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Hysteroscopic Endometrial Embryo Delivery (HEED)

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1. Introduction

It has been over 30 years since the first successful pregnancy using in vitro fertilization (IVF). There have been major advancements in the different components of IVF such as ovulation induction protocols, oocyte retrieval techniques, and culture medium tailored to improving embryo quality (Gardner 1998). However, the discrepancy between women undergoing IVF with normal embryo development and live pregnancy rates continues to exist. It is estimated that up to 85% of replaced embryos fail to implant despite the selection of apparently normal embryos for transfer (Sallam 2002). This failure rate suggests that the embryo transfer stage is a key step to successful live pregnancy rates in assisted reproductive technology (ART) (Meldrum 1987).

Embryo transfer is traditionally performed by “blindly” replacing the embryos into the uterine cavity utilizing a transcervical catheter at approximately 2-5 days of development. This technique relies highly on the skill and tactile senses of the clinician. Many clinicians will transfer the embryos at a fixed distance (6 cm) from the external os; however, with varying cervical lengths and uterine anatomy, this often does not ensure optimal placement (Brown 2007). Recently, there have been many studies proposing potential embryo transfer related factors to the low success rate in pregnancy outcomes such as uterine contractions, expulsion of embryos, blood or mucus on the catheter tip, bacterial contamination of the catheter, and retained embryos (Schoolcraft 2001). Ultrasound guided embryo transfer (UGET) is currently suggested as the standard clinical practice and appears to improve the chances of live/ongoing and clinical pregnancies compared with clinical touch methods (Brown 2007). However, controversies still remain regarding the actual benefit of UGET in successful clinical pregnancy rates (Kosmas 1999). The subendometrial embryo delivery (SEED) technique has been previously reported to increase pregnancy rates and eliminate ectopic pregnancies associated with ART (KAMRAVA 2010). In this study, we set out to use a similar technique which utilized a mini-hysteroscope with a flexible catheter for direct delivery of embryo(s) at the 4-12 cell stage onto the endometrium under direct visualization. The hysteroscopic visual guidance ensures more precise and reliable placement at the desired location of the endometrium.

2. Materials and methods

Patients. 35 patients between 22 and 46 years of age undergoing IVF were included in this report. Informed consent was obtained prior to the start of the cycle. Controlled ovarian hyperstimulation was initiated with Follitropin β (Follistim®, Organon Pharmaceuticals, Inc.). Endogenous gonadotropins surge (i.e., the prevention of an LH surge) was controlled with ganirelix acetate (Antagon™, Organon Pharmaceuticals, Inc.). Oocyte retrieval was carried out in an office setting under local anesthesia and mild sedation. Oocytes were fertilized and cultured in a human tubal fluid formulated medium at 37 degrees C and 5% CO₂ in air". Embryos were transferred at 48-72 hours post fertilization (Figure 1a). All women received some type of luteal support, be it progesterone or hCG (3000 IU of hCG at 3 and 6 days post retrieval) (Figure 1b, c). Serum hCG was quantified at 10 days after the last hCG; a concentration of 5 IU/ml with a delayed menses was used as confirmation of pregnancy.

Description of Hysteroscopic Endometrial Embryo Delivery (HEED):

A transvaginal ultrasound of the uterus is performed and the direction and thickness of the endometrial lining is ascertained. With patient in dorsolithotomy position, a bivalved speculum is placed in the vagina and the cervix exposed. Vagina and cervix are washed with modified HAM's solution. Subsequently, 10 cc of 1% xylocaine is injected bilaterally in the utero-sacral nerve endings.

Fig. 1. Mini flexible hysteroscope (Storz®, LA, CA USA)

The cervix is grasped with an allis clamp and stabilized. Nitrogen gas is used as the distention media throughout the procedure via a hysteroscopic insufflator. A 3 mm flexible hysteroscope (Figure 1) loaded with embryo catheter containing the embryos (Figure 2) is then gently inserted through the cervical os under direct visualization of the cervical canal into the uterine cavity. Once the cavity is visualized, it is then further advanced to the fundus of the uterus. The loaded embryo transfer catheter (Precision Reproduction, LA, CA USA) is then advanced to 1.5 cm from the tip of the hysteroscope and placed over the point of embryo deposition, half way between the lowest point of the fundus in the midline and the tubal opening into the uterus (Figure 3). The embryos are then gently released by the embryologist. Our results show that hysteroscopic guided early embryo transfer results in a high pregnancy outcome, 2-3x greater than “blind” transfer technique rates. Direct visualization provides an objective, visually confirmed, replicable technique for embryo transfer. The end result is less operator dependent and in contrast to routine ET techniques in which operator experience may account for the variable overall pregnancy rates (Garcia 2002). Hysteroscopic direct embryo delivery may circumvent many of the known and previously reported embryo transfer related factors associated with poor outcomes. Many of our patients had failed prior IVF-ET attempts due to multiple etiologies.

Fig. 2. The flexible catheter for embryo delivery (Precision Reproduction, LLC. LA, CA USA)

A light weight flexible minihysteroscope was used for visualization of the endometrial cavity (Figure 1d) (Storz®, LA, CA USA). The scope incorporates a flexible distal end of 3mm in diameter with a straight through operating channel. In addition, the optic filter is directly connected to a light source, decreasing the weight of the scope and giving a better

“feel” for the scope. The transfer catheter (Precision Reproduction, LLC, LA, CA USA) is polycarbonate based with a tapered tip (to 500 μm), beveled to 60°.

Fig. 3. Placement of embryo(s) under hysteroscopic guidance; arrow points to the tip of the catheter; catheter entry at 8 o'clock position.

3. Results

35 cycles were started and all had retrievals. 22 cycles involved use of intra-cytoplasmic sperm injection (ICSI) due to male factor problems. Endometrial thickness varied between 7 and 16 mm. 22 cycles had transfers on day 2 and 13 cycles had transfers on day 3. There were 16 positive β hCG's greater than 5 IU/ml twelve days after embryo transfer. Of these, 2 had biochemical pregnancies, and 12 had clinical pregnancies as evidenced by presence of gestational sac by ultrasound examination at five weeks of gestation and presence of the fetus and a heart beat at six weeks of gestation. There were 5 first trimester spontaneous abortions at 7-8 weeks of gestation. Seven(7) patients have delivered healthy babies at term; there were 2 ectopic pregnancies (Table 1).

	Day 2 Transfer average age 3835	Day 3 Transfer average age 38
Total Start	22	13
Cancelled	0	0
Retrievals	22	13
Transfers	22	13
Total + Beta	9	7
Chemical Preg	2	0
Spontaneous ab	3	2
Multiple Preg	3	1
Ectopic preg	1	1
LiveBorn	3(13%)	4(30%)

Table 1. Results from HEED on day2 and day 3 transfers

4. Discussion

As may have been expected, the average age of patients for transfers on day 3 versus day 2 was lower (35 vs. 38 years of age), as they had better quality embryos which made it more feasible to continue embryo culture 1 day longer. Interestingly enough, the live pregnancy rate was also higher in day 3 transfers (31% vs. 15%).

Advantages of hysteroscopic guided direct embryo delivery include objectivity and replicability of the procedure. This unique and significant aspect of the procedure increases the reliability of correct entry into the uterine cavity with direct visual confirmation. Furthermore, placement and subsequent implantation at a precise location, with minimal volume of transfer media, provides an obvious benefit to patients with distorted uterine cavities, myomas, and adenomyosis and uterine adhesions. Visualization also provides the advantage of maneuvering along the contours of the uterus, thus decreasing the rate of trauma to the endometrial lining. In addition, performing gas distension of the uterus by an inert gas (N₂), the catheter tip is less likely to come into contact with the uterine fundus which has been associated with stimulating uterine contractions and creating an unfavorable environment for implantation (Kovacs 1999, Lesny 1998). It has been reported that high frequency uterine contractions are associated with a lower ongoing clinical pregnancy rate and complete expulsion of the embryo (Fanchin 1998). It has also been postulated that the expulsion of the embryo into the lower uterine segment may result in higher rates of cervical ectopic pregnancy and placenta previas (Romundstad 2006; Schoolcraft 2001).

Witnessing uterine contractions hysteroscopically can also guide the clinician to abort and defer the procedure, thus decreasing costs, multiple failed attempts of ET, embryo loss, and risk of cervical ectopics and placenta previas. Direct visualization of the catheter tip ensures that the embryos are not retained in the catheter or lost. Viser et al. found a lower pregnancy rate when retained embryos were present (3% vs. 20.3%). In addition, catheter tip visualization allowed us to deliver smaller aliquot volumes for ET (5µl) as opposed to routine volumes (30µl). Smaller volume allows better handling of the embryo for proper orientation to the uterine lining, stabilizing the position and has been reported to increase pregnancy and implantation rates (Meldrum 1987). It may also contribute to the reduced

ectopic pregnancy rates, as larger volumes have been associated with increased ectopic pregnancy risk (Marcus 1995). Expulsion of this low volume of transfer media, carrying the embryo(s), from the tip of the catheter can only now be verified under direct visualization. In the “blind” procedure there is a real concern that this tiny droplet can be dragged into the lower uterine segment or into the cervical canal or out of the uterus along with the catheter during the final withdrawal of the catheter after embryo transfer.

The potential disadvantage and risk of this technique is disruption of the uterine lining, however the risk is postulated to be less than “blind” and ultrasound guided transfers due to the advantage of direct visualization of the uterine lining and not requiring movement of the catheter to facilitate identification during ultrasound (Garcia-Velasco 2002). In addition, visualization allows one to place the embryo at a different location if trauma ensues. The major drawback to its acceptance is that hysteroscopy is an invasive procedure. However, as opposed to rigid endoscopes which may cause trauma to the uterus, the hysteroscope used in this study is a mini hysteroscope with a 3 mm diameter and flexible tip that allows one to easily follow the curvature of the uterus. The catheter used is semi-rigid to prevent kinkage as it passes through the endoscope yet with flexibility to bend with the endoscope. In our study, no disruption to the uterine lining or uterine bleeding occurred. Increased cost is another drawback, however utilizing a hysteroscope will decrease the costs from multiple failed IVF-ET attempts and improve patient satisfaction.

5. Conclusion

Hysteroscopic endometrial embryo delivery (HEED) is a beneficial technique in increasing clinical pregnancy rates, especially in patients with repeated failed IVF-ET attempts. Due to the objective and replicable nature of the hysteroscopic procedure along with increased accuracy of placement of embryo(s), efforts in reducing multiple pregnancies should now be more focused on increasing our knowledge of selecting embryo(s) with high survival potential for embryo transfer. Ectopic pregnancies from IVF will be minimized by using lower transfer volumes of 5 μ l and visually confirmed positional placement of embryos away from the uterine cornu. Ectopics are almost eliminated when using the SEED technique for blastocyst embryo transfer.

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