

# We are IntechOpen, the world's leading publisher of Open Access books Built by scientists, for scientists

6,900

Open access books available

185,000

International authors and editors

200M

Downloads

Our authors are among the

154

Countries delivered to

TOP 1%

most cited scientists

12.2%

Contributors from top 500 universities



WEB OF SCIENCE™

Selection of our books indexed in the Book Citation Index  
in Web of Science™ Core Collection (BKCI)

Interested in publishing with us?  
Contact [book.department@intechopen.com](mailto:book.department@intechopen.com)

Numbers displayed above are based on latest data collected.  
For more information visit [www.intechopen.com](http://www.intechopen.com)



# Duplication of Coagulation Factor Genes and Evolution of Snake Venom Prothrombin Activators

Shiyang Kwong<sup>1</sup> and R. Manjunatha Kini<sup>1,2</sup>

<sup>1</sup>*Department of Biological Sciences, Faculty of Science  
National University of Singapore, Singapore*

<sup>2</sup>*Department of Biochemistry, Medical College of Virginia  
Virginia Commonwealth University, Richmond, Virginia*

<sup>1</sup>*Singapore*

<sup>2</sup>*USA*

## 1. Introduction

Snake venom is a complex mixture of pharmacologically active molecules which are responsible for immobilization, paralysis, death and digestion of prey organisms. This armory of toxins has evolved to target two key systems, namely the neuromuscular and circulatory systems, in order to induce rapid immobilization and death. So far, several hundreds of protein toxins from snake venoms have been purified and characterized. Most of these toxins have been documented to be structurally, and at times functionally, similar to proteins expressed in different tissues of the body. For example, elapid phospholipase A<sub>2</sub> toxins are structurally and catalytically similar to mammalian pancreatic phospholipase A<sub>2</sub> enzymes (Robin Doley et al. 2009). Similarly, sarafotoxins are structurally and functionally similar to endothelins produced primarily in endothelium (Landan et al. 1991a). Based on such structural and functional similarities, it is hypothesized that toxin proteins are “recruited” from body proteins by gene duplication (Fry 2005). Accordingly, the genes of body proteins are duplicated and modified to have differential and specific expression in venom glands. This phenomenon is broadly termed as “recruitment”. This “recruitment” process of body proteins has not only been observed in snakes but also in various other venomous animals, such as cone snails, spiders, scorpions and sea anemones as well as hematophagous animals (Fry et al. 2009). Although this overarching concept existed in the field of snake venom toxins for decades, there is not much direct molecular evidence for this process of “recruitment”.

Our laboratory has extensively characterized prothrombin activators from Australian elapid snake venoms and documented their structural and functional similarity with mammalian plasma coagulation factors. Through systematic, detailed studies, we provided the molecular details of the “recruitment” of venom prothrombin activators from plasma coagulation factors after gene duplication. We also identified several key structural changes that make these prothrombin activators better toxins. In this chapter, we will describe the first molecular evidence for the “recruitment” process and the evolution of prothrombin activators in venoms of Australian elapid snakes.

2. Snake venom prothrombin activators

As mentioned above, the circulatory system is one of the main targets of snake venom toxins. These toxins affect heart function (e.g., cardiotoxins), vasculature and blood pressure (e.g., sarafotoxins, natriuretic peptides and hemorrhagic toxins), blood coagulation (e.g., procoagulant and anticoagulant proteins), platelet aggregation (e.g., agonists and antagonists) and fibrinolysis (e.g., direct and indirect fibrinolytic proteinases) (Braud et al. 2000; Chow and Kini 2001; Hutton and Warrell 1993; Kini 2004; Kini and Chow 2001; Kini and Evans 1990; Markland 1997; Markland 1998; Morita 2004). All procoagulant proteins are generally proteinases that promote blood coagulation by activating zymogens of specific plasma coagulation factors (Davie 2003). Prothrombin activators are a group of procoagulant proteins that specifically activate prothrombin to thrombin, which then induces blood coagulation through fibrin clot formation.

Several prothrombin activators have been identified and characterized from snake venoms (Gao et al. 2002; Hasson et al. 2003; Joseph and Kini 2001; Kornalik and Blomback 1975; Morita and Iwanaga 1978; Rosing and Tans 1991; Rosing and Tans 1992; Schieck et al. 1972; Silva et al. 2003; Speijer et al. 1986; St Pierre et al. 2005; Yamada and Morita 1997), and based on their properties (structure, cofactor requirements and end-products formed), these snake venom prothrombin activators have been classified into four groups (Kini et al. 2001) (Table 1). Group A and B prothrombin activators, such as ecarin from *Echis carinatus* venom (Kornalik and Blomback 1975; Morita and Iwanaga 1978; Nishida et al. 1995; Schieck et al. 1972) and multactivase from *E. multisquamatus* venom (Yamada and Morita 1997), are metalloproteinases which induce coagulation by converting prothrombin to meizothrombin. These proteins are structurally distinct from blood coagulation factors. Group C and D prothrombin activators, such as oscutarin from *Oxyranus scutellatus* venom (Owen and Jackson 1973; Speijer et al. 1986; Walker et al. 1980; Welton and Burnell 2005) and notecarin from *Notechis scutatus scutatus* venom (Tans et al. 1985), are serine proteinases that induce coagulation by converting prothrombin to thrombin. These proteins exhibit functional similarity to mammalian plasma coagulation factors. However, no detailed structural information of these proteins was available. To fill this void, we initiated structural studies of these prothrombin activators. We characterized one representative each of group C (pseutarin C) and group D (trocarin D) prothrombin activators. Our results show that snake venom prothrombin activators are structurally and functionally similar to mammalian plasma coagulation factors.

Group	Cofactor Requirements	Type of proteinase	Product formed	Estimated Size	Similar to Plasma Coagulation Factors	Examples
A	None	Metalloproteinase	Meizothrombin	~47 kDa	None	Ecarin
B	Ca <sup>2+</sup>			Two subunits of ~25 and ~60 kDa		Carinactivase, multactivase
C	Ca <sup>2+</sup> plus phospholipids	Serine proteinase	Thrombin	Two subunits of ~60 and ~220 kDa	"FXa-FVa" complex	Pseutarin C oscutarin,
D	Ca <sup>2+</sup> plus phospholipids plus factor Va			~60 kDa	FXa	Trocarin D, hopsarin D, notanarin D

Table 1. Classification of snake venom prothrombin activators

## 2.1 Group D prothrombin activators

Group D prothrombin activators are found exclusively in the venom of Australian elapid snakes (Rosing and Tans 1991). Notecarin from *Notechis scutatus scutatus* venom was the first member of this group to be isolated and characterized (Tans et al. 1985). Since then, similar prothrombin activators have been characterized from several other snake venoms. They are glycoproteins with a molecular weight of ~50 kDa (Table 1). As a group of proteins, they share striking resemblances and requirements for optimal activity with activated mammalian plasma coagulation factor X (FXa) (Table 1) (Joseph et al. 1999; Marsh et al. 1997; Rao and Kini 2002; Stocker et al. 1994; Tans et al. 1985).

Venom of the Australian elapid *Tropidechis carinatus* (rough-scaled snake) was documented to have procoagulant properties 29 years ago (Chester and Crawford 1982). A prothrombin activator was isolated using gel filtration and benzamidine-based affinity chromatography and was partially characterized (Marsh et al. 1997). Our laboratory purified a prothrombin activator, trocarin D, from *T. carinatus* venom to homogeneity using a series of high performance liquid chromatography techniques including gel filtration, ion-exchange and reverse-phase chromatographies (Joseph et al. 1999). This purification procedure was refined to a single-step reverse-phase chromatographic method and was subsequently used for the purification of several other group D prothrombin activators such as notanarin D from *N. ater niger* venom, notecarin D from *N. scutatus* venom and hopsarin D from *Hoplocephalus stephensi* venom (Rao et al. 2003a). Our laboratory characterized trocarin D for its functional and structural properties in detail as a representative of group D prothrombin activators.

Functionally, trocarin D has properties which are similar to mammalian plasma coagulation FXa. They both promote blood coagulation by activating prothrombin to thrombin (Joseph et al. 1999). Trocarin D and FXa achieve activation of prothrombin by cleaving the same peptide bonds (Arg<sub>274</sub>-Thr<sub>275</sub> and Arg<sub>323</sub>-Ile<sub>324</sub>). Both proteins have identical co-factor requirements of Ca<sup>2+</sup> ions, phospholipids and activated factor V (FVa) for their optimal activities (Joseph et al. 1999). We determined the amino acid sequence of trocarin D and its precursor using Edman degradation (Joseph et al. 1999) and cDNA sequencing (Reza et al. 2005a), respectively. Trocarin D and mammalian FXa share significant sequence identity (~53-60%) and exhibit identical domain architecture (Joseph et al. 1999; Rao et al. 2003a) (Figure 1). Both proteins comprise two chains: a heavy chain, which has a serine proteinase with the characteristic catalytic triad (His<sub>42</sub>, Asp<sub>88</sub> and Ser<sub>185</sub>), and a light chain, which has a Gla domain followed by two epidermal-growth factor-like domains (EGF-I and EGF-II). These two chains are held together by a single inter-chain disulfide bond (Joseph et al. 1999) (Figure 1). The differences between trocarin D and mammalian FXa reside in an insertion in the heavy chain, the size of the activation peptide and post-translational modifications. Firstly, there is a 12-residue insert in the heavy chain of trocarin D (Reza et al. 2005a). However, the functional importance of this insertion is not clear. Secondly, the activation peptide of trocarin D precursor is only 27 residues long (Reza et al. 2005a) compared to the activation peptides of mammalian FXs which ranges from 48 to 52 residues (Figure 1). Lastly, post-translational modifications show that trocarin D is glycosylated, but mammalian FXa is not. In addition, trocarin D also contains a O-linked carbohydrate at Ser<sub>52</sub> of the light chain and a N-linked carbohydrate at Asn<sub>45</sub> of the heavy chain (Joseph et al. 1999) (Figure 1). Interestingly, the O-linked carbohydrate moiety has a N-acetylglucosamine moiety, which is found commonly in nuclear and cytoplasmic proteins but rarely in secreted proteins (Hanover et al. 1987; Holt et al. 1987; Holt and Hart 1986; Snow et al. 1987). The

function of glycosylation is hypothesized to protect trocarin D from inactivation by proteolysis and confer it with thermal stability (Rao et al. 2003a; Wang et al. 1996). Although human FX contains two *O*-glycosylation and two *N*-glycosylation moieties (Inoue and Morita 1993), all the carbohydrate moieties are exclusively found on the activation peptide, which is removed when FX is activated. Therefore, activated mammalian FXa is not glycosylated. In contrast, the short activation peptide of trocarin D does not have any glycosylation sites. The last difference in post-translational modification is that the Asp<sub>63</sub> of FX light chain EGF-I domain is  $\beta$ -hydroxylated (McMullen et al. 1983; Stenflo et al. 1987), but the corresponding residue in trocarin D is not (Joseph et al. 1999) (Figure 1).

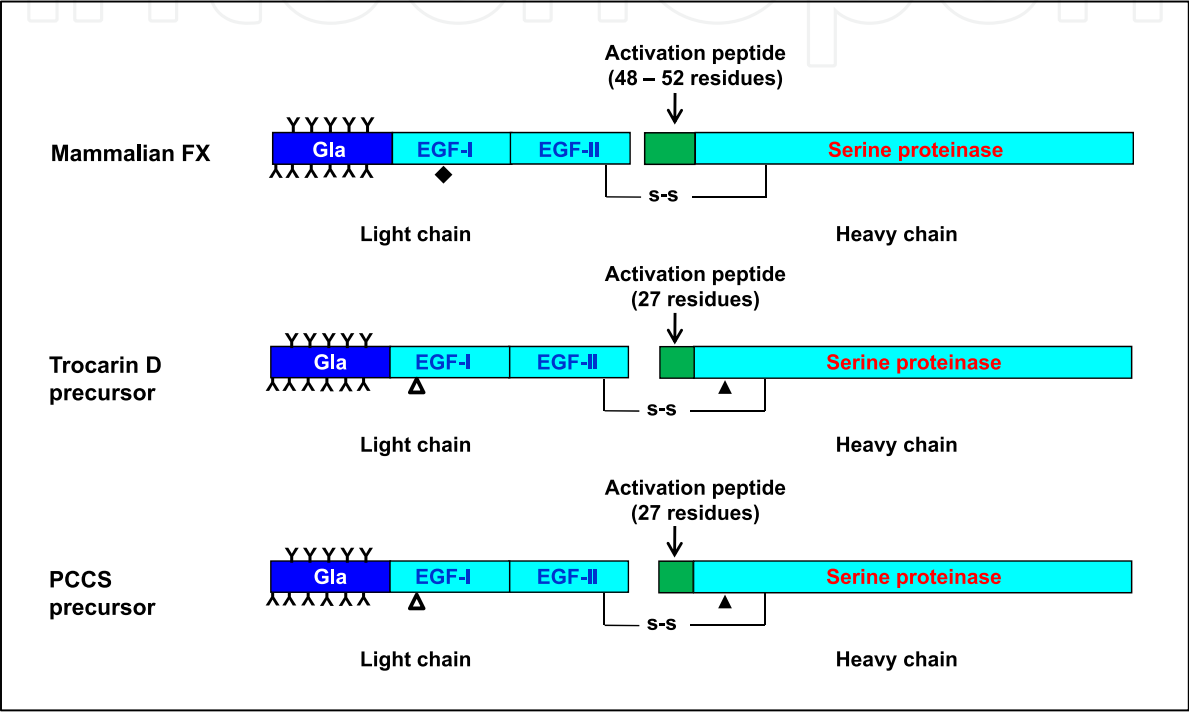


Fig. 1. Domain architecture of mammalian FX, and precursors of trocarin D and PCCS. The Ys represent Gla residues; open triangles represent the *O*-linked carbohydrate at Ser<sub>52</sub> on trocarin D and PCCS; solid triangles represent the *N*-linked carbohydrate at Asn<sub>45</sub> on trocarin D and PCCS; the solid diamond represents  $\beta$ -hydroxylation at Asp<sub>62</sub> on mammalian FXa.

### 2.2 Group C prothrombin activators

The group C snake venom prothrombin activators are also found exclusively in the venoms of Australian elapid snakes (Rosing and Tans 1991). Oscutarin from *Oxyuranus scutellatus* venom was the first member of the group C prothrombin activators to be isolated and characterized (Owen and Jackson 1973; Speijer et al. 1986; Walker et al. 1980; Welton and Burnell 2005). This group of proteins is generally ~300 kDa in size and comprises two subunits (~60 and ~220 kDa) (Table 1). The smaller enzymatic subunit is a serine proteinase and has characteristics of FXa, whereas the nonenzymatic subunit resembles activated mammalian plasma coagulation factor V (FVa). Overall, group C prothrombin activators have striking resemblances to and similar co-factor requirements as the mammalian plasma coagulant “FXa-FVa” complex (Filippovich et al. 2005; Masci et al. 1988; Rao and Kini 2002; Speijer et al. 1986; Walker et al. 1980) (Table 1).



Pseutarin C was purified from *P. textilis* venom, and it activates prothrombin to thrombin. For its optimal activity, pseutarin C requires only  $\text{Ca}^{2+}$  ions and phospholipids (Rao and Kini 2002). These functional characteristics are similar to that of the mammalian “FXa-FVa” complex. As with other group C prothrombin activators, pseutarin C comprises two subunits of ~60 kDa and ~220 kDa (Rao and Kini 2002) (Table 1). The smaller subunit, with serine proteinase activity, was termed the pseutarin C catalytic subunit (PCCS) and the larger subunit, which has no enzymatic activity, was termed the pseutarin C nonenzymatic subunit (PCNS) (Rao and Kini 2002). A comparison of the protein quantities in the venom and plasma revealed that pseutarin C is expressed ~4,200 times higher in the venom than the amount of FV and FX in the plasma (Rao and Kini 2002). We purified pseutarin C and its subunits and characterized them both functionally and structurally as representatives of the group C prothrombin activators.

1. *Pseutarin C catalytic subunit (PCCS)* – Functionally, PCCS is similar to mammalian FXa and group D prothrombin activators (Rao and Kini 2002). They have the same co-factor requirements, including  $\text{Ca}^{2+}$  ions, phospholipids and FVa, for their optimal activity and activate prothrombin by cleaving the same two peptide bonds ( $\text{Arg}_{274}\text{-Thr}_{275}$  and  $\text{Arg}_{323}\text{-Ile}_{324}$ ). The enzymatic activity ( $V_{\text{max}}$ ) of PCCS is enhanced by the presence of FVa (Rao and Kini 2002). The amino acid sequence of PCCS and its precursor was determined using both Edman degradation and cDNA sequencing (Rao et al. 2004; Rao and Kini 2002). Structurally, PCCS is also similar to mammalian FXa and group D prothrombin activators (Figure 1). Its sequence shows ~42% identity to mammalian FXa and 74-83% identity to group D prothrombin activators (Rao et al. 2004). Like mammalian FXa and group D prothrombin activators (Rao et al. 2004), the domain architecture of PCCS consists of a light and a heavy chain that are linked by a single disulfide bond (Rao et al. 2004). The light chain has a Gla domain followed by two EGF-like domains, and the heavy chain contains a serine proteinase domain (Figure 1). Despite such functional and structural similarities, the differences between PCCS and mammalian FXa reside in the size of the activation peptide and post-translational modifications (Rao et al. 2004). Similar to trocarin D precursor, the activation peptide of PCCS precursor is 27 residues long and is significantly shorter than those of mammalian FXs (Figure 1). Like trocarin D, it has an insertion in its heavy chain. Interestingly, the PCCS insert is 13 residues long and is distinctly different from the 12-residue insert in trocarin D (Rao et al. 2004; Reza et al. 2005b). This strongly indicates that the evolution of groups C and D prothrombin activators are independent. Like trocarin D, the  $\text{Ser}_{52}$  and  $\text{Asn}_{45}$  residues of the light and heavy chains of PCCS are O- and N-glycosylated, respectively (Figure 1). However, as mentioned previously, these two residues have no post-translational modifications in mammalian FX/FXa. These functional and structural characteristics suggest that PCCS is a homologue of mammalian FXa and group D prothrombin activators.

2. *Pseutarin C nonenzymatic subunit (PCNS)* – Structurally, PCNS is similar to mammalian FV. The amino acid sequence of PCNS and its precursor was determined using Edman degradation and cDNA sequencing (Rao et al. 2003b; Rao and Kini 2002). PCNS shares ~50% identity with the mammalian FV and has identical domain architecture with mammalian FV (Rao et al. 2003b). Both PCNS and mammalian FV have six domains: A1, A2, B, A3, C1 and C2 (Figure 2). Domains A and C are functionally important, and these domains are highly conserved in PCNS and FVs of other species (Rao et al. 2003b). These structural similarities suggest that PCNS is a homologue of mammalian FV (Rao et al. 2003b).

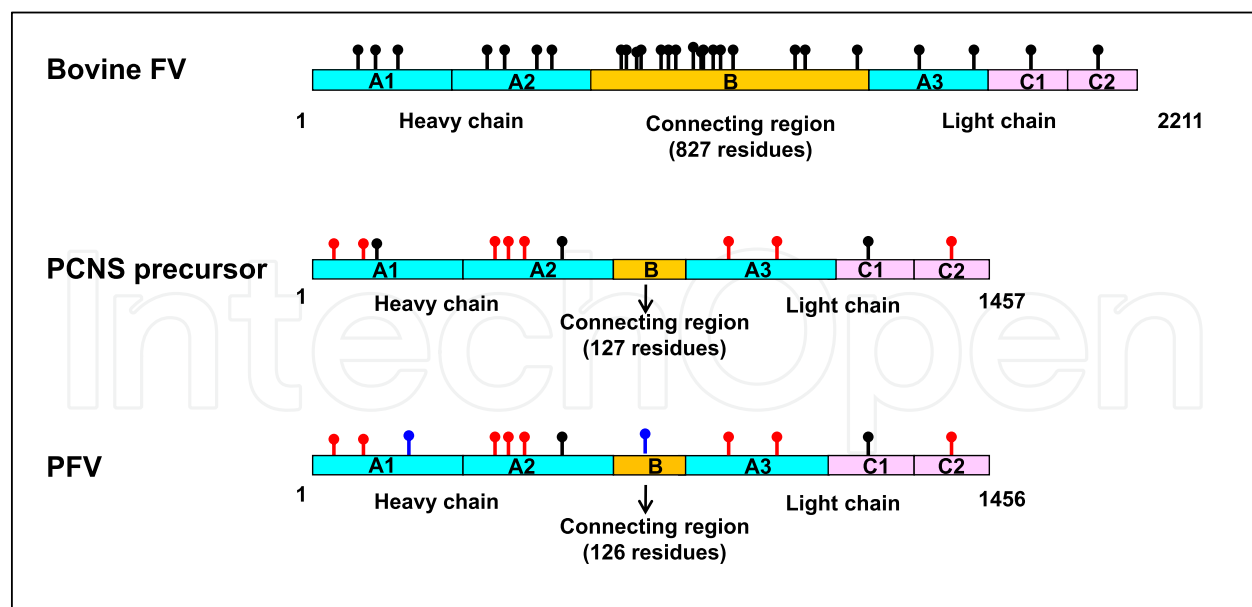


Fig. 2. Domain architecture of bovine FV, PCNS precursor and PFV. N-glycosylation sites are shown as colored knobs. Black knobs are conserved, while red knobs are found only in PCNS and PFV, and blue knobs are found only in PFV.

Despite being a FV homologue, PCNS shows several differences with FV from other species. Firstly, the domain B size of PCNS is significantly smaller (127 residues) than that of fishes (fugu: 530 residues and zebrafish: 756 residues) and mammals (murine: 843 residues, bovine: 869 residues, and human: 882 residues) (Rao et al. 2003b) (Figure 2). During FV activation, domain B is removed by thrombin or FXa by cleavage at three activation sites: Arg<sub>709</sub>, Arg<sub>1018</sub> and Arg<sub>1545</sub> (bovine FV numbering) (Foster et al. 1983; Nesheim et al. 1979; Suzuki et al. 1982) (Figure 3B). Although only two of these sites (Arg<sub>709</sub> and Arg<sub>1545</sub>) are conserved in PCNS (Figure 3B), the complete domain B can still be cleaved off during the activation of PCNS (Rao et al. 2003b). Thus, the difference in domain B size should not have any effect on the function of PCNS. Secondly, PCNS and FV of other species have different post-translation modifications. While bovine FV has 29 glycosylation sites, PCNS has only 11 potential N-glycosylation sites (Rao et al. 2003b) (Figure 2). Mammalian FV is phosphorylated at the Ser<sub>692</sub> residue, but PCNS is not (Rao et al. 2003b). In addition, there are six sulfation sites in human FV that are absent in PCNS (Rao et al. 2003b). This difference in post-translation modifications is interesting, as sulfation and phosphorylation are important for regulating the activation of human FV by thrombin (Kalafatis et al. 1994; Pittman et al. 1994).

Aside from these differences, it is noted that PCNS possesses certain modifications and associations that allow it to function efficiently as a toxin (Bos et al. 2009). Firstly, PCNS has evolved a way to evade inactivation by protein C. In the human coagulation system, protein C is activated by thrombin in the presence of thrombomodulin, and activated protein C (APC) subsequently inactivates FVa in a negative feedback loop (Esmon 2001). This inactivation occurs by proteolytic cleavage at three sites on the FVa heavy chain: Arg<sub>306</sub>, Arg<sub>506</sub> and Arg<sub>662</sub> (Kalafatis et al. 1994; Mann et al. 1997) in bovine FVa (Figure 3B). PCNS is able to evade APC inactivation, as it does not have any of the three APC cleavage sites (Rao et al. 2003b). Even an alternate less efficient cleavage site at Arg<sub>316</sub> (van der Neut et al. 2004b)

is not conserved in PCNS (Rao et al. 2003b). Secondly, FVa is inactivated by phosphorylation at Ser<sub>692</sub> (Kalafatis et al. 1994). This phosphorylation site is not present in PCNS (Rao et al. 2003b). Lastly, PCNS is shielded from APC inactivation (Nesheim et al. 1982; Rao et al. 2003b) and is kept activated through its constant and stable association with FXa-like PCCS (Rao et al. 2003a; Rao et al. 2004; Thorelli et al. 1998) in the venom. We have shown experimentally that pseutarin C is unaffected by APC, while bovine FXa-FVa complex is completely inactivated (Bos et al. 2009; Rao et al. 2003b) (Figure 4). Overall, PCNS is a good example of how a toxin gene is duplicated from an ancestral gene and undergoes modifications to gain unique characteristics that allow it to function efficiently as a toxin.

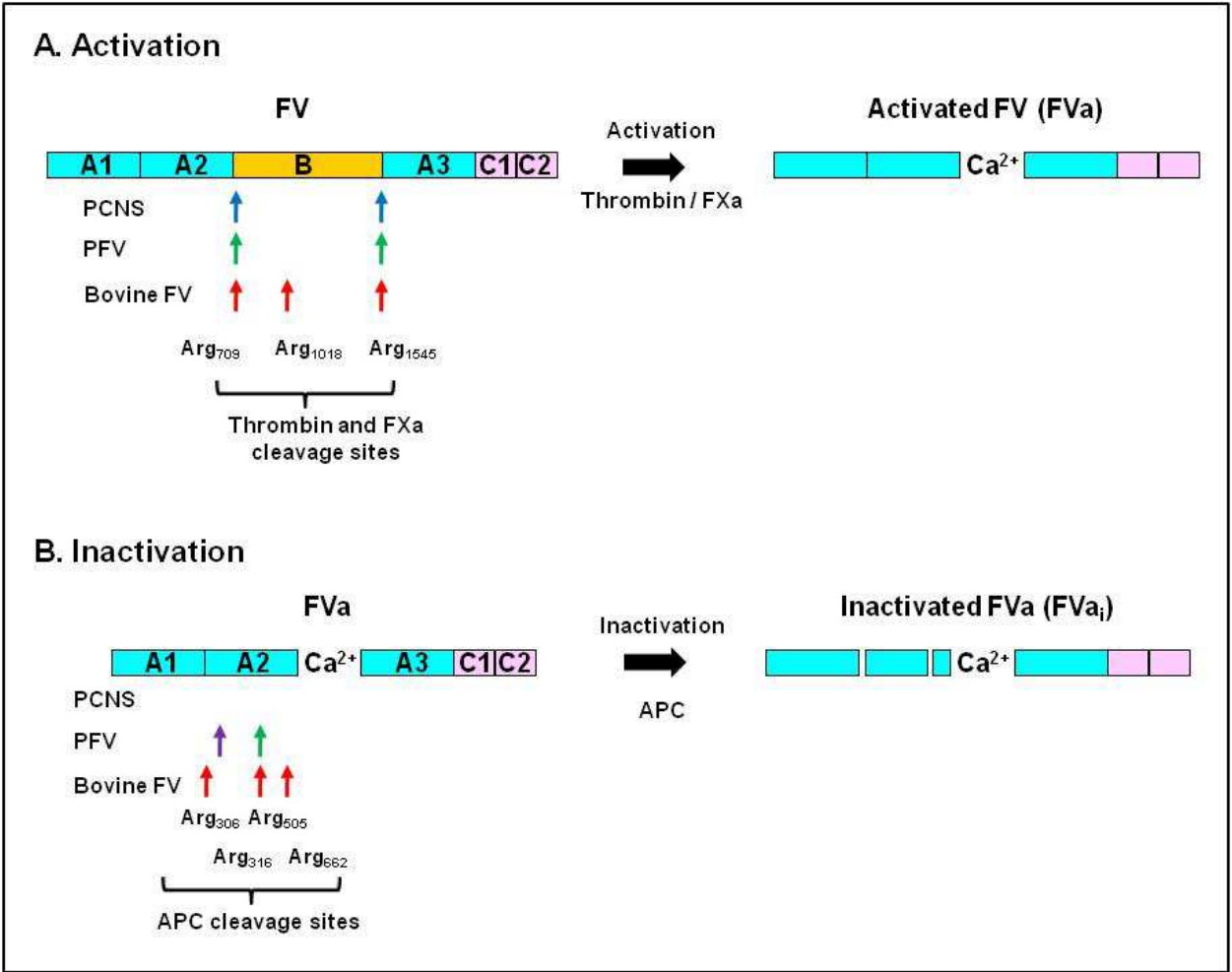


Fig. 3. Functionally important proteolytic sites in PCNS and FV. (A) Activation sites. Thrombin and FXa cleavage sites of PCNS, PFV and bovine FV are shown in blue, green and red, respectively. Critical activation sites are conserved in PCNS and PFV. (B) Inactivation sites. Activated protein C (APC) cleavage sites of PFV and bovine FV are shown in green and red, respectively. In addition to Arg<sub>505</sub>, PFV has a primitive inactive site at Arg<sub>316</sub> shown in purple arrow. Inactivation sites are missing in PCNS.



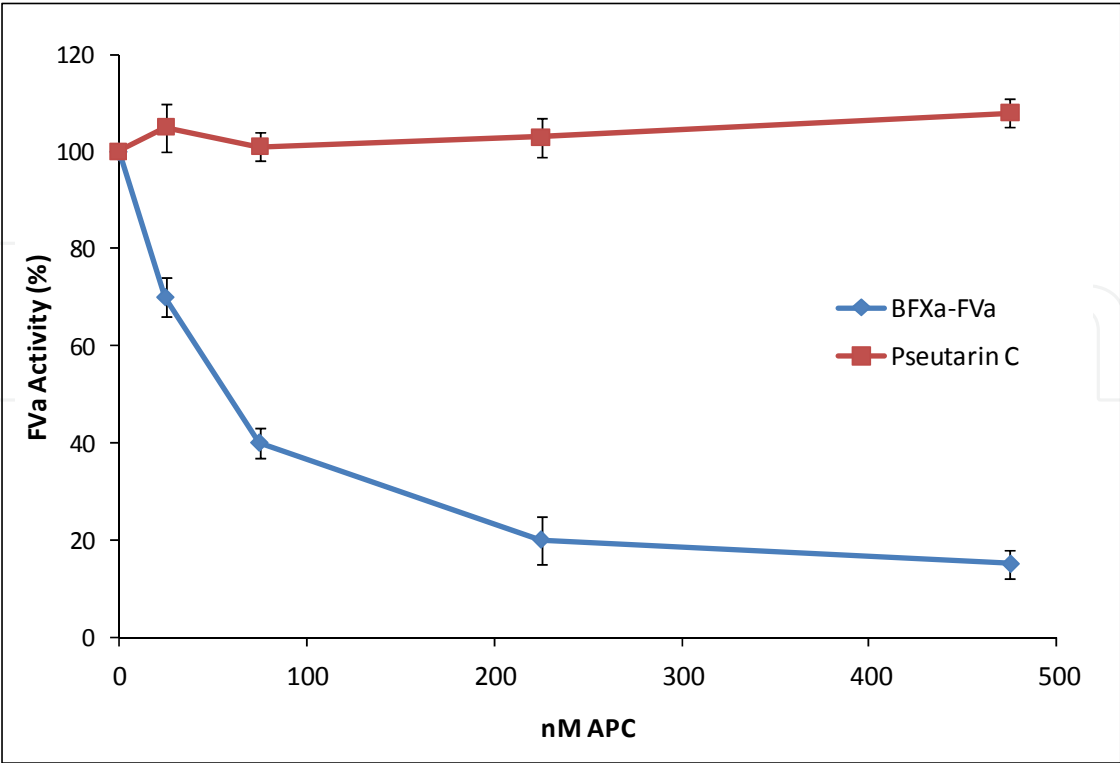


Fig. 4. APC resistance assay (Rao et al. 2003b). Varying concentrations of APC were added either to pseutarin C (E; 8 nM) or bovine FXa-FVa (F; FXa 42 nM, FVa 2 nM) complex which was diluted in 50 mM Tris-HCl buffer (pH 7.5) containing 100 mM NaCl, 5 mM CaCl<sub>2</sub>, and 0.5 mg/mL BSA. The reaction mixture was incubated for 30 minutes at room temperature. Prothrombin was added to a final concentration of 2.8 μM and thrombin formed was assayed using thrombin-specific chromogenic substrate S-2238. Each point represents an average of 2 independent experiments each carried out in triplicates.

3. Parallel prothrombin activator system in Australian elapid snakes

As described above, groups C and D snake venom prothrombin activators are functional and structural homologues of mammalian blood coagulation factors. As snakes are vertebrates, their hemostatic system should contain plasma coagulation factors. Thus, Australian elapid snakes should possess parallel prothrombin activating systems: one in their venom, which is used as an offensive weapon to attack the hemostatic system of the prey, and the other in their plasma, which is used for their own hemostatic purpose. We examined the presence of plasma coagulation factors in the snake’s hemostatic system and determined the relationship between the snake venom and plasma coagulation factors.

3.1 Trocarin D and FX from *Tropidechis carinatus* (TrFX)

Since the liver mainly expresses plasma coagulation factors, the cDNA encoding *T. carinatus* FX (TrFX) was sequenced from liver tissue (Reza et al. 2005a). The deduced amino acid sequence of TrFX is similar to mammalian FX (~50%) and trocarin D (~80%) (Figure 5). Structurally, TrFX is similar to trocarin D. They both have conserved cysteine residues and identical domain architecture. However, there are some differences between TrFX and trocarin D. The activation peptide of TrFX is similar to the mammalian FXs and not to that of

the venom prothrombin activators (Reza et al. 2005b). It is 57 residues long compared to 27 residues in trocarin D (Figure 5). In addition, there is no 12-residue insert in the heavy chain of TrFX as was observed to be present in the trocarin D precursor (Reza et al. 2005b). These differences in amino acid sequences, and the lengths of activation peptides and insertion in the heavy chain, suggest that TrFX and trocarin D are encoded by two independent genes. Hence, this confirms the presence of a parallel prothrombin activator system. TrFX is more similar to trocarin than to mammalian FX in terms of post-translational modifications (Reza et al. 2005a). TrFX and trocarin D both have N- and O-glycosylation modifications that are not found in mammalian FXs (as described previously).



Fig. 5. Alignment of deduced amino acid sequences of FX-like proteins from *T. carinatus* (TrFX and trocarin D) and *P. textilis* (PCCS, PFX1 and PFX2) snakes.

Trocarin D and TrFX differ in their physiological roles. Trocarin D plays an offensive role as a toxin in the venom that is used for killing prey. Upon envenomation, like other prothrombin activators (Masci et al. 1988; Rao et al. 2003a), it induces cyanation and death in experimental animals (Joseph et al. 1999) through disseminated intravascular coagulopathy. On the other hand, TrFX plays a crucial role in the coagulation cascade and prevents excessive blood loss by promoting blood coagulation when there is a vascular injury. Trocarin D is an active enzyme and is found in large quantities in the venom. In contrast, TrFX is found as a zymogen, which gets activated only when required and is found in much smaller concentrations in the plasma. Real-time polymerase chain reaction (RT-PCR) was used to determine the amount of expression of these two closely related proteins in the liver and venom gland. The results indicate that trocarin D is expressed in the venom gland but

not in the liver, while TrFX is expressed in the liver but not in the venom gland. Further, the expression of trocarin D is ~30 times higher in the venom gland than TrFX in the liver (Reza et al. 2007). Such differential expression patterns of trocarin D and TrFX strongly support the distinct physiological roles of these two proteins.

3.2 PCCS and FX from *Pseudonaja textilis* (PFX)

To understand the evolution of group C prothrombin activators, we also determined the cDNA sequence of the *P. textilis* FX (PFX) from the liver. Interestingly, two PFX isoforms (PFX1 and PFX2) were detected in the liver, and their cDNA sequences are ~85% similar (Reza et al. 2006). The domain architecture and cysteine residues of these two isoforms are also conserved compared to group D prothrombin activators. Amino acid sequence comparison shows that PFX1 is more similar to TrFX (~94%), while PFX2 is more similar PCCS and trocarin D (~90%) (Figure 5). Further, PFX1 has a longer activation peptide, similar to plasma FXs, whereas PFX2 has a shorter activation peptide, similar to PCCS and trocarin D. Also, PFX2 has a 9-residue insert, which is not present in PFX1. These structural differences suggest that PFX1, PFX2 and PCCS are encoded by three independent genes and that PFX2 is an evolutionary intermediate between PFX1 and PCCS (Reza et al. 2006) (Figure 5). This similarly confirms the presence of a parallel prothrombin activator system. The expression profiles of PFX1, PFX2 and PCCS were determined in liver and venom gland tissues by RT-PCR (Reza et al. 2006). The results show that PFX1 and PFX2 are expressed only in the liver, while PCCS is expressed only in the venom gland. PFX1 is also found to be expressed ~55,000 times higher than PFX2 in the liver, and PCCS is expressed ~80 times higher in the venom gland than is PFX1 in the liver (Reza et al. 2006). In summary, the sequence comparisons and expression profiles indicate that PCCS has evolved from PFX1 by gene duplication and PFX2 is an intermediary product of this “recruitment” process (Figure 6).

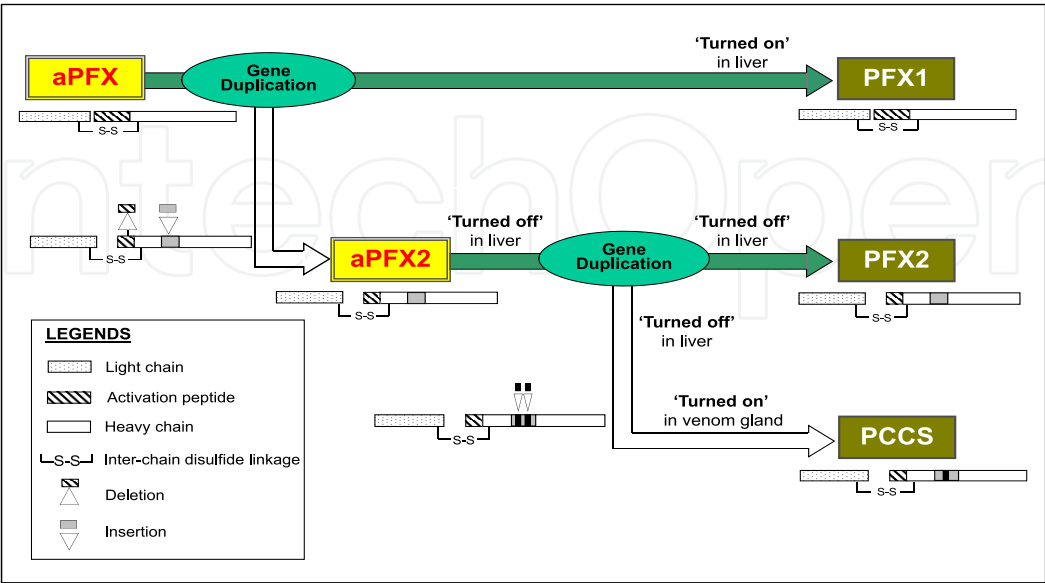


Fig. 6. Schematic diagram showing the probable evolutionary path in the recruitment of FX protein as toxin in the venom (Reza et al. 2006).

### 3.3 PCNS and FV from *Pseudonaja textilis* (PFV)

The cDNA sequence of *P. textilis* FV (PFV) was determined from its liver (Minh et al. 2005). The deduced amino acid sequence of PFV shows similarities to other mammalian and non-mammalian FVs (~50%) and PCNS (~96%) and shares identical domain architecture (Minh et al. 2005). Like the FVs of other species, PFV and PCNS comprise A1, A2, B, A3, C1 and C2 domains (Figure 3A). Functionally important domains A and C are highly conserved in both PFV and PCNS, whereas domain B is the most variable (Minh et al. 2005). The domain B (126 residues) of PFV is one residue shorter than that of PCNS (127 residues), and is much shorter than that of mammalian and non-mammalian FVs. A more detailed comparison shows that all the FXa and thrombin proteolytic cleavage sites (which are important for activation of these nonenzymatic proteins) are conserved in PFV and PCNS (Figure 3A). However, PFV has an additional FXa proteolytic cleavage site at Arg<sub>1765</sub> (Minh et al. 2005; Rao et al. 2003b). This cleavage site also exists in mammalian FV but not in FVs of teleosts (Minh et al. 2005). This is evolutionarily interesting as this additional cleavage site may be a characteristic found only in tetrapod FVs. However, the functional implication of this cleavage with regards to procoagulant activity of FV is not yet known.

As mentioned previously, PCNS has evolved to be resistant to inactivation by activated protein C (APC), which is crucial to its function as a toxin. On the other hand, PFV is similar to other FV, as it can still be inactivated by APC. PFV can be inactivated by APC by cleavage at Arg<sub>316</sub>, a primitive inactive site (van der Neut et al. 2004a), and at Arg<sub>506</sub> (Minh et al. 2005) (Figure 3B). The expression profiles of PFV and PCNS in the liver and the venom gland were determined using RT-PCR. As with other venom prothrombin activator genes, PCNS is expressed only in the venom gland, while PFV is expressed only in the liver. It was found that PCNS is expressed ~280 times higher in the venom gland than is PFV in the liver (Minh et al. 2005). Thus, PCNS and PFV have differential expressions (Minh et al. 2005).

Based on sequence comparisons, we confirmed the presence of parallel prothrombin activator systems in Australian elapid snakes and showed for the first time that groups C and D prothrombin activators in snake venom and their plasma coagulation factor counterparts are closely related. We also proposed that these venom prothrombin activators evolved from their plasma coagulation factor counterparts by gene duplication and were subsequently modified to function efficiently as toxins.

## 4. Phylogenetic relationship between snake venom and plasma prothrombin activators

A phylogenetic tree of the snake venom and plasma prothrombin activators with other known FX sequences was constructed to understand their evolutionary relationships using zebrafish FX as the out group (Reza et al. 2006; St Pierre et al. 2005). All the reptilian sequences form a monophyletic group (Reza et al. 2006) (Figure 7). Within the reptilian clade, group C and D prothrombin activators appear as two separate clades on the tree. This indicates that, despite their similarities, group C and D prothrombin activators have originated independently. Interestingly, the PFX2 sequence is found nested within the group C prothrombin activators. This supports the hypothesis that PFX2 is an evolutionary intermediate of PCCS from PFX1. Based on the topology of the phylogenetic tree, it is suggested that these snake venom prothrombin activators have been “recruited” through independent evolutionary events (Reza et al. 2006) (Figure 7).

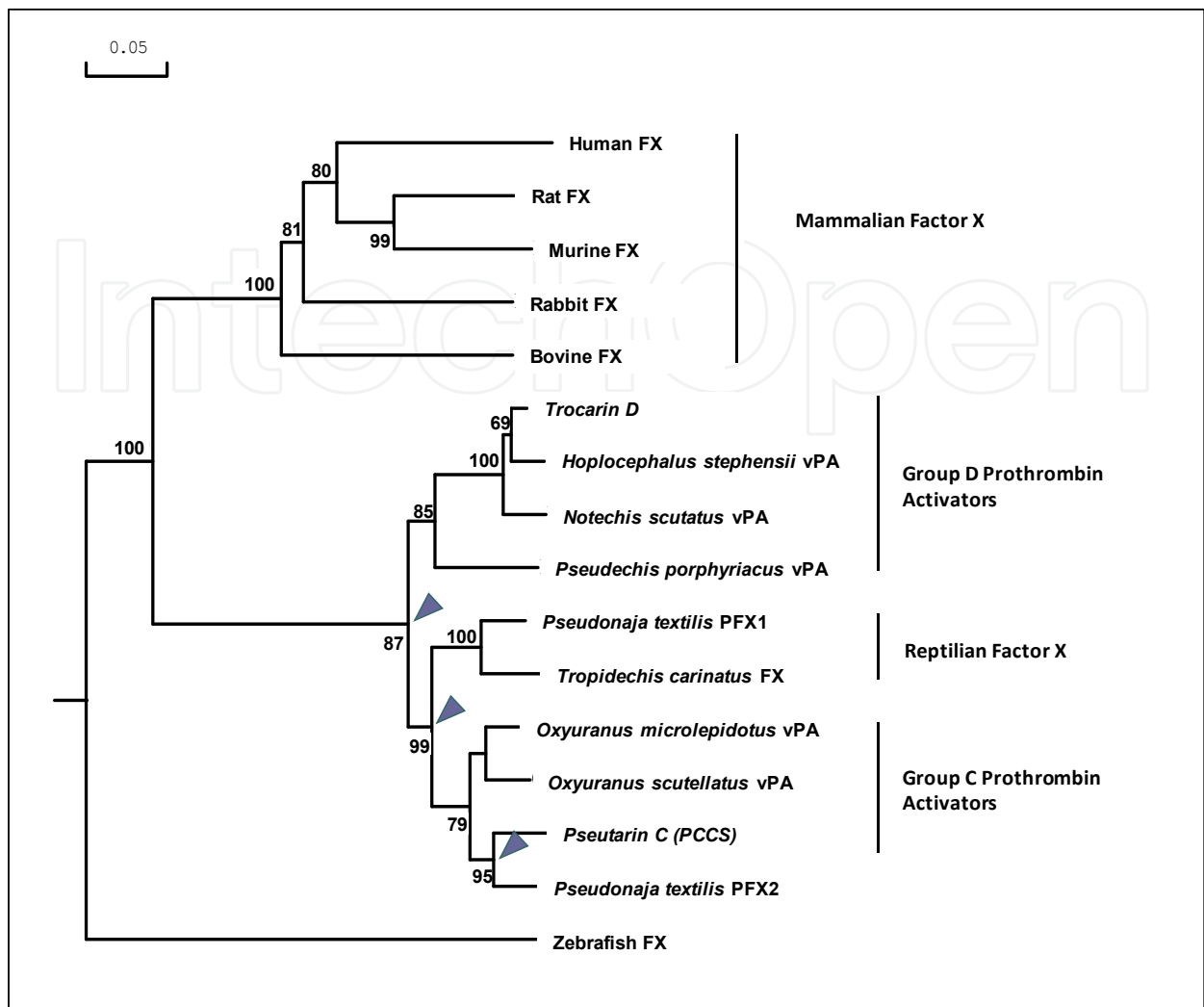


Fig. 7. Phylogenetic relationships of snake venom and plasma prothrombin activators with other known FX sequences (Reza et al. 2006). “vPA” is an abbreviation for venom prothrombin activators. Arrows indicate the three independent “recruitment” events of snake venom prothrombin activators.

### 5. Comparison of trocarin D and TrFX genes

In the previous sections, we have described how the venom prothrombin activators have been modified to gain certain characteristics, such as resistance to inactivation, which enables them to function better as toxins. However, differential and tissue-specific expression of venom prothrombin activators and their plasma coagulation factors is also important for their respective physiological roles. The expression of toxins should be venom gland-specific and inducible to higher levels. This is so the snake can protect itself against its own venom toxins and replenish its venom supply quickly. Conversely, plasma coagulant factors are mainly expressed in the liver at constitutently low levels so that they can be activated to induce blood coagulation during vascular injuries.

To understand how the venom prothrombin activators are regulated for tissue specificity and level of expression, we determined the gene structure of trocarin D and TrFX. Based on the cDNA sequences of trocarin D and TrFX, and that of mammalian FX gene, primers were



designed, and the gene sequences were determined using genomic DNA PCR and genome walking strategies (Reza et al. 2006). The gene organizations of trocarin D and TrFX are identical (eight exons and seven introns). The intron sequences were highly similar to each other with only differences in the promoter and intron 1 regions (Figure 8). Such similarities strongly support our findings that there are two closely related, parallel prothrombin activator systems, and that the venom prothrombin activators are “recruited” from plasma coagulation factors through gene duplication. The duplicated FX gene was subsequently modified and “recruited” for expression in the venom gland as a venom prothrombin activator.

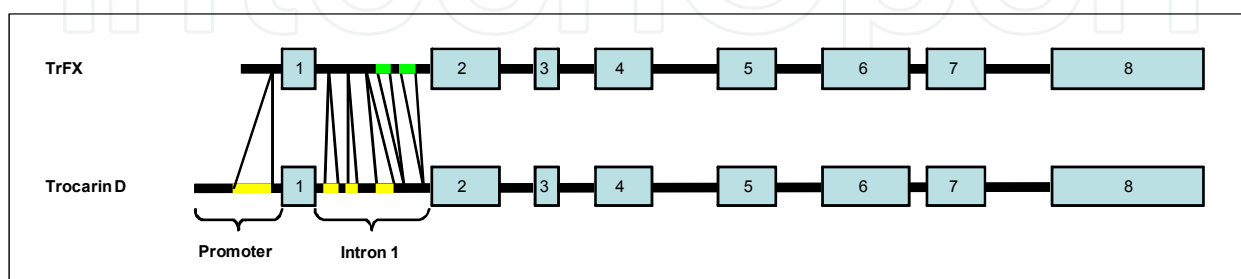


Fig. 8. Gene organization comparison of trocarin D and TrFX. Exons are shown as numbered boxes. The differences in the promoter and intron 1 regions are indicated in yellow (insertions in trocarin D gene) and green (deletions in TrFX gene) (Reza et al., 2007).

### 5.1 *Cis*-elements in trocarin D promoter region

The overlapping promoter regions of trocarin D and TrFX were characterized by comparing them against previously characterized human (Hung et al. 2001; Hung and High 1996) and murine (Wilberding and Castellino 2000) FX promoter regions (Reza et al. 2007) (Figure 9). Based on these comparisons, four conserved *cis*-regulatory elements in the trocarin D and TrFX promoter regions were identified (Figure 9): (i) a CCAAT box (Hung et al. 2001; Hung and High 1996; Wilberding and Castellino 2000), (ii) a gut-specific transcription factor GATA-4 binding site (Hung et al. 2001), (iii) a liver-specific transcription factor HNF-4 (Hung and High 1996), and (iv) multiple Sp1/Sp3 binding sites (Hung et al. 2001).

Comparison of the trocarin D and TrFX promoter regions reveals that trocarin D has a 264 bp insertion (Figure 8 and 9). This 264 bp is located from -33 to -297 bp upstream of the trocarin D start codon (ATG) (Figure 9). This insertion is postulated to play a major role in the recruitment of the duplicated TrFX gene by causing it to be exclusively expressed in the venom gland as the procoagulant toxin, trocarin D. Hence, it was termed Venom Recruitment/Switch Element (*VERSE*). This segment was characterized for its *cis*-elements and gene-regulatory role using luciferase assays in primary venom gland cells and mammalian cell lines (Kwong et al. 2009). The *VERSE* promoter was found to be responsible for the elevation of expression levels, but not tissue-specific expression, of trocarin D. In terms of *cis*-element characterization, besides confirming the presence of two TATA-boxes, one GATA box and one Y-box, three novel *cis*-elements were also identified (Figure 9). Functionally, it is found that both TATA boxes (TLB2 and TLB3) are functional. However, TLB2 is the primary TATA box which initiates and directs transcription start site (Kwong et al. 2009) (Figure 9).

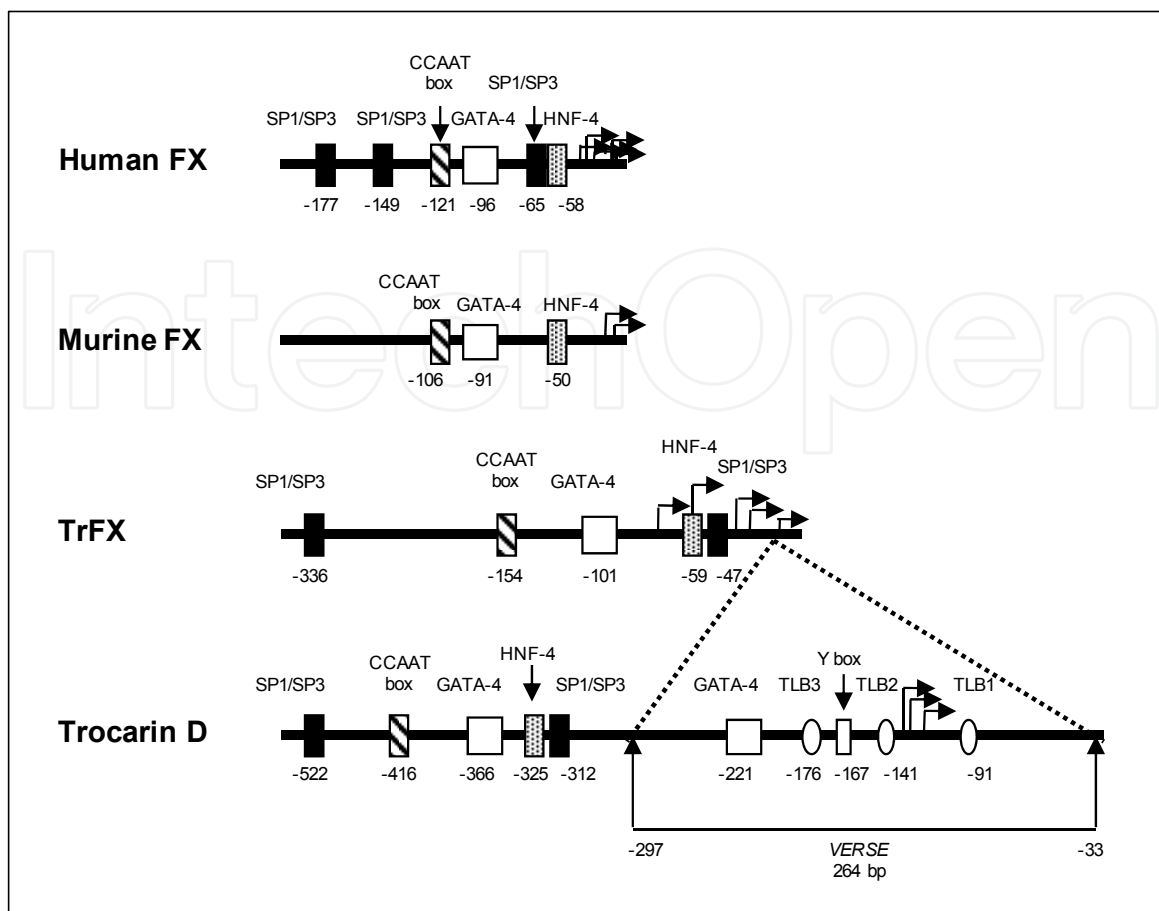


Fig. 9. Comparison of promoter regions in mammalian and *T. carinatus* prothrombin activator genes (Reza et al., 2007).

## 5.2 Comparison of trocarin D and TrFX first introns

The intron 1 size of trocarin D is 7911 bp, while that of TrFX is 5293 bp. The difference in size is explained by three insertions and two deletions in the trocarin D intron 1 region (Reza et al. 2007) (Figure 10). Bioinformatics analysis of these insertion/deletion segments reveals that they are novel. The three insertion segments within intron 1 of trocarin D region are 214 bp, 1975 bp and 2174 bp in size with respective positions at 128 bp, 914 bp and 3300 bp on the trocarin D gene (Figure 10). Upon closer analysis of the insertion segment sequences, it is observed that the first insert within intron 1 of trocarin D is almost an exact repeat (96.33% identity) of the intron 1 segment spanning from 3082 bp to 3299 bp. The other two inserts seem to be inverted repeats of each other (~71% identity). The third insert shows potential of being a Scaffold/Matrix Attachment Region (S/MAR) due to: (i) a high AT content (Cockerill and Garrard 1986; Liebich et al. 2002; Zhou and Liu 2001), (ii) a topoisomerase II (Boulikas 1993), (iii) a S/MAR consensus motif (van Drunen et al. 1997), (iv) a significant over-representation of characteristic hexanucleotides (Liebich et al. 2002), and (v) an ATTA motif and an AT-rich region with H-box (Will et al. 1998). The two deletion segments within intron 1 of trocarin D are 255 bp and 1406 bp in size with respective positions at 2610 bp and 3770 bp on the TrFX gene (Reza et al. 2007) (Figure 10). As the *VERSE* promoter of trocarin D does not regulate tissue-specific expression (Kwong et al. 2009), it is postulated that these insertions and deletions in the

intron 1 of trocarin D could contain *cis*-elements that are responsible for the venom gland-specific expression of trocarin D. In summary, the characterization of the *VERSE* promoter and intron 1 regions of trocarin D has increased our understanding regarding gene regulation of venom prothrombin activators.

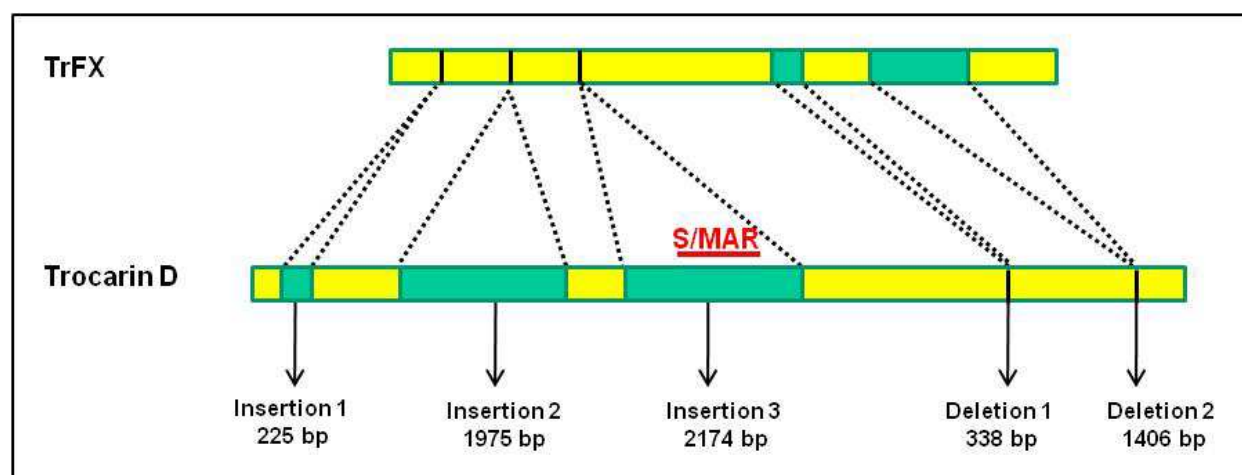


Fig. 10. Comparison of intron 1 regions in TrFX and trocarin D genes (Reza et al., 2007).

## 6. Gene duplication in snake venom toxin diversification

Besides “recruitment”, gene duplication also plays an important role in the diversification of venom toxins. This diversification is essential for the development of novel toxins. This diversification through gene duplication is evident from the many toxin isoforms present in the snake venom. Interestingly, each isoform varies in its function and gene regulation.

Gene duplication has also led to neofunctionalization of venom toxins, which has led to the new families of snake venom toxins (St Pierre et al. 2008) and addition of new members within these families (Fry et al. 2003; Landan et al. 1991b; Lynch 2007; Moura-da-Silva et al. 1996). The three-finger toxin (3FTx) multigene family is a good example of neofunctionalization by gene duplication (Fry et al. 2003). Structurally, all the members of this family have very well-conserved cysteine residues and share a common structure of three beta-stranded loops extending from a central core. However, they exhibit a wide variety of pharmacological effects. For example, acetylcholinesterase inhibition (fasciculin from *Dendroaspis angusticeps* venom), neurotoxicity ( $\alpha$ -bungarotoxin from *Bungarus multicinctus* venom), cardiotoxicity ( $\beta$  cardiotoxin from *Ophiophagus hannah* venom), and many others (for details, see (Kini and Doley 2010)). Neofunctionalization occurs when a toxin gene undergoes gene duplication and the duplicated gene is mutated within the functional sites, which often results in new ligand-binding specificities (Kini 2002).

Besides neofunctionalization, changes in gene regulation are also the outcomes of gene duplication. This can be seen in two isoforms present in the venom of *Naja sputatrix*: cardiotoxin and  $\alpha$ -neurotoxin (Ma et al. 2001). Besides varying in function, these two isoforms have different expression levels in the venom gland. Cardiotoxin constitutes 60% of the venom while the  $\alpha$ -neurotoxin makes up only 3% of the venom. Gene duplication is evident from the gene comparison whereby the structures and amino acid sequence of these

two toxins are very well-conserved (Ma et al. 2002). The main difference lies in the promoter segment, where it was found that the  $\alpha$ -neurotoxin promoter contains a stronger silencer element, which is responsible for significantly reducing its expression level in the venom (Ma et al. 2001; Ma et al. 2002).

In the case of venom prothrombin activators, we have shown that they have been "recruited" from the gene of an ancestral plasma prothrombin activator protein through gene duplication. The duplicated gene underwent modifications in its regulatory and coding regions to gain toxin characteristics. *VERSE* segments were inserted in the promoter regions of trocarin D and PCCS and are responsible for their elevated level of expression. Insertion/deletion segments in their intron 1 regions are postulated to be responsible for venom-gland specific expression. Modifications in the gene-coding regions enable prothrombin activators to function better as toxin by gaining certain characteristics such as resistance to inactivation.

## 7. Conclusion

Gene duplication has played a major role in the development of snake venom toxins. Our findings on venom prothrombin activators and blood coagulation factors have captured the first molecular evidence of gene duplication. The characterization of the differences in their genes, i.e. *VERSE* segment and intron 1 of trocarin D, has increased our understanding of gene regulation of snake venom toxins. It is shown that the *VERSE* segment is responsible for the elevation of gene expression and that the intron 1 is probably responsible venom gland-specific expression (unpublished observations). We identified three novel *cis*-elements in the *VERSE* segment, and these play important roles in gene regulation. It would be interesting to further characterize them and their *trans*-factor partners to determine how various *trans*-factors interact with each other to regulate gene expression. The answers to some of these questions will increase our overall understanding of gene regulation.

## 8. References

- Bos, M.H., M.Boltz, L.St Pierre, P.P.Masci, J.de Jersey, M.F.Lavin, and R.M.Camire. 2009. "Venom factor V from the common brown snake escapes hemostatic regulation through procoagulant adaptations." *Blood*. 114:686-692.
- Boulikas, T. 1993. "Nature of DNA sequences at the attachment regions of genes to the nuclear matrix." *J.Cell Biochem*. 52:14-22.
- Braud, S., C.Bon, and A.Wisner. 2000. "Snake venom proteins acting on hemostasis." *Biochimie*. 82:851-859.
- Chester, A. and G.P.Crawford. 1982. "In vitro coagulant properties of venoms from Australian snakes." *Toxicon*. 20:501-504.
- Chow, G. and R.M.Kini. 2001. "Exogenous factors from animal sources that induce platelet aggregation." *Thromb.Haemost*. 85:177-178.
- Cockerill, P.N. and W.T.Garrard. 1986. "Chromosomal loop anchorage of the kappa immunoglobulin gene occurs next to the enhancer in a region containing topoisomerase II sites." *Cell*. 44:273-282.

- Davie, E.W. 2003. "A brief historical review of the waterfall/cascade of blood coagulation." *J.Biol.Chem.* 278:50819-50832.
- Esmon, C.T. 2001. "Role of coagulation inhibitors in inflammation." *Thromb.Haemost.* 86:51-56.
- Filippovich, I., N.Sorokina, L.St Pierre, S.Flight, J.de Jersey, N.Perry, P.P.Masci, and M.F.Lavin. 2005. "Cloning and functional expression of venom prothrombin activator protease from *Pseudonaja textilis* with whole blood procoagulant activity." *Br.J.Haematol.* 131:237-246.
- Foster, W.B., M.E.Nesheim, and K.G.Mann. 1983. "The factor Xa-catalyzed activation of factor V." *J.Biol.Chem.* 258:13970-13977.
- Fry, B.G. 2005. "From genome to "venome": molecular origin and evolution of the snake venom proteome inferred from phylogenetic analysis of toxin sequences and related body proteins." *Genome Res.* 15:403-420.
- Fry, B.G., K.Roelants, D.E.Champagne, H.Scheib, J.D.Tyndall, G.F.King, T.J.Nevalainen, J.A.Norman, R.J.Lewis, R.S.Norton, C.Renjifo, and R.C.de la Vega. 2009. "The toxicogenomic multiverse: convergent recruitment of proteins into animal venoms." *Annu.Rev.Genomics Hum.Genet.* 10:483-511.
- Fry, B.G., W.Wuster, R.M.Kini, V.Brusic, A.Khan, D.Venkataraman, and A.P.Rooney. 2003. "Molecular evolution and phylogeny of elapid snake venom three-finger toxins." *J.Mol.Evol.* 57:110-129.
- Gao, R., R.M.Kini, and P.Gopalakrishnakone. 2002. "A novel prothrombin activator from the venom of *Micropechis ikaheka*: isolation and characterization." *Arch. Biochem. Biophys.* 408:87-92.
- Hanover, J.A., C.K.Cohen, M.C.Willingham, and M.K.Park. 1987. "O-linked N-acetylglucosamine is attached to proteins of the nuclear pore. Evidence for cytoplasmic and nucleoplasmic glycoproteins." *J.Biol.Chem.* 262:9887-9894.
- Hasson, S.S., R.D.Theakston, and R.A.Harrison. 2003. "Cloning of a prothrombin activator-like metalloproteinase from the West African saw-scaled viper, *Echis ocellatus*." *Toxicon.* 42:629-634.
- Holt, G.D., R.S.Haltiwanger, C.R.Torres, and G.W.Hart. 1987. "Erythrocytes contain cytoplasmic glycoproteins. O-linked GlcNAc on Band 4.1." *J.Biol.Chem.* 262:14847-14850.
- Holt, G.D. and G.W.Hart. 1986. "The subcellular distribution of terminal N-acetylglucosamine moieties. Localization of a novel protein-saccharide linkage, O-linked GlcNAc." *J.Biol.Chem.* 261:8049-8057.
- Hung, H.L. and K.A.High. 1996. "Liver-enriched transcription factor HNF-4 and ubiquitous factor NF-Y are critical for expression of blood coagulation factor X." *J.Biol.Chem.* 271:2323-2331.
- Hung, H.L., E.S.Pollak, R.D.Kudaravalli, V.Arruda, K.Chu, and K.A.High. 2001. "Regulation of human coagulation factor X gene expression by GATA-4 and the Sp family of transcription factors." *Blood.* 97:946-951.
- Hutton, R.A. and D.A.Warrell. 1993. "Action of snake venom components on the haemostatic system." *Blood Rev.* 7:176-189.



- Inoue, K. and T.Morita. 1993. "Identification of O-linked oligosaccharide chains in the activation peptides of blood coagulation factor X. The role of the carbohydrate moieties in the activation of factor X." *Eur.J.Biochem.* 218:153-163.
- Joseph, J.S., M.C.Chung, K.Jeyaseelan, and R.M.Kini. 1999. "Amino acid sequence of trocarin, a prothrombin activator from *Tropidechis carinatus* venom: its structural similarity to coagulation factor Xa." *Blood.* 94:621-631.
- Joseph, J.S. and R.M.Kini. 2001. "Snake venom prothrombin activators homologous to blood coagulation factor Xa." *Haemostasis.* 31:234-240.
- Kalafatis, M., M.D.Rand, and K.G.Mann. 1994. "The mechanism of inactivation of human factor V and human factor Va by activated protein C." *J.Biol.Chem.* 269:31869-31880.
- Kini, R.M. 2002. "Molecular moulds with multiple missions: functional sites in three-finger toxins." *Clin.Exp.Pharmacol.Physiol.* 29:815-822.
- Kini, R.M. 2004. "Platelet aggregation and exogenous factors from animal sources." *Curr.Drug Targets.Cardiovasc.Haematol.Disord.* 4:301-325.
- Kini, R.M. and G.Chow. 2001. "Exogenous inhibitors of platelet aggregation from animal sources." *Thromb.Haemost.* 85:179-181.
- Kini, R.M. and R.Doley. 2010. "Structure, function and evolution of three-finger toxins: mini proteins with multiple targets." *Toxicon.* 56:855-867.
- Kini, R.M. and H.J.Evans. 1990. "Effects of snake venom proteins on blood platelets." *Toxicon.* 28:1387-1422.
- Kini, R.M., T.Morita, and J.Rosing. 2001. "Classification and nomenclature of prothrombin activators isolated from snake venoms." *Thromb.Haemost.* 86:710-711.
- Kornalik, F. and B.Blomback. 1975. "Prothrombin activation induced by Ecarin - a prothrombin converting enzyme from *Echis carinatus* venom." *Thromb.Res.* 6:57-63.
- Kwong, S., A.E.Woods, P.J.Mirtschin, R.Ge, and R.M.Kini. 2009. "The recruitment of blood coagulation factor X into snake venom gland as a toxin: the role of promoter Cis-elements in its expression." *Thromb.Haemost.* 102:469-478.
- Landan, G., A.Bdolah, Z.Wollberg, E.Kochva, and D.Graur. 1991a. "Evolution of the sarafotoxin/endothelin superfamily of proteins." *Toxicon.* 29:237-244.
- Landan, G., A.Bdolah, Z.Wollberg, E.Kochva, and D.Graur. 1991b. "Evolution of the sarafotoxin/endothelin superfamily of proteins." *Toxicon.* 29:237-244.
- Liebich, I., J.Bode, I.Reuter, and E.Wingender. 2002. "Evaluation of sequence motifs found in scaffold/matrix-attached regions (S/MARs)." *Nucleic Acids Res.* 30:3433-3442.
- Lynch, V.J. 2007. "Inventing an arsenal: adaptive evolution and neofunctionalization of snake venom phospholipase A2 genes." *BMC.Evol.Biol.* 7:2.
- Ma, D., A.Armugam, and K.Jeyaseelan. 2001. "Expression of cardiotoxin-2 gene. Cloning, characterization and deletion analysis of the promoter." *Eur.J.Biochem.* 268:1844-1850.
- Ma, D., A.Armugam, and K.Jeyaseelan. 2002. "Alpha-neurotoxin gene expression in *Naja sputatrix*: identification of a silencer element in the promoter region." *Arch.Biochem.Biophys.* 404:98-105.

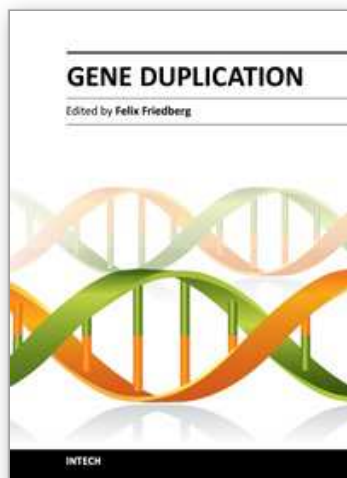
- Mann, K.G., M.F.Hockin, K.J.Begin, and M.Kalafatis. 1997. "Activated protein C cleavage of factor Va leads to dissociation of the A2 domain." *J.Biol.Chem.* 272:20678-20683.
- Markland, F.S. 1997. "Snake venoms." *Drugs.* 54 Suppl 3:1-10.
- Markland, F.S. 1998. "Snake venoms and the hemostatic system." *Toxicon.* 36:1749-1800.
- Marsh, N.A., T.L.Fyffe, and E.A.Bennett. 1997. "Isolation and partial characterization of a prothrombin-activating enzyme from the venom of the Australian rough-scaled snake (*Tropidechis carinatus*)." *Toxicon.* 35:563-571.
- Masci, P.P., A.N.Whitaker, and J.de Jersey. 1988. "Purification and characterization of a prothrombin activator from the venom of the Australian brown snake, *Pseudonaja textilis textilis*." *Biochem.Int.* 17:825-835.
- McMullen, B.A., K.Fujikawa, W.Kisiel, T.Sasagawa, W.N.Howald, E.Y.Kwa, and B.Weinstein. 1983. "Complete amino acid sequence of the light chain of human blood coagulation factor X: evidence for identification of residue 63 as beta-hydroxyaspartic acid." *Biochemistry.* 22:2875-2884.
- Minh, L.T., M.A.Reza, S.Swarup, and R.M.Kini. 2005. "Gene duplication of coagulation factor V and origin of venom prothrombin activator in *Pseudonaja textilis* snake." *Thromb.Haemost.* 93:420-429.
- Morita, T. 2004. "C-type lectin-related proteins from snake venoms." *Curr.Drug Targets.Cardiovasc.Haematol.Disord.* 4:357-373.
- Morita, T. and S.Iwanaga. 1978. "Purification and properties of prothrombin activator from the venom of *Echis carinatus*." *J.Biochem.(Tokyo).* 83:559-570.
- Moura-da-Silva, A.M., R.D.Theakston, and J.M.Crampton. 1996. "Evolution of disintegrin cysteine-rich and mammalian matrix-degrading metalloproteinases: gene duplication and divergence of a common ancestor rather than convergent evolution." *J.Mol.Evol.* 43:263-269.
- Nesheim, M.E., W.M.Canfield, W.Kisiel, and K.G.Mann. 1982. "Studies of the capacity of factor Xa to protect factor Va from inactivation by activated protein C." *J.Biol.Chem.* 257:1443-1447.
- Nesheim, M.E., J.B.Taswell, and K.G.Mann. 1979. "The contribution of bovine Factor V and Factor Va to the activity of prothrombinase." *J.Biol.Chem.* 254:10952-10962.
- Nishida, S., T.Fujita, N.Kohno, H.Atoda, T.Morita, H.Takeya, I.Kido, M.J.Paine, S.Kawabata, and S.Iwanaga. 1995. "cDNA cloning and deduced amino acid sequence of prothrombin activator (ecarin) from Kenyan *Echis carinatus* venom." *Biochemistry.* 34:1771-1778.
- Owen, W.G. and C.M.Jackson. 1973. "Activation of prothrombin with *Oxyranus scutellatus* scutellatus (Taipain snake) venom." *Thromb.Res.* 3:705-714.
- Pittman, D.D., K.N.Tomkinson, D.Michnick, U.Selighsohn, and R.J.Kaufman. 1994. "Posttranslational sulfation of factor V is required for efficient thrombin cleavage and activation and for full procoagulant activity." *Biochemistry.* 33:6952-6959.
- Rao, V.S., J.S.Joseph, and R.M.Kini. 2003a. "Group D prothrombin activators from snake venom are structural homologues of mammalian blood coagulation factor Xa." *Biochem.J.* 369:635-642.

- Rao, V.S. and R.M.Kini. 2002. "Pseutarin C, a prothrombin activator from *Pseudonaja textilis* venom: its structural and functional similarity to mammalian coagulation factor Xa-Va complex." *Thromb.Haemost.* 88:611-619.
- Rao, V.S., S.Swarup, and R.M.Kini. 2003b. "The nonenzymatic subunit of pseutarin C, a prothrombin activator from eastern brown snake (*Pseudonaja textilis*) venom, shows structural similarity to mammalian coagulation factor V." *Blood.* 102:1347-1354.
- Rao, V.S., S.Swarup, and R.M.Kini. 2004. "The catalytic subunit of pseutarin C, a group C prothrombin activator from the venom of *Pseudonaja textilis*, is structurally similar to mammalian blood coagulation factor Xa." *Thromb.Haemost.* 92:509-521.
- Reza, A., S.Swarup, and R.M.Kini. 2005a. "Two parallel prothrombin activator systems in Australian rough-scaled snake, *Tropidechis carinatus*. Structural comparison of venom prothrombin activator with blood coagulation factor X." *Thromb.Haemost.* 93:40-47.
- Reza, M.A., L.T.Minh, S.Swarup, and R.M.Kini. 2006. "Molecular evolution caught in action: gene duplication and evolution of molecular isoforms of prothrombin activators in *Pseudonaja textilis* (brown snake)." *J.Thromb.Haemost.* 4:1346-1353.
- Reza, M.A., S.Swarup, and R.M.Kini. 2005b. "Gene structures of trocarin D and coagulation factor X, two functionally diverse prothrombin activators from Australian rough scaled snake." *Pathophysiol.Haemost.Thromb.* 34:205-208.
- Reza, M.A., S.Swarup, and R.M.Kini. 2007. "Structure of two genes encoding parallel prothrombin activators in *Tropidechis carinatus* snake: gene duplication and recruitment of factor X gene to the venom gland." *J.Thromb.Haemost.* 5:117-126.
- Robin Doley, Xingding Zhou, and R.M.Kini. 2009. "Snake Venom Phospholipase A2 Enzymes." In Stephen P.Mackessy, editor, *Handbook of Venoms and Toxins of Reptiles*. CRC Press. 173-205.
- Rosing, J. and G.Tans. 1991. "Inventory of exogenous prothrombin activators. For the Subcommittee on Nomenclature of Exogenous Hemostatic Factors of the Scientific and Standardization Committee of the International Society on Thrombosis and Haemostasis." *Thromb.Haemost.* 65:627-630.
- Rosing, J. and G.Tans. 1992. "Structural and functional properties of snake venom prothrombin activators." *Toxicon.* 30:1515-1527.
- Schieck, A., E.Habermann, and F.Kornalik. 1972. "The prothrombin-activating principle from *Echis carinatus* venom. II. Coagulation studies in vitro and in vivo." *Naunyn Schmiedebergs Arch.Pharmacol.* 274:7-17.
- Silva, M.B., M.Schattner, C.R.Ramos, I.L.Junqueira-de-Azevedo, M.C.Guarnieri, M.A.Lazzari, C.A.Sampaio, R.G.Pozner, J.S.Ventura, P.L.Ho, and A.M.Chudzinski-Tavassi. 2003. "A prothrombin activator from *Bothrops erythromelas* (jararaca-da-seca) snake venom: characterization and molecular cloning." *Biochem.J.* 369:129-139.
- Snow, C.M., A.Senior, and L.Gerace. 1987. "Monoclonal antibodies identify a group of nuclear pore complex glycoproteins." *J.Cell Biol.* 104:1143-1156.

- Speijer, H., J.W.Govers-Riemslog, R.F.Zwaal, and J.Rosing. 1986. "Prothrombin activation by an activator from the venom of *Oxyuranus scutellatus* (Taipan snake)." *J.Biol.Chem.* 261:13258-13267.
- St Pierre, L., S.T.Earl, I.Filippovich, N.Sorokina, P.P.Masci, J.De Jersey, and M.F.Lavin. 2008. "Common evolution of waprin and kunitz-like toxin families in Australian venomous snakes." *Cell Mol.Life Sci.* 65:4039-4054.
- St Pierre, L., P.P.Masci, I.Filippovich, N.Sorokina, N.Marsh, D.J.Miller, and M.F.Lavin. 2005. "Comparative analysis of prothrombin activators from the venom of Australian elapids." *Mol.Biol.Evol.* 22:1853-1864.
- Stenflo, J., A.Lundwall, and B.Dahlback. 1987. "beta-Hydroxyasparagine in domains homologous to the epidermal growth factor precursor in vitamin K-dependent protein S." *Proc.Natl.Acad.Sci.U.S.A.* 84:368-372.
- Stocker, K., H.Hauer, C.Muller, and D.A.Triplett. 1994. "Isolation and characterization of Textarin, a prothrombin activator from eastern brown snake (*Pseudonaja textilis*) venom." *Toxicon.* 32:1227-1236.
- Suzuki, K., B.Dahlback, and J.Stenflo. 1982. "Thrombin-catalyzed activation of human coagulation factor V." *J Biol Chem.* 257:6556-6564.
- Tans, G., J.W.Govers-Riemslog, J.L.van Rijn, and J.Rosing. 1985. "Purification and properties of a prothrombin activator from the venom of *Notechis scutatus scutatus*." *J.Biol.Chem.* 260:9366-9372.
- Thorelli, E., R.J.Kaufman, and B.Dahlback. 1998. "Cleavage requirements of factor V in tissue-factor induced thrombin generation." *Thromb.Haemost.* 80:92-98.
- van der Neut, K.M., R.J.Dirven, H.L.Vos, G.Tans, J.Rosing, and R.M.Bertina. 2004b. "Factor Va is inactivated by activated protein C in the absence of cleavage sites at Arg-306, Arg-506, and Arg-679." *J.Biol.Chem.* 279:6567-6575.
- van der Neut, K.M., R.J.Dirven, H.L.Vos, G.Tans, J.Rosing, and R.M.Bertina. 2004a. "Factor Va is inactivated by activated protein C in the absence of cleavage sites at Arg-306, Arg-506, and Arg-679." *J.Biol.Chem.* 279:6567-6575.
- van Drunen, C.M., R.W.Oosterling, G.M.Keultjes, P.J.Weisbeek, R.van Driel, and S.C.Smeekens. 1997. "Analysis of the chromatin domain organisation around the plastocyanin gene reveals an MAR-specific sequence element in *Arabidopsis thaliana*." *Nucleic Acids Res.* 25:3904-3911.
- Walker, F.J., W.G.Owen, and C.T.Esmon. 1980. "Characterization of the prothrombin activator from the venom of *Oxyuranus scutellatus scutellatus* (taipan venom)." *Biochemistry.* 19:1020-1023.
- Wang, C., M.Eufemi, C.Turano, and A.Giartosio. 1996. "Influence of the carbohydrate moiety on the stability of glycoproteins." *Biochemistry.* 35:7299-7307.
- Welton, R.E. and J.N.Burnell. 2005. "Full length nucleotide sequence of a factor V-like subunit of oscutarin from *Oxyuranus scutellatus scutellatus* (coastal Taipan)." *Toxicon.* 46:328-336.
- Wilberding, J.A. and F.J.Castellino. 2000. "Characterization of the murine coagulation factor X promoter." *Thromb.Haemost.* 84:1031-1038.

- Will, K., G.Warnecke, N.Albrechtsen, T.Boulikas, and W.Deppert. 1998. "High affinity MAR-DNA binding is a common property of murine and human mutant p53." *J.Cell Biochem.* 69:260-270.
- Yamada, D. and T.Morita. 1997. "Purification and characterization of a Ca<sup>2+</sup> -dependent prothrombin activator, multactivase, from the venom of *Echis multisquamatus*." *J.Biochem.(Tokyo)*. 122:991-997.
- Zhou, C.Z. and B.Liu. 2001. "Identification and characterization of a silk gland-related matrix association region in *Bombyx mori*." *Gene*. 277:139-144.





### **Gene Duplication**

Edited by Prof. Felix Friedberg

ISBN 978-953-307-387-3

Hard cover, 400 pages

**Publisher** InTech

**Published online** 17, October, 2011

**Published in print edition** October, 2011

The book Gene Duplication consists of 21 chapters divided in 3 parts: General Aspects, A Look at Some Gene Families and Examining Bundles of Genes. The importance of the study of Gene Duplication stems from the realization that the dynamic process of duplication is the "sine qua non" underlying the evolution of all living matter. Genes may be altered before or after the duplication process thereby undergoing neofunctionalization, thus creating in time new organisms which populate the Earth.

### **How to reference**

In order to correctly reference this scholarly work, feel free to copy and paste the following:

Shiyang Kwong and R. Manjunatha Kini (2011). Duplication of Coagulation Factor Genes and Evolution of Snake Venom Prothrombin Activators, Gene Duplication, Prof. Felix Friedberg (Ed.), ISBN: 978-953-307-387-3, InTech, Available from: <http://www.intechopen.com/books/gene-duplication/duplication-of-coagulation-factor-genes-and-evolution-of-snake-venom-prothrombin-activators>

**INTECH**  
open science | open minds

### **InTech Europe**

University Campus STeP Ri  
Slavka Krautzeka 83/A  
51000 Rijeka, Croatia  
Phone: +385 (51) 770 447  
Fax: +385 (51) 686 166  
[www.intechopen.com](http://www.intechopen.com)

### **InTech China**

Unit 405, Office Block, Hotel Equatorial Shanghai  
No.65, Yan An Road (West), Shanghai, 200040, China  
中国上海市延安西路65号上海国际贵都大饭店办公楼405单元  
Phone: +86-21-62489820  
Fax: +86-21-62489821

© 2011 The Author(s). Licensee IntechOpen. This is an open access article distributed under the terms of the [Creative Commons Attribution 3.0 License](https://creativecommons.org/licenses/by/3.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

IntechOpen

IntechOpen