκBα kinase (IKK) and phosphorylated IκBα and p65, leading to cell cycle arrest and apoptosis. NF-kB is overexpressed in various tumors and cancer cell lines including melanoma cells. Electrophoretic mobility shift and gene reporter assays have demonstrated that curcumin suppresses constitutive NF-kB activation in melanoma cells and increases the number of cells in the sub G1 phase of cell cycle (Marin et al., 2007). Interestingly, curcumin shows selectivity in inducing apoptosis of melanoma cells but not melanocytes.

Other mechanisms of the anti-inflammatory-related cancer effect of curcumin have been proposed including suppression of cyclooxygenase 2 (COX-2). A wide range of stimuli mediates COX-2 expression and overexpression of this molecule is found in various cancer types in association with accelerated cell growth, antiapoptotic activity, angiogenesis, and metastasis (Prescott and Fitzpatrick, 2000). In a cigarette-smoking (CS) study using non-small cell lung cancer model, CS exposure activates NF-κB and subsequently induces COX-2 expression, which is inhibited by pre-treatment with curcumin (Shishodia et al., 2003). This chemopreventive property of curcumin is also observed in smokeless-tobacco mediated

activation of NF-κB and its downstream target COX-2 (Sharma et al., 2006). Since inflammation contributes to tumor initiation, and curcumin possesses anti-inflammatory activity, curcumin could be beneficial in cancer prevention.

3.2 Inhibition of tumor growth and cell proliferation

Excessive proliferation is a hallmark of cancer (Hanahan and Weinberg, 2000). Aberrational cell division is an important basis of cancer development allowing formation and expansion of tumor growth. In normal cells, cell proliferation requires the growth signal that is generated by cell-cell interaction, neighbour cells and extracellular matrix. Cancer cells can further generate their own growth signals, upregulate growth receptors, and be desensitized to antigrowth factors. Upon transmitting the growth signals to their receptors, downstream mediators are activated which drive the quiescent cells to proliferative cycle (Evan and Vousden, 2001). Curcumin has been shown to modulate the expression and activity of growth factors, including epidermal growth factor (EGF) and insulin-like growth factor (IGF) (Table 2). Overexpression of EGF and its receptor, EGFR, was found in human prostate cancer cells undergoing rapid growth expansion (Cai et al., 2008). It has been demonstrated that curcumin blocks the EGF pathway through downregulation of EGFR, suppression of intrinsic EGFR tyrosine kinase activity, and inhibition of ligand-induced EGFR activity in both androgen-dependent and androgen-independent prostate cancer cells (Dorai et al., 2000).

Inhibition of cancer proliferation and tumor growth			
Cancer	Mechanism	References	
Prostate cancer cells	Downregulation of EGFR, suppression of intrinsic EGFR tyrosine kinase activity, and inhibition ligand-induced EGFR activity	Dorai et al., 2000	
Prostate and breast cancer cells	Downregulation of cyclin D1	Mukhopadhyay et al., 2002	
Breast cancer cells	Suppression IGF system	Xia et al., 2007	
Breast cancer cells	Induction of cell cycle arrest at S, G2/M phase by upregulation of p21	Chiu & Su, 2009	
Human epidermoid carcinoma cells	Inhibit EGFR activity	Korutla & Kumar, 1994	
Colon carcinoma cells	Induction of cell cycle arrest at S, G2/M phase	Chen et al., 1999	
Human head and neck squamous carcinoma cells	Downregulation of cyclin D1	Aggarwal et al., 2004	
Human biliary cancer cells	Downregulation of cyclin D1	Prakobwong et al., 2011	
Human hepatocarcinoma cells	Induction of cell cycle arrest at S, G2/M phase	Cheng et al., 2010	

Table 2. Inhibition of cancer proliferation and tumor growth

Likewise, insulin-like growth factor has been implicated in the regulation of normal cell growth, in which its atypical expression is found in human cancer cells (Samani et al., 2007). Curcumin abrogates IGF-1 system in MCF-7 human breast carcinoma cells (Xia et al., 2007). It decreases IGF-1 secretion in concomitant with the increasing IGF binding protein IGFBP-3 in a dose-dependent fashion. Furthermore, it abolishes IGF-1-stimulated MCF-7 growth through the suppression of IGF-1 mediated receptor activity and downregulation of IGF receptor mRNA expression.

Deregulation of cell cycle causes limitless replicative potential of cancer. Curcumin exhibits antiproliferative activity via the regulation of cell cycle (Table 2). It disrupts the progression of cell cycle by increasing the number of cancer cells at S, G2/M phase, hence preventing cell entering next cycle (Chen et al., 1999). Additional mechanistic studies indicate that curcumin downregulates cyclin D1 expression through inhibition of its promoter activity and enhanced degradation (Mukhopadhyay et al., 2002). Cyclin D1 is a subunit of cyclin-dependent kinase (Cdk)-4 and Cdk-6 which plays a key role in determining the cell cycle progression from G1 to S phase (Baldin et al., 1993) and overexpression of this gene is a common event in several forms of cancer (Knudsen et al., 2006). Similarly, in the presence of curcumin, cyclin D1 is suppressed as a result of NF- κ B inhibition as observed in human head carcinoma, neck squamous carcinoma, and biliary cancer cells (Aggarwal et al., 2004; Prakobwong et al., 2011).

3.3 Induction of cancer cell apoptosis

Apoptosis plays an essential role in various physiological and pathological processes (Hengartner, 2000). Tissue homeostasis maintains the balance of cell proliferation and cell death as part of normal tissue development. Dysregulation of this process can lead to several diseases including cancer. Avoidance of apoptotic cell death is a major characteristic of malignant cells in response to stress conditions, which is achieved by activating the antiapoptotic signals or inhibiting proapoptotic signals (Hanahan and Weinberg, 2000). Several strategies have been developed to overcome this defective mechanism in cancers and curcumin has shown promising activities that modulate this mechanism in favor of cancer cell apoptosis.

Apoptosis generally occurs through two main pathways, intrinsic and extrinsic (Lavrik et al., 2005). The intrinsic pathway is initiated by several cellular stresses such as DNA damage which activates proapoptotic proteins, notably the Bcl-2 family proteins, causing mitochondrial membrane permeabilization and subsequent activation of the caspase cascade. In the extrinsic pathway, the interaction between death ligands and death receptors results in the assembly of death-inducing signaling complex (DISC) and the activation of initiator caspases. Curcumin is known to overcome the apoptosis resistance of cancer cells through both pathways (Table 3). In human acute myelogenous leukemia HL-60 cells, curcumin induces apoptosis by suppressing the expression of antiapoptotic Bcl-2 and Bcl-xL, causing cytochrome c release, caspase-3 activation, and PARP cleavage (Anto et al., 2002). Curcumin also stimulates the death receptor pathway through caspase-8 activation but overexpression of the DISC protein FADD cannot protect the cells from apoptosis in response to curcumin. In A549 lung adenocarcinoma cells, curcumin treatment up-regulates the mitochondrial Bax protein expression, suggesting the intrinsic pathway as a major pathway of curcumin-induced apoptosis in this cell type (Chen et al., 2010).

352

Cancer	Mechanism	References Anto et al., 2002	
Human acute myelogenous leukemia cells	Downregulation of Bcl-2 and Bcl- xL		
Lung adenocarcinoma cells	Upregulation of Bax and Downregulation of Bcl-2	Chen et al., 2010	
Rat histocytoma cells	Induction of ROS production	Bhaumik et al., 1999	
Human gingival fibroblasts and human submandibular gland carcinoma cells	Induction of ROS production	Atsumi et al., 2006	
Human breast cancer cells	Downregulation of anti-apoptotic and upregulation of proapoptotic proteins in a p53-dependent manner	Choudhuri et al., 2002	
Human breast cancer cells	Inhibition of PI3K/Akt pathway	Squires et al., 2003	
Human breast and hepatic cancers cells	Glutathione depletion and ROS production	Syng-Ai et al., 2004	
Human ovarian cancer cells	Upregulation of caspase-3 and downregulation of NF-кВ expression	Zheng et al., 2002	
Human colon cancer cells	Induction of ROS production and deactivation of JNK pathway	Moussavi et al., 2006	
Human colon adenocarcinoma cells	Downregulation of anti-apoptotic and upregulation of proapoptotic proteins in a p53-dependent manner	Song et al., 2005	
Human melanoma cells	Activation of caspase-3, and -8, Fas receptor aggregation, suppression of NF-κB activation, and downregulation of XIAP expression	Bush et al., 2001	
T cell leukemia cells	Inhibition of PI3K/Akt pathway	Hussain et al., 2006	
Prostate cancer cells	Inhibition of PI3K/Akt pathway	Shankar & Srivastava, 2007	
Human glioblastoma cells	Increasing Bax:Bcl-2 ratio, activation of caspase-8, -9, and -3, downregulation of NF-кВ	Karmakar et al., 2006	
Breast cancer cells	Inhibition of MAPK and PI3K/ Akt pathway	Squires et al., 2003	

Table 3. Molecular mechanisms of curcumin-induced apoptosis

In human melanoma cells, curcumin induces apoptosis through the extrinsic pathway by activating caspase-8 (Bush et al, 2001). Inhibition of caspase-8 by specific caspase-8 inhibitor or pan-caspase inhibitor prevents cancer cell apoptosis, whereas specific caspase-9 inhibitor lacks this ability. The underlying apoptosis mechanism involves the induction of Fas receptor oligomerization independent of Fas ligand, supporting the role of death receptor modification in curcumin-induced apoptosis. In some cancers such as human glioblastoma T98G cells, curcumin induces apoptosis through both pathways (Karmakar et al., 2006).

The activation of apoptotic pathwas by curcumin is regulated by reactive oxygen species (ROS). In various cancer cell types including H460 non-small cell lung cancer cells (Chanvorachote et al., 2009), AK5 rat histocytoma cells (Bhaumik et al., 1999), human gingival fibroblasts (HGF), and human submandibular gland carcinoma (HSG) cells (Atsumi et al., 2006), the induction of apoptosis by curcumin requires ROS generation. Likewise, in MCF-7, MDAMB, and HepG2 cells, apoptosis is mediated through oxidative stress induced by curcumin as a result of glutathione depletion (Syng-Ai et al., 2004). In colon cancer cells, curcumin-induced cell death is associated with ROS generation that converges on JNK activation (Moussavi et al., 2006).

Other mechanisms of curcumin-induced apoptosis have been reported. In human HT-29 colon adenocarcinoma (Song et al., 2005) and breast cancer cells (Choudhuri et al., 2002), curcumin upregulates proapoptotic proteins and downregulates antiapoptotic proteins in the Bcl-2 family through p53-dependent mechanism. In T cell leukemia (Hussain et al., 2006) and breast cancer cells (Squires et al., 2003), apoptosis induced by curcumin involves PI3K/Akt signaling pathway. Curcumin inhibits Akt phosphorylation and upregulates p53 expression which induces the proapoptotic Bcl-2 family proteins facilitating cell apoptosis (Shankar and Srivastava, 2007).

3.4 Sensitization of cancer cells to chemotherapy

In addition to its direct apoptosis-inducing effect, curcumin has been reported to sensitize cancer cells to chemotherapy-induced cell death. The main problem of cancer chemotherapy is the acquisition of apoptosis resistance of cancer cells and the cytotoxicity to normal cells. Combination therapy is an alternative approach that can overcome this problem by potentiating the effects of combination drugs and reducing their cytotoxicity by using optimal dosage regimens. Curcumin is one such agent that has been investigated for its effect in combination therapy (Table 4). In non-small cell lung cancer cells, we have recently shown that cotreatment of the cells with cisplatin and curcumin results in a substantial increase in cancer cell death as compared to cisplatin treatment alone (Chanvorachote et al., 2009). Since cisplatin-based therapy is the first line drug for treatment of lung cancer and the efficacy of this drug is frequently attenuated by the development of drug resistance especially in advanced stage cancer (Chang, 2011), curcumin has the potential to overcome this resistance problem. Curcumin promotes the apoptotic effect of cisplatin by inducing superoxide anion and downregulating the anti-apoptotic Bcl-2 protein which facilitates the cancer cell killing by cisplatin.

In pancreatic cancer cells, the acquisition of apoptosis resistance to the combination therapy of gemcitabine and capecitabine (a prodrug of 5-fluorouracil, 5-FU) is frequently observed. It is thought that overexpression of the multidrug resistance-associated protein 5 (MRP5), which promotes cellular efflux of the drugs (Szakacs et al., 2006), contributes to the

354

resistance. Curcumin was shown in a recent study to inhibit MRP5 activity and increase the sensitivity of pancreatic cancer cells to 5-FU-induced toxicity in a dose-dependent manner (Li et al., 2010).

Cancer cells	Chemotherapeutic drugs	References
Non-small cell lung cancer cells	Cisplatin	Chanvorachote et al., 2009
Pancreatic cancer cells	5-Fluorouracil	Li et al., 2010
Hela cells	Taxol	Bava et al., 2005
Human hepatic cancer cells	Doxorubicin	Notarbartolo et al., 2005
Hepatocellular carcinoima cells	Doxorubicin	Chuang et al., 2002
Human ovarian adenocarcinoma cells	Paclitaxel	Ganta & Amiji, 2009
Human melanoma cells	Tamoxifen	Chatterjee & Pandey, 2011
Human colorectal cancer cells	Oxaliplatin	Howells et al., 2010
Bladder cancer cells	Gemcitabine	Tharakan et al., 2010

Table 4. Curcumin potentiates chemotherapeutic agent-induced cancer cell death

Curcumin also augments the therapeutic effect of taxol in Hela cells (Bava et al., 2005). The synergistic mechanism involves the inhibition of NF- κ B activation and Akt phosphorylation which results in increased apoptosis and decreased DNA synthesis of cancer cells independent of tubulin polymerization. The increased susceptibility of cancer cells to chemotherapeutic agents by curcumin might overcome the drug resistance problem, thus improving the clinical outcomes.

3.5 Inhibition of cancer metastasis

Cancer metastasis is the spread of cancer cells from the initiation site to other parts of the body, and particularly presented in several advanced stage cancers which are difficult to treat (Gubta and Massagué, 2006). Cancer metastasis is a multistep process involving complex interactions between the disseminating cancer cells and their microenvironment. When transformed cells are initiated and continue to grow at the primary site, angiogenic factors are synthesized for vascularization which increases the likelihood of tumor cells to enter in the blood stream or lymphatic system and colonize at distant sites. Once at the new sites, the extravasation of cancer cells allows the formation and growth of secondary tumors which complete the metastatic process (Fidler, 2003). Agents that inhibit metastasis provide a major advantage in treating cancers. Several studies have shown that curcumin inhibits cancer angiogenesis, migration and invasion by interacting with key regulatory molecules as summarized in Table 5.

It is well established that vascular endothelial growth factor (VEGF) and matrix metalloproteinase family proteins (MMP) are essential factors in the angiogenesis and invasion of cancer cells (Carmeliet, 2005; Helmestin et al., 1994). In non-small cell lung cancer cells, VEGF, MMP-9 and MMP-2 are inhibited by curcumin through MEKK and ERK-dependent pathways, resulting in inhibition of cell migration and invasion (Lin et al., 2009). Similarly, in a human glioblastoma xenograft mouse model, curcumin inhibits tumor growth,

suppresses angiogenesis, and increases animal survival through the inhibition of MMP-9 and neovascularization (Perry et al., 2010). Curcumin also acts as a potent inhibitor of breast cancer cell motility and invasion through the attenuation of MMP-3 which acts as an invasive factor in this cancer cell type (Boonrao et al., 2010).

Cancer	Effects	Mechanism	References
Human non-small cell lung cancer	Inhibition of cell invasion and migration	Inhibition of VEGF, MMP-9, and MMP-2 through MEKK and ERK pathway	Lin et al., 2009
Human glioblastoma	Suppression of angiogenesis	Inhibition of MMP-9	Perry et al., 2010
Human breast cancer	Inhibition of cancer motlity and invasion	Attenuation of MMP-3 activity	Boonrao et al., 2010
Human non-small cell lung cancer	Sensitization of cancer cell anoikis	Downregulation of Bcl-2 through proteasomal degradation	Pongrakhananon et al., 2010
Human lung cancer	Inhibition invasion and metastasis	Activation of tumor suppressor HLJ1 through JNK/JunD pathway	Chen et al., 2008
Human colon cancer	Inhibition of migration	Inhibition of neurotensin- mediated activator protein-1 and NF-κB activation, and suppression of neurotensin- stimulated IL-8 gene induction	Wang et al., 2006
Prostate cancer	Inhibition of invasion	Downregulation of MMP-2 and MMP-9	Hong et al., 2006
Human fribosarcoma	Inhibition of migration and invasion	Downregulation of MMP-2, MMP-9, uPA, MT1-MMP, and TIMP-2	Yodkeeree et al., 2008

Table 5. Antimetastatic properties of curcumin

Several studies have investigated the molecular mechanisms of cancer cell survival during metastasis. Survival of primary cancer cells in the circulation is a key factor determining its metastatic ability. In general, most adherent cells undergo apoptosis when detached due to improper environmental conditions. This detachment-induced apoptosis or anoikis which is often impaired in metastatic cancers (Mehlen and Puisieux, 2006) has increasingly been recognized as an important mechanism for controlling cancer cell dissemination and invasion to secondary sites. A recent study by our group has shown that curcumin can sensitize non-small cell lung cancer cells to anoikis (Pongrakhananon et al., 2010) through a mechanism that involves Bcl-2 downregulation through ubiquitin-proteasomal degradation. This process is dependent on ROS generation, partcularly superoxide anion which mediates the Bcl-2 degradation process, consistent with the previous findings on the pro-oxidant properties of curcumin (Bhaumik et al., 1999; Khar et al., 2001; Wang et al., 2008).

3.6 Animal studies

Several animal studies have been reported on the anticancer and chemopreventive effects of curcumin in various cancer types. Since the *in vivo* chemopreventive effect of curcumin has earlier been described under 3.1.1 and summarized in Table 1, we will focus on the direct *in vivo* anticancer properties of curcumin.

The anticancer property of curcumin in prostate cancer was investigated by using prostate cancer cells implanted into nude mice (Dorai et al., 2001). Dietary curcumin at the concentration of 2% was given to the mice, and after 6 weeks of treatment the animals were examined for tumor growth, apoptosis, and vascularity. The results showed that curcumin was able to decrease tumor volume, increase cancer cell apoptosis, and inhibit vascular angiogenesis as indicated by the reduction in microvessel density. A similar study using a murine xenograft model of human lung carcinoma cells was reported (Su et al., 2010). In this study, NCI-H460 cells were implanted subcutaneously into nude mice, and curcumin (30 and 45 mg/kg of bodyweight) was intraperitoneally injected into the mice every 4 days after the tumor reached 100 mm³ in size. Curcumin was shown to significantly decrease the tumor size as compared to non-treated control.

The antitumor property of curcumin-encapsulated nanoparticles was investigated in the xenograft mouse model of human pancreatic cancer (Bisht et al., 2010). The nanoparticle formulation was injected twice daily for 3 weeks into the xenografted mice. Plasma concentration of curcumin was sustained in the treated mice at Tmax of 2.75 ± 1.50 h and Cmax of $17,176 \pm 5,176$ ng/ml. Tumor volume was substantially decreased in curcumin treated group. A greater antitumor effect was observed when curcumin was combined with gemcitabine. Curcumin also exhibited antimetastatic activity which was greatly enhanced by the combination treatment. The underlying mechanism of curcumin action involves NF- κ B activation and downregualtion of cyclin D1 and MMP-9.

Curcumin also improves the therapeutic activity of paclitaxel in breast cancer (Aggarwal et al., 2005). Since most metastatic breast cancers acquire apoptosis resistance to paclitaxel, which is the first line therapy for breast cancer, a combination therapy with apoptosissensitizing agents such as curcumin could be beneficial. In a mouse model of breast cancer metastasis, curcumin was shown to decrease the incidence of breast cancer metastasis to the lung as compared to paclitaxel treatment alone. Tissue sections from treated animals showed that NF- κ B, MMP-9, and COX-2 expression were increased in the paclitaxel-treated group but suppressed in the curcumin co-treatment group, supporting the ability of curcumin to abrogate paclitaxel resistance in this metastatic breast cancer model.

3.7 Clinical studies

Several clinical studies are ongoing to investigate the efficacy and safety of curcumin as a preventive treatment agent for a variety of cancers. In prospective phase I clinical trials, curcumin was shown to be safe even at high doses (Cheng et al., 2001). In this study, patients with premalignant lesions caused by oral leukoplakia, intestinal metaplasia, uterine cervical intraepithelial neoplasia, skin Bowen's disease, and bladder cancer were given a curcumin tablet which was taken orally for the period of three months at the daily dose of 500, 1000, 2000, 4000, and 8000 mg. No toxicity was observed in these patients even the highest dose (8000 mg/day). However, this dose was unacceptable by the patients due to its bulky volume. Peak serum concentration of curcumin after the 4000 mg administration was $0.51\pm0.11 \mu$ M, and $0.63\pm0.06 \mu$ M and $1.77\pm1.87 \mu$ M respectively after the 6000 mg and 8000 mg dosing. A similar study showed that a single dose of curcumin up to 12,000 mg had no

dose-limiting toxic effect in healthy volunteers, not even minor adverse effects such as diarrhea (Lao et al., 2006).

Expanding from the above findings, another phase I clinical study was conducted in patients with colon adenocarcinoma to investigate the phamacodynamic of curcumin (Sharma et al., 2004). Curcuminoid, formulated as 500 mg in soft gelatin capsule containing 450 mg of curcumin, 40 mg of desmethoxycurcumin, and 10 mg of bisdesmethoxycurcumin, was taken orally at the dose of 450, 900, 1800, and 3600 mg/day of curcumin for 4 months. Since curcumin is known to induce glutathione S-transferase (GST), suppress prostaglandin E2 (PGE2) production, and inhibit oxidative DNA adduct (M1G) formation, these biomarkers are frequently used to indicate curcumin efficacy. Curcumin and its metabolites were collected from plasma, urine, and feces, and analyzed to assess the pharmacokinetic parameters. The results showed that curcumin was well tolerated by the patients without a dose-limiting toxicity, except in a few cases where patients reported a minor gastrointestinal upset. The result also showed that curcumin at the dose of 3600 mg/day was suitable for phase II evaluation. Curcumin was shown to inhibit PGE2 without affecting GST and M1G, suggesting that GST and M1G may not be useful as indicators for curcumin efficacy. Furthermore, curcumin and its glucuronide and sulfate metabolites were found in the plasma and urine. The presence of these metabolites at all time points indicates that curcumin has poor systemic availability when given orally.

Consistent with the above finding, numerous other studies have demonstrated low systemic bioavailability of curcumin resulting from poor absorption, rapid metabolism, and rapid systemic elimination (Hsu et al., 2007; Ireson et al., 2001; Maiti et al., 2007; Garcea et al., 2004). Glucuronide and sulfate metabolites of curcumin are rapidly detected in the peripheral and portal circulation after curcumin administration (Garcea et al., 2004). In this study, patients with liver metastasis from colorectal adenocarcinoma were administered orally with curcumin at the daily dose of 450, 1800, and 3600 mg for a week. No curcumin or its metabolites was detected in the bile or hepatic tissue, indicating that curcumin is not suitable for treating patients with tumors distant from the absorption site.

In a phase II clinical study conducted in patients with advanced pancreatic cancer, curcumin was administered orally at the dose of 8 g/day for 8 weeks (Dhillon et al., 2008). The treatment was well tolerated by the patients with no systemic side effects, while effectively reducing the tumor size and the activation of NF- κ B and COX-2. Mechanistically, curcumin induces cancer cell apoptosis through an upregulation of p53 in the tumor tissues (He et al., 2011).

4. Conclusion

Curcumin has a great potential in cancer therapy and is gaining wide acceptance as a preventive treatment agent due to its safety. It affects multiple steps in the carcinogenic process, which is important in avoiding chemoresistance. Clinical studies have indicated its efficacy as a single agent or in combination therapy; however, more rigorous testing are needed. Furthermore, problems associated with the low bioavailability of curcumin, including poor absorption, rapid metabolism, and limited tissue distribution, must be addressed. Current strategies that have been investigated to overcome these problems include alternative administration routes, chemical modifications, and various drug delivery and formulation strategies. These strategies will likely benefit the development of curcumin as an anticancer agent.

5. Acknowledgment

This work was supported by NIH grants R01-HL076340, R01-HL076340-04S1, and R01-HL095579.

6. References

- Aggarwal, S., Takada, Y., Singh, S., Myers, J.N., & Aggarwal, B.B. (2004) Inhibition of growth and survival of human head and neck squamous cell carcinoma cells by curcumin via modulation of nuclear factor-kappaB signaling. *International Journal of Cancer*, Vol. 111, No. 5, (September 2004), pp. 679-692, ISSN 1097-2015
- Aggarwal, B.B., Shishodia, S., Takada, Y., Banerjee, S., Newman, R.A., Bueso-Ramos, C.E., & Price, J.E. (2005) Curcumin suppresses the paclitaxel-induced nuclear factorkappaB pathway in breast cancer cells and inhibits lung metastasis of human breast cancer in nude mice. *Clinical Cancer Research*, Vol. 11, No. 20, (October 2005), pp. 7490-7498, ISSN 1078-0432
- Amanda, M.G., & Robert, A.O. (2008) Curcumin and resveratrol inhibit nuclear factorkappaB-mediated cytokine expression in adipocytes. *Nutrition and Metabolism*, Vol. 12, No. 5, (June 2008), pp. 1-13, ISSN 1743-7075
- Anto, R.J., George, J., Babu, K.V., Rajasekharan, K.N., & Kuttan, R. (1996) Antimutagenic and anticarcinogenic activity of natural and synthetic curcuminoids. *Mutation Research*, Vol. 370, No. 2, (September 1996), pp. 127-131, ISSN 1383-5742
- Anto, R.J., Mukhopadhyay, A., Denning, K., & Aggarwal, B.B. (2002). Curcumin (diferuloylmethane) induces apoptosis through activation of caspase-8, BID cleavage and cytochrome c release: Its suppression by ectopic expression of bcl-2 and bcl-xl. *Carcinogenesis*, Vol. 23, No. 1, (January 2002), pp. 143-150, ISSN 0143-3334
- Atsumi, T., Tonosaki, K., & Fujisawa, S. (2006). Induction of early apoptosis and ROSgeneration activity in human gingival fibroblasts (HGF) and human submandibular gland carcinoma (HSG) cells treated with curcumin. Archives of Oral Biology, Vol. 51, No. 10, (October 2006), pp. 913-921, ISSN0003-9969
- Azuine, M.A., Kayal, J.J., & Bhide, S.V. (1992) Protective role of aqueous turmeric extract against mutagenicity of direct-acting carcinogens as well as benzopyrene-induced genotoxicity and carcinogenicity. *Journal of Cancer Research and Clinical Oncology*, Vol. 118, No. 6, pp. 447-452, ISSN 1432-1335
- Baldin, V., Lukas, J., Marcote, M.J., Pagano, M. & Draetta, G. (1993) Cyclin D1 is a nuclear protein required for cell cycle progression in G1. *Genes & Development*, Vol. 7, No. 5, (May 1993), pp. 812-21, ISSN1549-5477
- Bava, S.V., Puliappadamba, V.T., Deepti, A., Nair, A., Karunagaran, D., & Anto, R.J. (2005). Sensitization of taxol-induced apoptosis by curcumin involves down-regulation of nuclear factor-kappaB and the serine/threonine kinase akt and is independent of tubulin polymerization. *The Journal of Biological Chemistry*, Vol. 280, No. 8, (February 2005), pp. 6301-6308, ISSN 1083-351X
- Bhaumik, S., Anjum, R., Rangaraj, N., Pardhasaradhi, B.V.V., & Khar, A. (1999) Curcumin mediated apoptosis in AK-5 tumor cells involves the production of reactive oxygen

intermediates. FEBS Letters, Vol. 456, No. 2, (August 1999), pp.311-314, ISSN 0014-5793

- Bisht, S., Mizuma, M., Feldmann, G., Ottenhof, N.A., Hong, S.M., Pramanik, D., Chenna, V., Karikari, C., Sharma, R., Goggins, M.G., Rudek, M.A., &Maitra, A. (2010). Systemic administration of polymeric nanoparticle-encapsulated curcumin (NanoCurc) blocks tumor growth and metastases in preclinical models of pancreatic cancer. *Molecular Cancer Therapeutics*, Vol. 9, No. 8, (August 2010), pp. 2255-2264, ISSN 1535-7163
- Boonrao, M., Yodkeeree, S., Ampasavate, C., Anuchapreeda, S., & Limtrakul, P. (2010). The inhibitory effect of turmeric curcuminoids on matrix metalloproteinase-3 secretion in human invasive breast carcinoma cells. *Archives of Pharmacal Research*, Vol. 33, No. 7, (July 2010), pp. 989-998, ISSN 1976-3786
- Brouet, I., & Ohshima, H. (1995)Curcumin, an anti-tumor promoter and anti-inflammatory agent, inhibits induction of nitric oxide synthase in activated macrophages.*Biochemical and Biophysical Research Communications*, Vol. 206, No. 2, (January 1995), pp. 533-540, ISSN 1090-2104
- Bush, J.A., Cheung, K.J., Jr, & Li, G. (2001). Curcumin induces apoptosis in human melanoma cells through a fas receptor/caspase-8 pathway independent of p53. *Experimental Cell Research*, Vol. 271, No. 2, (December 2001), pp. 305-314, ISSN 0014-4827
- Cai, C.Q., Peng, Y., Buckley, M.T., Wei, J., Chen, F., Liebes, L., Gerald, W.L., Pincus, M.R., Osman, I. & Lee, P. (2008). Epidermal growth factor receptor activation in prostate cancer by three novel missense mutations. *Oncogene*, Vol. 27, No. 22, (January 2008), pp. 3201-3210, ISSN 0950-9232
- Carmeliet, P. (2005). VEGF as a key mediator of angiogenesis in cancer. *Oncology*, Vol. 69, No. Suppl 3, (November 2005), pp. 4-10, ISSN 1423-0232
- Chang, A. (2011). Chemotherapy, chemoresistance and the changing treatment landscape for NSCLC. *Lung Cancer (Amsterdam, Netherlands)*, Vol. 71, No. 1, (January 2011), pp. 3-10, ISSN 0169-5002
- Chanvorachote, P., Pongrakhananon, V., Wannachaiyasit, S., Luanpitpong, S., Rojanasakul, Y., & Nimmanit U (2009) Curcumin sensitizes lung cancer cells to cisplatin-induced apoptosis through superoxide anion-mediated Bcl-2 degradation. *Cancer Investigation*, Vol. 26, No. 6, (July 2009), pp. 624-635, ISSN 1532-4192
- Chatterjee, S.J., & Pandey, S. (2011). Chemo-resistant melanoma sensitized by tamoxifen to low dose curcumin treatment through induction of apoptosis and autophagy. *Cancer Biology & Therapy*, vol. 11, No. 2, (January 2011), pp. 216-228, ISSN 1555-8576
- Chen, H., Zhang, Z.S., Zhang, Y.L., & Zhou, D.Y. (1999). Curcumin inhibits cell proliferation by interfering with the cell cycle and inducing apoptosis in colon carcinoma cells. *Anticancer Research*, Vol. 19, No.5A, (Septober-October 1999), pp. 3675-3680, ISSN 1791-7530
- Chen, H.W., Lee, J.Y., Huang, J.Y., Wang, C.C., Chen, W.J., Su, S.F., Huang, C.W., Ho, C.C., Chen, J.J., Tsai, M.F., Yu, S.L., & Yang, P.C. (2008) Curcumin inhibits lung cancer cell invasion and metastasis through the tumor suppressor HLJ1. *Cancer Research*, Vol. 68, No. 18, (September 2008), pp. 7428-7438, ISSN 1538-7445

- Chen, Q.Y., Lu, G.H., Wu, Y.Q., Zheng, Y., Xu, K., Wu, L.J., Jiang, Z.Y., Feng, R., & Zhou, J.Y. (2010) Curcumin induces mitochondria pathway mediated cell apoptosis in A549 lung adenocarcinoma cells. *Oncology Report*, Vol. 23, No. 5, (May 2010), pp. 1285-1292, ISSN 1792- 2431
- Cheng, A.L., Hsu, C.H., Lin, J.K., Hsu, M.M., Ho, Y.F., Shen, T.S., Ko, J.Y., Lin, J.T., Lin, B.R., Ming-Shiang, W., Yu, H.S., Jee, S.H., Chen, G.S., Chen, T.M., Chen, C.A., Lai, M.K., Pu, Y.S., Pan, M.H., Wang, Y.J., Tsai, C.C., & Hsieh, C.Y. (2001) Phase I clinical trial of curcumin, a chemopreventive agent, in patients with high-risk or pre-malignant lesions. *Anticancer Research*, Vol. 21, No. 4B, (July-August 2001), pp. 2895-2900, ISSN 1791-7530
- Cheng, C.Y., Lin, Y.H., & Su, C.C. (2010). Curcumin inhibits the proliferation of human hepatocellular carcinoma J5 cells by inducing endoplasmic reticulum stress and mitochondrial dysfunction. *International Journal of Molecular Medicine*, Vol. 26, No. 5, (November 2010), pp. 673-678, ISSN 1791-244X
- Chignell, C.F., Bilski, P., Reszka, K.J., Motten, A.G., Sik, R.H., & Dahl, T.A. (1994). Spectral and photochemical properties of curcumin. *Photochemistry and Photobiology*, Vol. 59, No. 3, (March 1994), pp. 295-302, ISSN 1751-1097
- Chiu, T.L., & Su, C.C. (2009) Curcumin inhibits proliferation and migration by increasing the Bax to Bcl-2 ratio and decreasing NF-kappaBp65 expression in breast cancer MDA-MB-231 cells. *International Journal of Molecular Medicine*, Vol. 23, No. 4, (April 2009), pp. 469-475, ISSN 1791- 244X
- Choudhuri, T., Pal, S., Agwarwal, M.L., Das, T., & Sa, G. (2002). Curcumin induces apoptosis in human breast cancer cells through p53-dependent bax induction. *FEBS Letters*, Vol. 512, No.1-3, (February 2002), pp. 334-340, ISSN 0014-5793
- Chuang, S.E., Cheng, A.L., Lin, J.K., & Kuo, M.L. (2000). Inhibition by curcumin of diethylnitrosamine-induced hepatic hyperplasia, inflammation, cellular gene products and cell-cycle-related proteins in rats.*Food and Chemical Toxicology : An International Journal Published for the British Industrial Biological Research Association*, Vol. 38, No. 11, (November 2000), pp. 991-995, ISSN 1873-6351
- Chuang, S.E., Yeh, P.Y., Lu, Y.S., Lai, G.M., Liao, C.M., Gao, M., & Cheng, A.L. (2002) Basal levels and patterns of anticancer drug-induced activation of nuclear factor-kappaB (NF-kappaB), and its attenuation by tamoxifen, dexamethasone, and curcumin in carcinoma cells. *Biochemical Pharmacology*, Vol. 63, No. 9, (May 2002), pp. 1709-1716, ISSN 1873-2968
- Colotta, F., Allavena, P., Sica, A., Garlanda, C., &Mantovani, A. (2009)Cancer-related inflammation, the seventh hallmark of cancer: links to genetic instability. *Carcinogenesis*, Vol. 7, No. 30, (May 2009), pp. 1073-1081, ISSN 1460-2180
- Devasena, T., Rajasekaran, K.N., Gunasekaran, G., Viswanathan, P., & Menon, V.P. (2003)Anticarcinogenic effect of bis-1,7-(2-hydroxyphenyl)-hepta-1,6-diene-3,5dione a curcumin analog on DMH-induced colon cancer model. *Pharmacological Research*, Vol. 47, No. 2, (Febrauary 2003), pp. 133-140, ISSN 1096-1186
- Dhillon, N., Aggarwal, B.B., Newman, R.A., Wolff, R.A., Kunnumakkara, A.B., Abbruzzese, J.L., Ng, C.H., Badmaev, E., & Kurzrock, R. (2008). Phase II trial of curcumin in patients with advanced pancreatic cancer. *Clinical Cancer Research : An Official*

Journal of the American Association for Cancer Research, Vol. 14, No. 14, (July 2008), pp. 4491-4499, ISSN 1557-3265

- Diehl, J.A. (2002) Cycling to cancer with cyclin D1. *Cancer Biology & Therapy*, Vol. 1, No. 3, (May-June 2002), pp. 226-231, ISSN 1555-8576
- Dorai, T., Gehani, N., & Katz, A. (2000). Therapeutic potential of curcumin in human prostate cancer. II. curcumin inhibits tyrosine kinase activity of epidermal growth factor receptor and depletes the protein. *Molecular Urology*, Vol. 4, No. 1, pp. 1-6, ISSN 1091-5362
- Dorai, T., Cao, Y.C., Dorai ,B., Buttyan, R., &Katz, A.E. (2001) Therapeutic potential of curcumin in human prostate cancer. III. Curcumin inhibits proliferation, induces apoptosis, and inhibits angiogenesis of LNCaP prostate cancer cells in vivo. *The prostate*, Vol. 47, No. 4, (June 2001), pp. 293-303, ISSN 1097-0045
- Evan, G.I., & Vousden, K.H. (2001) Proliferation, cell cycle and apoptosis in cancer. *Nature*, Vol. 6835, No. 411, (May 2001), pp. 342-348, ISSN 1476-4687
- Fidler, I.J. (2003) The pathogenesis of cancer metastasis: the seed and soil hypothesis revisited. *Nature Reviews Cancer*, Vol. 3, No.6, (June 2003), pp. 453-458, ISSN 1474-1768
- Ganta, S., &Amiji, M. (2009) Coadministration of paclitaxel and curcumin in nanoemulsion formulations to overcome multidrug resistance in tumor cells. *Molecular Pharmacology*, Vol. 6, No. 3, (May-June 2009), pp. 928-939, ISSN 1543-8392
- Garcea, G., Jones, D.J., Singh, R., Dennison, A.R., Farmer, P.B., Sharma, R.A., Steward, W.P., Gescher, A.J., & Berry, D.P. (2004). Detection of curcumin and its metabolites in hepatic tissue and portal blood of patients following oral administration. *British Journal of Cancer*, Vol. 90, No. 5, (March 2004), pp. 1011-1015, ISSN 0007-0920
- Gupta, G.P., & Massague, J. (2006). Cancer metastasis: Building a framework. *Cell*, Vol. 127, No. 4, (November 2006), pp. 679-695, ISSN 0092-8674
- Han, S.S., Chung, S.T., Robertson, D.A., Ranjan, D., & Bondada, S. (1999) Curcumin causes the growth arrest and apoptosis of B cell lymphoma by downregulation of egr-1, cmyc, bcl-XL, NF- kappa B, and p53. *Clinical Immunology*, Vol. 93, No. 2, (November 1999), pp. 152-161, ISSN 1521-7035
- Hanahan, D.,& Weinberg, R.A. (2000) The Hallmarks of cancer. *Cell*, Vol. 1, No. 100, (January 2000), pp. 57-70, ISSN0092-8674
- He, Z.Y., Shi, C.B., Wen, H., Li, F. L., Wang, B.L., & Wang, J. (2011). Upregulation of p53 expression in patients with colorectal cancer by administration of curcumin. *Cancer Investigation*, Vol. 29, No. 3, (March 2011), pp. 208-213, ISSN 0735-7907
- Hengartner, M.O. (2000) The biochemistry of apoptosis. *Nature*, Vol. 407, No. 6805, (October 2000), pp. 770–776, ISSN 0028-0836
- Himelstein, B.P., Canete-Soler, R., Bernhard, E.J., Dilks, D.W., & Muschel, R.J. (1994). Metalloproteinases in tumor progression: The contribution of MMP-9. *Invasion & Metastasis*, Vol. 14, No. 1-6, pp. 246-258, ISSN 0251-1789
- Hong, J.H., Ahn, K.S., Bae, E., Jeon, S.S., & Choi, H.Y. (2006) The effects of curcumin on the invasiveness of prostate cancer in vitro and in vivo. *Prostate Cancer and Prostatic Diseases*, Vol. 9, No. 2, (January 2006), pp. 147-52, ISSN 1476-5608
- Howells, L.M., Sale, S., Sriramareddy, S.N., Irving, G.R., Jones, D.J., Ottley, C.J., Pearson, D.G., Mann, C.D., Manson, M.M., Berry, D.P., Gescher, A., Steward, W.P., &Brown,

K. (2010). Curcumin ameliorates oxaliplatin-induced chemoresistance in HCT116 colorectal cancer cells in vitro and in vivo. *International Journal of Cancer. Journal International Du Cancer*, (Septemper 2010), ISSN 1097-0215

- Hsu, C.H., & Cheng, A.L. (2007) Clinical studies with curcumin. *Advances in Experimental Medicine and Biology*, Vol. 595, pp. 471-480, ISSN 0065-2598
- Hussain, A.R., Al-Rasheed, M., Manogaran, P.S., Al-Hussein, K.A., Platanias, L.C., Al Kuraya, K., & Uddin, S. (2006). Curcumin induces apoptosis via inhibition of PI3'kinase/AKT pathway in acute T cell leukemias. *Apoptosis : An International Journal* on Programmed Cell Death, Vol. 11, No. 2, (February 2006), pp. 245-254, ISSN 1573-675X
- Ikezaki, S., Nishikawa, A., Furukawa, F., Kudo, K., Nakamura, H., Tamura, K., & Mori, H. (2001)Chemopreventive effects of curcumin on glandular stomach carcinogenesis induced by N-methyl-N'-nitro-N-nitrosoguanidine and sodium chloride in rats. *Anticancer Research*, Vol. 21, No. 5, (September-October 2001), pp. 3407-3411, ISSN 1791-7530
- Ireson, C., Orr, S., Jones, D.J., Verschoyle, R., Lim, C.K., Luo, J.L, Howells, L., Plummer, S., Jukes, R., Williams, M., Steward, W.P., &Gescher, A. (2001). Characterization of metabolites of the chemopreventive agent curcumin in human and rat hepatocytes and in the rat in vivo, and evaluation of their ability to inhibit phorbol esterinduced prostaglandin E2 production. *Cancer Research*, Vol. 61, No. 3, (February 2001), pp. 1058-1064, ISSN 1538-7445
- Jagetia, G.C., & Rajanikant, G.K. (2005) Curcumin treatment enhances the repair and regeneration of wounds in mice exposed to hemibody gamma irradiation. *Plastic and Reconstructive Surgery*, Vol. 115, No. 2, (February 2005), pp. 515-528, ISSN 1529-4242
- Kakar, S. S., & Roy, D. (1994). Curcumin inhibits TPA induced expression of c-fos, c-jun and c-myc proto-oncogenes messenger RNAs in mouse skin. *Cancer Letters*, Vol. 87, No. 1, (November 1994), pp. 85-89, ISSN 0304-3835
- Karin, M., Cao, Y., Greten, F.R., & Li, Z.W. (2002)NF-kB in cancer: from innocent bystander to major culprit. *Nature Reviews Cancer*, Vol. 4, No. 2, (April 2002), pp. 301-310, ISSN 1474-1768
- Karmakar, S., Banik, N.L., Patel, S.J., & Ray, S.K. (2006). Curcumin activated both receptormediated and mitochondria-mediated proteolytic pathways for apoptosis in human glioblastoma T98G cells. *Neuroscience Letters*, Vol. 407, No. 1, (October 2006), pp. 53-58, ISSN 0304-3940
- Kastan, M.B., & Bartek, J. (2004) Cell-cycle checkpoints and cancer. *Nature*, Vol. 432, No. 7015, (November 2004), pp. 316-23, ISSN 1476-4687
- Khar, A., Ali, A.M., Pardhasaradhi, B.V.V., Varalakshmi, C., Anjum, R., & Kumari, A.L. (2001) Induction of stress response renders human tumor cell lines resistant to curcumin-mediated apoptosis: role of reactive oxygen intermediates. *Cell Stress Chaperones*, Vol. 6, No. 4, (October 2006), pp. 368-376, ISSN1466-1268
- Knudsen, K.E., Diehl, J.A., Haiman, C.A., & Knudsen, E.S. (2006). Cyclin D1: Polymorphism, aberrant splicing and cancer risk. *Oncogene*, Vol. 25, No. 11, (March 2006), pp. 1620-1628, ISSN1476-5594

- Korutla, L., & Kumar, R. (1994). Inhibitory effect of curcumin on epidermal growth factor receptor kinase activity in A431 cells. *Biochimica Et Biophysica Acta*, Vol. 1224, No. 3, (December 1994), pp. 597-600. ISSN 0006-3002
- Lao, C.D., Ruffin, M.T. 4th, Normolle, D., Heath, D.D., Murray, S.I., Bailey, J.M., Boggs, M.E., Crowell, J., Rock, C.L., & Brenner, D.E. (2006) Dose escalation of a curcuminoid formulation.*BMC Complementary and Alternative Medicine*, Vol. 12, No. 6, (March 2006), pp. 10, ISSN 1472-6882
- Lavrik, I.N., Golks, A., & Krammer, P.H. (2005) Caspases: pharmacological manipulation of cell death. *The Journal of Clinical Investigation*, Vol. 115, No. 10, (October 2005), pp. 2665-2672, ISSN 1558-8238
- Lee, K.W., Kim, J.H., Lee, H.J., & Surh, Y.J. (2005) Curcumin inhibits phorbol ester-induced upregulation of cyclooxygenase-2 and matrix metalloproteinase-9 by blocking ERK1/2 phosphorylation and NF-kappaB transcriptional activity in MCF10A human breast epithelial cells. *Antioxidant and Redox Signaling*, Vol. 7, No. 11-12, (November-December 2005), pp. 1612-1620, ISSN 1523-0864
- Li, Y., Revalde, J.L., Reid, G., & Paxton, J.W. (2010). Modulatory effects of curcumin on multi-drug resistance-associated protein 5 in pancreatic cancer cells. *Cancer Chemotherapy and Pharmacology*,(November 2010), ISSN 1432-0843
- Limtrakul, P., Anuchapreeda, S., Lipigorngoson, S., & Dunn, F.W. (2001). Inhibition of carcinogen induced c-ha-ras and c-fos proto-oncogenes expression by dietary curcumin. *BMC Cancer*, Vol. 1, No. 1, (Junuary 2011), ISSN 1471-2407
- Lin, S.S., Lai, K.C., Hsu, S.C., Yang, J.S., Kuo, C.L., Lin, J.P., Ma, Y.S., Wu, C.C., & Chung, J.G. (2009) Curcumin inhibits the migration and invasion of human A549 lung cancer cells through the inhibition of matrix metalloproteinase-2 and -9 and vascular endothelial growth factor (VEGF). *Cancer Letters*, Vol. 285, No. 2, (May 2009), pp. 127-133, ISSN 1872-7980
- Liu, E., Wu, J., Cao, W., Zhang, J., Liu, W., Jiang, X., & Zhang, X. (2007). Curcumin induces G2/M cell cycle arrest in a p53-dependent manner and upregulates ING4 expression in human glioma. *Journal of Neuro-Oncology*, Vol. 85, No. 3, (December 2007), pp. 263-270, ISSN 1573-7373
- Lu, Y.P., Chang, R.L., Huang, M.T., & Conney, A.H. (1993). Inhibitory effect of curcumin on 12-O-tetradecanoylphorbol-13-acetate-induced increase in ornithine decarboxylase mRNA in mouse epidermis.*Carcinogenesis*, Vol. 14, No. 2, (February 1993), pp. 293-297, ISSN 1460-2180
- Mai, S.,& Mushinski, J.F. (2003) c-Myc-induced genomic instability. *Journal of Environmental Pathology, Toxicology and Oncology*, Vol. 22, No. 3, pp. 179-199, ISSN 0731-8898
- Maiti, K., Mukherjee, K., Gantait, A., Saha, B.P., & Mukherjee, P.K. (2007). Curcuminphospholipid complex: Preparation, therapeutic evaluation and pharmacokinetic study in rats. *International Journal of Pharmaceutics*, Vol. 330, No. 1-2, (February 2007), pp. 155-163, ISSN 0378-5173
- Mantovani, A., Allavena, P., Sica, A., & Balkwill, F. (2008) Cancer-related inflammation. *Nature*, Vol. 7203, No. 454, (July 2008), pp. 436-444, ISSN 1476-4687
- Marin, Y.E., Wall, B.A., Wang, S., Namkoong, J., Martino, J.J., Suh, J., Lee, H.J., Rabson, A.B., Yang, C.H., Chen, S., & Ryu, J.H. (2007). Curcumin downregulates the constitutive

activity of NF-kappaB and induces apoptosis in novel mouse melanoma cells. *Melanoma Research*, Vol. 17, No. 5, (October 2007), pp. 274-283, ISSN 1473-5636

- Mehlen, P., & Puisieux, A. (2006) Metastasis: a question of life or death. *Nature Reviews Cancer*, Vol. 6, No. 6, (June 2006), pp. 449-458, ISSN 1474-1768
- Moussavi, M., Assi, K., Gómez-Muñoz, A., & Salh, B. (2006) Curcumin mediates ceramide generation via the de novo pathway in colon cancer cells. *Carcinogenesis*, Vol. 27, No. 8, (August 2006), pp. 1636-1644, ISSN 1460-2180
- Mukhopadhyay, A., Banerjee, S., Stafford, L.J., Xia, C., Liu, M., & Aggarwal, B.B. (2002). Curcumin-induced suppression of cell proliferation correlates with downregulation of cyclin D1 expression and CDK4-mediated retinoblastoma protein phosphorylation. *Oncogene*, Vol. 21, No. 57, (December 2002), pp. 8852-8861, ISSN 1476-5594
- Notarbartolo, M., Poma, P., Perri, D., Dusonchet, L., Cervello, M., & D'Alessandro, N. (2005) Antitumor effects of curcumin, alone or in combination with cisplatin or doxorubicin, on human hepatic cancer cells. analysis of their possible relationship to changes in NF-kB activation levels and in IAP gene expression. *Cancer Letters*, Vol. 224, No. 1, (June 2005), pp. 53-65, ISSN 1872- 7980
- Park, C., Kim, G.Y., Kim, G.D., Choi, B.T., Park, Y.M., & Choi, Y.H. (2006) Induction of G2/M arrest and inhibition of cyclooxygenase-2 activity by curcumin in human bladder cancer T24 cells. *Oncology Reports*, Vol. 15, No. 5, (May 2006), pp. 1225-1231, ISSN 1791-2431
- Pereira, M.A., Grubbs, C.J., Barnes, L.H., Li, H., Olson, G.R., Eto, I., Juliana, M., Whitaker, L.M., Kelloff, G.J., Steele, V.E., & Lubet, R.A. (1996). Effects of the phytochemicals, curcumin and quercetin, upon azoxymethane-induced colon cancer and 7,12dimethylbenz[a]anthracene-induced mammary cancer in rats. *Carcinogenesis*, Vol. 17, No. 6, (June 1996), pp. 1305-1311, ISSN 1460-2180
- Perry, M.C., Demeule, M., Regina, A., Moumdjian, R., & Beliveau, R. (2010). Curcumin inhibits tumor growth and angiogenesis in glioblastoma xenografts. *Molecular Nutrition & Food Research*, Vol. 54, No. 8, (August 2010), pp. 1192-1201, ISSN 1613-4133
- Philip, S., Bulbule, A., & Kundu, G.C. (2004) Matrix metalloproteinase-2: mechanism and regulation of NF-kappaB-mediated activation and its role in cell motility and ECMinvasion. *Glycoconjugate Journal*, Vol. 21, No. 8-9, (November 2004), pp. 429-441, ISSN 1573-4986
- Pongrakhananon, V., Nimmannit, U., Luanpitpong, S., Rojanasakul, Y., & Chanvorachote, P. (2010) Curcumin sensitizes non-small cell lung cancer cell anoikis through reactive oxygen species- mediated Bcl-2 downregulation. *Apoptosis*, Vol. 15, No. 5, (May 2010), pp. 574-585, ISSN 1360- 8185
- Prakobwong, S., Gupta, S.C., Kim, J.H., Sung, B., Pinlaor, P., Hiraku, Y., Wongkham, S., Sripa, B., Pinlaor, S., & Aggarwal, B.B. (2011). Curcumin suppresses proliferation and induces apoptosis in human biliary cancer cells through modulation of multiple cell signaling pathways. *Carcinogenesis*, (February 2011), ISSN1460-2180
- Prasad, N.S., & Sarasija, S. (1997) Spectrophotometric estimation of curcumin. *Indian Drugs*, vol. 34, pp. 227-228, ISSN 0019-462X

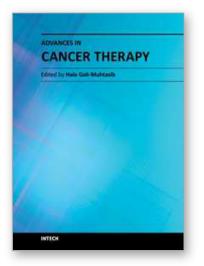
- Prescott, S.M., & Fitzpatrick, F.A. (2000) Cyclooxygenase-2 and carcinogenesis. *Biochimica et biophysica acta*, Vol. 1470, No. 2, (March 2000), pp. M69-78, ISSN 0006-3002
- Rajalingam, K., Schreck, R., Rapp, U.R., & Albert, S. (2007) Ras oncogenes and their downstream targets. *Biochimica et Biophysica Acta*, Vol. 1773, No. 8, (January 2007), pp. 1177-1195, ISSN 0006-3002
- Romanska, H.M., Polak, J.M., Coleman, R.A., James, R.S., Harmer, D.W., Allen, J.C., & Bishop, A.E. (2002)iNOS gene upregulation is associated with the early proliferative response of human lung fibroblasts to cytokine stimulation. *The Journal of Pathology*, Vol. 3, No. 197, (July 2002), pp. 372–379, ISSN 1096-9896
- Samani, A.A., Yakar, S., LeRoith, D., & Brodt, P. (2007) The role of the IGF system in cancer growth and metastasis: overview and recent insights. *Endocrine Reviews*, Vol. 28, No. 1, (Febrauary 2007), pp. 20-47, ISSN 1945-7189
- Shankar, S., & Srivastava, R.K. (2007). Involvement of bcl-2 family members, phosphatidylinositol 3'-kinase/AKT and mitochondrial p53 in curcumin (diferulolylmethane)-induced apoptosis in prostate cancer.*International Journal of Oncology*, Vol. 30, No.4, (April 2007), pp. 905-918, ISSN 1019-6439
- Sharma, C., Kaur, J., Shishodia, S., Aggarwal, B.B., & Ralhan, R. (2006). Curcumin down regulates smokeless tobacco-induced NF-kappaB activation and COX-2 expression in human oral premalignant and cancer cells. *Toxicology*, Vol. 228, No. 1, (November 2006), pp. 1-15, ISSN 0300-483X
- Sharma, R.A., Euden, S.A., Platton, S.L., Cooke, D.N., Shafayat, A., Hewitt, H.R., Marczylo, T.H., Morgan, B., Hemingway, D., Plummer, S.M., Pirmohamed, M., Gescher, A.J., & Steward, W.P. (2004) Phase I clinical trial of oral curcumin: biomarkers of systemic activity and compliance. *Clinical Cancer Research*, Vol. 15, No. 20, (October 2004), pp. 6847-6854, ISSN 1078-0432
- Shishodia, S., Potdar, P., Gairola, C.G., & and Aggarwal, B.B. (2003)Curcumin (diferuloylmethane) down-regulates cigarette smoke-induced NF-kBactivation through inhibition of IkBa kinase in human lung epithelial cells:correlation with suppression of COX-2, MMP-9 and cyclin D1. *Carcinogenesis*, Vol.24, No.7, pp.1269-1279, ISSN 1460-2180
- Shishodia, S., Amin, H.M., Lai, R., & Aggarwal, B.B. (2005). Curcumin (diferuloylmethane) inhibits constitutive NF-kappaB activation, induces G1/S arrest, suppresses proliferation, and induces apoptosis in mantle cell lymphoma. *Biochemical Pharmacology, Vol. 70, No.* 5, (September 2005), pp. 700-713, ISSN 1873-2968
- Simon, A., Allais, D.P., Duroux, J.L., Basly, J.P., Durand-Fontanier, S., & Delage, C. (1998) Inhibitory effect of curcuminoids on MCF-7 cell proliferation and structure-activity relationships. *Cancer Letters*, Vol. 129, No. 1, (July 1998), pp. 111-116, ISSN 1872-7980
- Singh, M., & Singh, N. (2009) Molecular mechanism of curcumin induced cytotoxicity in human cervical carcinoma cells *Molecular and Cellular Biochemistry*, Vol. 325, No. 1-2, (February 2009), pp. 107-119, ISSN 1573-4919
- Song, G., Mao, Y.B., Cai, Q.F., Yao, L.M., Ouyang, G.L., & Bao, S.D. (2005). Curcumin induces human HT-29 colon adenocarcinoma cell apoptosis by activating p53 and regulating apoptosis-related protein expression. *Brazilian Journal of Medical and Biological Research*, Vol. 38,No. 12, (December 2005), pp. 1791-1798, ISSN 1414-431X

- Squires, M.S., Hudson, E.A., Howells, L., Sale, S., Houghton, C.E., Jones, J.L., Fox, L.H., Dickens, M., Prigent, S.A. & Manson, M.M. (2003). Relevance of mitogen activated protein kinase (MAPK) and phosphotidylinositol-3-kinase/protein kinase B (PI3K/PKB) pathways to induction of apoptosis by curcumin in breast cells. *Biochemical Pharmacology*, Vol. 65, No. 3, (February 2003), pp. 361-376, ISSN1873-2968
- Su, C.C., Yang, J.S., Lu, C.C., Chiang, J.H., Wu, C.L., Lin, J.J., Lai, K.C., Hsia, T.C., Lu, H.F., Fan, M.J., & Chung, G.J. (2010). Curcumin inhibits human lung large cell carcinoma cancer tumour growth in a murine xenograft model. *Phytotherapy Research* : *PTR*, Vol. 24, No. 2, (February 2010), pp. 189-192, ISSN 1099-1573
- Syng-Ai, C., Kumari, A.L., & Khar, A. (2004). Effect of curcumin on normal and tumor cells: Role of glutathione and bcl-2. *Molecular Cancer Therapeutics*, Vol. 3, No. 9, (September 2004), pp. 1101-1108, ISSN 1538-8514
- Szakacs, G., Paterson, J.K., Ludwig, J.A., Booth-Genthe, C. & Gottesman, M.M. (2006) Targeting multidrug resistance in cancer. *Nature Reviews. Drug Discovery*, Vol. 5, No. 3, (March 2006), pp. 219-234, ISSN 1474-1784
- Tharakan, S.T., Inamoto, T., Sung, B., Aggarwal, B.B., & Kamat, A.M. (2010). Curcumin potentiates the antitumor effects of gemcitabine in an orthotopic model of human bladder cancer through suppression of proliferative and angiogenic biomarkers. *Biochemical Pharmacology*, Vol. 79, No. 2, (January 2010), pp. 218-228, ISSN 0006-2952
- Ventura, A., Kirsch, D.G., McLaughlin, M.E., Tuveson, D.A., Grimm, J., Lintault, L., Newman, J., Reczek, E.E., Weissleder, R., & Jacks, T. (2007) Restoration of p53 function leads to tumour regression in vivo. *Nature*, Vol. 445, No.7128, (February 2007), pp. 661-665, ISSN 0028-0836
- Vogelstein, B., & Kinzler, K.W. (2004) Cancer genes and the pathways they control. *Nature Medicine*, Vol. 8, No. 10, (August 2004), pp. 789-799, ISSN 1546-170X
- Wang, L., Chanvorachote, P., Toledo, D., Stehlik, C., Mercer, R.R., Castranova, V., & Rojanasakul, Y. (2008) Peroxide is a key mediator of Bcl-2 down-regulation and apoptosis induction by cisplatin in human lung cancer cells. *Molcular Pharmacology*, Vol. 73, No. 1, (October 2007), pp. 119-127, ISSN 1521-0111
- Wang, X., Wang, Q., Ives, K.L., & Evers, B.M. (2006). Curcumin inhibits neurotensinmediated interleukin-8 production and migration of HCT116 human colon cancer cells. *Clinical Cancer Research : An Official Journal of the American Association for Cancer Research*, Vol. 12, No. 18, (September 2006), pp. 5346-5355, ISSN 1557-3265
- Wang, Y.J., Pan, M.H., Cheng, A.L., Lin, L.I., Ho, Y.S., Hsieh, C.Y., & Lin, J.K. (1997). Stability of curcumin in buffer solutions and characterization of its degradation products. *Journal of Pharmaceutical and Biomedical Analysis*, Vol. 15, No. 12, (August 1997), pp. 1867-1876, ISSN 1873-264X
- Xia, Y., Jin, L., Zhang, B., Xue, H., Li, Q., & Xu, Y. (2007). The potentiation of curcumin on insulin-like growth factor-1 action in MCF-7 human breast carcinoma cells. *Life Sciences*, Vol. 80, No. 23, (May 2007), pp. 2161-2169, ISSN 0024-3205
- Yang, F., Lim, G.P., Begum, A.N., Ubeda, O.J., Simmons, M.R., Ambegaokar, S.S., Chen, P.P., Kayed, R., Glabe, C.G., Frautschy, S.A., & Cole, G.M. (2005) Curcumin inhibits formation of amyloid beta oligomers and fibrils, binds plaques, and reduces

amyloid in vivo. *Journal of Biological Chemistry*, Vol. 280, No. 7, (December 2004), pp. 5892-5901, ISSN 1083-351X

- Yodkeeree, S., Garbisa, S., & Limtrakul, P. (2008). Tetrahydrocurcumin inhibits HT1080 cell migration and invasion via downregulation of MMPs and uPA. *Acta Pharmacologica Sinica*, Vol. 29, No.7, (July 2008), pp. 853-860, ISSN 1745-7254
- Zheng, L.D., Tong, Q.S., & Wu, C.H. (2002). Inhibitory effects of curcumin on apoptosis of human ovary cancer cell line A2780 and its molecular mechanism. *Ai Zheng Chinese Journal of Cancer*, Vol. 21, No. 12, (December 2002), pp. 1296-1300, ISSN 1000-467X





Advances in Cancer Therapy Edited by Prof. Hala Gali-Muhtasib

ISBN 978-953-307-703-1 Hard cover, 568 pages **Publisher** InTech **Published online** 21, November, 2011 **Published in print edition** November, 2011

The book "Advances in Cancer Therapy" is a new addition to the Intech collection of books and aims at providing scientists and clinicians with a comprehensive overview of the state of current knowledge and latest research findings in the area of cancer therapy. For this purpose research articles, clinical investigations and review papers that are thought to improve the readers' understanding of cancer therapy developments and/or to keep them up to date with the most recent advances in this field have been included in this book. With cancer being one of the most serious diseases of our times, I am confident that this book will meet the patients', physicians' and researchers' needs.

How to reference

In order to correctly reference this scholarly work, feel free to copy and paste the following:

Varisa Pongrakhananon and Yon Rojanasakul (2011). Anticancer Properties of Curcumin, Advances in Cancer Therapy, Prof. Hala Gali-Muhtasib (Ed.), ISBN: 978-953-307-703-1, InTech, Available from: http://www.intechopen.com/books/advances-in-cancer-therapy/anticancer-properties-of-curcumin



InTech Europe

University Campus STeP Ri Slavka Krautzeka 83/A 51000 Rijeka, Croatia Phone: +385 (51) 770 447 Fax: +385 (51) 686 166 www.intechopen.com

InTech China

Unit 405, Office Block, Hotel Equatorial Shanghai No.65, Yan An Road (West), Shanghai, 200040, China 中国上海市延安西路65号上海国际贵都大饭店办公楼405单元 Phone: +86-21-62489820 Fax: +86-21-62489821 © 2011 The Author(s). Licensee IntechOpen. This is an open access article distributed under the terms of the <u>Creative Commons Attribution 3.0</u> <u>License</u>, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

IntechOpen

IntechOpen