We are IntechOpen, the world's leading publisher of Open Access books Built by scientists, for scientists



186,000

200M



Our authors are among the

TOP 1% most cited scientists





WEB OF SCIENCE

Selection of our books indexed in the Book Citation Index in Web of Science™ Core Collection (BKCI)

Interested in publishing with us? Contact book.department@intechopen.com

Numbers displayed above are based on latest data collected. For more information visit www.intechopen.com



Androgens and Vascular Function

Mercedes Ferrer Departamento de Fisiología, Facultad de Medicina, Universidad Autónoma de Madrid and Instituto de Investigación Sanitaria IdiPAZ Spain

1. Introduction

It is widely recognized that vascular function is modulated by the endocrine system. So it is known that hormones as aldosterone, renin-angiotensin II system, thyroids hormones, oxitocine, ghrelin, vasopressine, etc exert action on the vascular tone (Axelband et al., 2011; Nguyen & Touyz, 2011; Szmydynger-Chodobska et al., 2011; Tesauro et al., 2010;). Regarding sex hormones, epidemiological studies have demonstrated that there is a gender difference in the morbidity associated with hypertension and that there is an increased prevalence of cardiovascular diseases in postmenopausal women (Leung et al., 2007; Teede, 2007). Simultaneously, androgens had been associated with an increased risk of cardiovascular disease, but recent studies have explored protective effects of androgens in males (Jones, 2010; Traish and Kypreos, 2011). For example, it has been demonstrated that men with coronary artery disease have decreased levels of testosterone, which were conversely correlated to the degree of coronary artery narrowing (Saad et al., 2008). Likewise, lower testosterone, predicted incident stroke and transient ischemic attack in older men (Braga-Basaria, 2006). In general terms, it seems to be demonstrated that men with cardiovascular disease had lower levels of testosterone, and what is more important, new emerging evidence points to androgen deficiency more likely to be associated with cardiovascular diseases than gender per se (Traish & Kypreos, 2011). Testosterone deficiency alters carbohydrate, lipid and protein metabolism, this contributing to oxidative stress, endothelial dysfunction and increased production of pro-inflammatory factors, promoting alterations on vascular function. Among the alterations induced by testosterone deficiency are the loss of muscle mass and strength, increasing visceral fat mass, reduced libido, erectile dysfunction, increased osteoporosis, lethargy, lack of energy, and changes in mood. In addition, testosterone deficiency has been associated with increased risk of metabolic syndrome, type 2 diabetes, obesity, insulin resistance and atherosclerosis (Jones & Saad, 2009; Kapoor et al., 2005).

Most of results obtained about that effects have been obtained from patients with prostate cancer subjected to androgen deprivation therapy. This therapy improves cancer related symptoms and quality of life (Bain, 2010), but shows side effects as sexual dysfunction, decreased lean body mass, decreased quality of life, osteoporosis, and detrimental changes in metabolic status (Basaria & Dobs, 2001; Chodak *et al.*, 2002; Smith *et al.*, 2002). Increased insulin levels and insulin resistance and increased prevalence of fasting hyperglycemia and hypertriglyceridemia have also been observed in men with prostate cancer treated with

androgen deprivation therapy (Braga-Bassaria *et al.*, 2006; Keating *et al.*, 2006). This type of therapy provides an invaluable method to correlate vascular alteration and sex hormone status. However, at mechanistic level the animal models are of valuable interest. In this sense our research group has been focused on analyzing the effects produced by the loss of gonadal function on vascular reactivity, as well as some of the underlaying mechanisms.

Vascular tone is regulated by several mechanisms in which nitric oxide (NO) plays an important role. NO is formed through several NO synthases (NOS), i.e., endothelial NOS (eNOS), inducible (iNOS), and neuronal (nNOS). Independently from the source of NO, one of the major downstream events occurring after NO release is an increase in cGMP formation through soluble guanylate cyclase stimulation, and the subsequent activation of cGMP-dependent protein kinase (PKG) (Murad, 1997). PKG contributes to reduce the intracellular calcium concentration through a wide spectrum of PKG substrates, leading to vasodilation (Lincoln *et al.*, 2001; Munzel *et al.*, 2003). In addition, NO (Bolotina *et al.*, 1994) and cGMP (Ferrer *et al.*, 1995) can induce membrane hyperpolarization and subsequent relaxation.

Concerning NO, and independently from the NO-activated signalling pathway, it is important to take into account that vascular function of endothelial NO depends on its bioavailability, which is determined by the rate of NO production and by its scavenging by superoxide anion. Therefore, the elimination of superoxide anion within the vessel wall is fundamental and it is performed by superoxide dismutases (SODs) (Wolin, 2002). It is well understood that alterations of different steps along the NO pathway determine its effect on the vascular tone.

Vascular tone is also regulated by prostanoids originated by arachidonic acid metabolism through the cyclooxygenase (COX) pathway (Henrion *et al.*, 1997). One of the best studied prostanoids is thromboxane A_2 (TXA₂) which has been implicated as mediator in diseases such as myocardial infarction and hypertension (FitzGerald et al., 1987). Additionally, the role of COX derivatives other than TXA₂, such as prostaglandin (PG) $F_{2\alpha}$ and PGE₂, or PGI₂ which can induce a vasoconstrictor or vasodilator response is the subject of numerous studies (Blanco-Rivero *et al.*, 2005; Félétou & Vanhoutte, 2006) since they could also participate in vascular dysfunction.

On the other hand, proteins kinases are important regulators of different cell signalling pathways. Among different proteins kinases, PKC merits special attention since it is able to regulate the activity of different enzymes such as NOS and COX-2 (Kim *et al.*, 2008; Shanmugam *et al.*, 2004). In turn, PKC can be activated by different reactive oxygen species. At this point, it is important to note that sex hormones possesses antioxidant properties, and the loss of gonadal function can induce oxidative stress and, in turn triggers modulatory actions in different cell signalling pathways that are working simultaneously to ensure the optimal response of the vessel to different stimuli.

For this, this chapter will review how the loss of gonadal function modifies the release and function of different mediators, such as reactive oxygen species, nitric oxide and prostanoids, and their functional involvement in the reactivity of aorta and mesenteric artery of the male rats. Future studies in the research field of androgens on cell signaling pathways will be commented, since they will be of important interest to implement therapeutic strategies that could improve vascular function.

2. Androgens and nitric oxide

The functional role of NO in vascular tone regulation has been widely reported (Furchgott & Zawadzki, 1980; Toda & Okamura, 2003; Vanhoutte 1996). NO is formed through several

NO synthases (NOS), i.e, endothelial NOS (eNOS), inducible (iNOS) and neuronal (nNOS) (Förstermann *et al.*, 1991). Most of the studies about the effects of androgens on vascular function have been focused on analyzing the interaction between androgens and endothelial NO (Tep-areenan *et al.*, 20043; Jones *et al.*, 2004). More specifically, most studies have been focused on analyzing the effects of androgenic derivatives on different aspects of the NO system, such us eNOS expression, NO release or NO vasodilator effect (Ceballos *et al.*, 1999; Hutchison *et al.*, 1997; Teoh *et al.*, 2000). However, few studies exist about the specific effect of endogenous male sex hormones on these aspects referred to endothelial or neuronal NO when they are simultaneously studied. Our data were obtained from aorta and mesenteric artery from intact and orchidectomized male Sprague-Dawley rats (6 months old), and seem not to be related to haemodynamic changes, since orchidectomy did not modify blood pressure (137 ± 5.8 mm Hg in control rats and of 145 ± 6.2 mm Hg in orchidectomized rats; *p* > 0.05). Moreover, the results on vascular function would have to be androgen-related, as confirmed by the decreased testosterone levels (control: 2404 ± 323 pg/mL; orchidectomized: 220 ± 49 pg/mL; n = 6; *p* < 0.001).

The reported effects of androgens on NO release are contradictory. Testosterone impairs relaxation and worsens endothelial dysfunction in male rabbits (Hutchison *et al.*, 1997). However, it has also been reported that testosterone or its derivatives increase eNOS (Liu & Dillon, 2002; Weiner *et al.*, 1994) and nNOS activity in the central nervous system of guinea pig (Weiner *et al.*, 1994) and mouse (Scordalakes *et al.*, 2002). Nevertheless, to our knowledge, we showed first evidence on the effect of endogenous male sex hormones on the nNOS/eNOS expression and nNOS-/eNOS-derived NO release in vascular tissues (Martin et al., 2005; Blanco-Rivero et al., 2007).

2.1 Neuronal nitric oxide

The analysis of the expression of nNOS, by Western blot, showed that it was higher in segments from control rats than from orchidectomized rats (Martín *et al.*, 2005). The NO release was quantified by measuring nitrite production and also the fluorescence emitted by DAF-2, and the results showed that electrical field stimulation EFS-induced NO release was similar in segments from both control and orchidectomized rats. The measured NO release seems to come from nerve endings, since preincubation with tetrodotoxin abolished the nitrite release in endothelial-denuded segments from both groups of rats. This result indicates that male sex hormones apparently do not modulate nNOS-derived NO release, in contrast to our observations with female sex hormones in which we found that the loss of these hormones provoked by ovariectomy increased nNOS-derived NO release (Minoves *et al.*, 2002).

To study the involvement of neuronal or endothelial NO on vasomotor responses, EFS induced contractile responses in endothelium-denuded mesenteric segments were analyzed by using vascular reactivity technique (Nielsen & Owman, 1971). The contractile response induced by EFS were practically abolished by tetrodotoxin and markedly reduced by phentolamine, the respective blockers for nerve impulse propagation and α -adrenoceptors in arteries from both control and orchidectomized rats. Therefore, these responses appear to be mediated by noradrenaline (NA) release from adrenergic nerve terminals and the subsequent activation of α -adrenoceptors in both experimental hormonal conditions, as has been described in other rat strains (Ferrer *et al.*, 2000; 2001).

The present results show that vasoconstrictor response to KCl and exogenous NA are diminished in segments from orchidectomized male rats, which is in agreement with most studies demonstrating that testosterone treatment enhanced the action of several contractile agents (Baker *et al.*, 1978; Calderone *et al.*, 2002; Greenberg *et al*, 1974). However, the responses induced by EFS was similar in both groups of rats Fig. 1 which would suggest that EFS increased the release of a vasoconstrictor factor, that could be the NA release from adrenergic endings in segments from orchidectomized animals.

Concerning this point, most studies have been performed in the central nervous system and, conflicting results exist. Thus, a decrease (Guan &Dluzen, 1991; Holmquist *et al.*, 1994), an increase (Shan & Dluzen, 2002; Siddiqui & Shah, 1997) and no modification (Agostini *et al.*, 1981; Chen *et al.*, 1999) of NA release have been reported in orchidectomized male animals. In a later study we demonstrated that the NA release was not modified by orchidectomy (Blanco-Rivero *et al.*, 2006).



Fig. 1. Contractile response induced by cumulative concentration-response curves to noradrenaline (NA) and by frequency-response curves in denuded mesenteric arteries from control and orchidectomized rats. Results (means \pm SEM) are expressed in milligram. Number of animals: 25-30. * *p*<0.01 compared with control rats (from Martín et al., 2005).

Since sex hormone modulation of the calcitonin gene related peptide (CGRP) system has been described (Sun *et al.*, 2001), and that the CGRP, the essential neurotransmitter in sensory nerves (Kawasaki *et al.*, 1988), had been proposed to play a role in vascular tone regulation, the participation of sensory innervation in the vasomotor response to EFS was assessed by the use of capsaicin, which selectively depletes the sensory nerves of the neurotransmitter (Li & Duckles, 1992). Capsaicin did not have a significant effect on the vasomotor response to either EFS or exogenous NA in mesenteric arteries from control and orchidectomized rats (data not shown), indicating that sensory innervation does not modulate the vasomotor response to EFS in our experimental conditions. Sun et al., (2001) analyzed how orchidectomy modulates CGRP release, while we analyzed the functional involvement of the sensory innervation in the response to EFS without separately studying CGRP release and/or response.

The NOS inhibitor L-NAME increased the vasoconstrictor response to EFS in segments from both control and orchidectomized rats. The fact that the endothelium was removed and that AMT, an inhibitor of iNOS (Ferrer *et al.*, 2000; Ishikawa & Quock, 2003; Tracey *et al.*, 1995), did not modify the response induced by EFS in segments from both rat groups, reinforced the neuronal origin for the NO release.



Fig. 2. Effect of L-NAME on the frequency-response curves performed in mesenteric arteries from control and orqchidectomized rats. Results are expressed as a percentage of a previous tone with 75 mM KCl. n, number of animals; *p<0.05; **p<0.01 compared with control rats (from Martín et al., 2005).

The greater response to EFS in the presence of L-NAME in segments from orchidectomized animals compared with the response obtained in segments from control rats Fig. 2 indicated differences in steps downstream from neuronal NO release, that will be considered later.

The reported effects of androgens on NO release are contradictory. Testosterone impairs relaxation and worsens endothelial dysfunction in male rabbits (Hutchison *et al.*, 1997). However, it has also been reported that testosterone or its derivatives increase eNOS (Liu & Dillon, 2002; Weiner *et al.*, 1994) and nNOS activity in the central nervous system of guinea pig (Weiner *et al.*, 1994) and mouse (Scordalakes *et al.*, 2002). Nevertheless, to our knowledge, there is no experimental evidence on the effect of male sex hormones on the nNOS expression and nNOS-derived NO release in vascular tissues. Therefore, we analysed the expression of nNOS by western blot, and found it was higher in segments from control rats than from orchidectomized rats. We also quantified the NO release induced by EFS in segments from both groups of rats by measuring nitrite production and the fluorescence emitted by DAF-2, and the results showed that the EFS-induced NO release was similar in segments from both control and orchidectomized rats.

2.2 Endothelial nitric oxide

The eNOS expression in aorta and mesenteric artery showed that not was modified by orchidectomy (Blanco-Rivero *et al.*, 2007; Martorell *et al.*, 2008), in contrast to that observed in endothelial-denuded mesenteric artery (Martín *et al.*, 2005), which indicates that endogenous male sex hormones act in a different manner depending on the target protein. Additionally, these results also indicate that effects induced by endogenous hormones are quite different from those induced by exogenous hormones, since it has been reported that androgenic derivatives can increase (Simoncini *et al.*, 2003), decrease (Chatrath *et al.*, 2003) or not affect (McNeill *et al.*, 1999) eNOS expression.

Regarding eNOS activity, androgen-induced increase (Liu & Dillon, 2002; Simoncini *et al.*, 2003) and decrease (Mukherjee *et al.*, 2001) have both been reported. Others researchers have

demonstrated that androgens increased (Orshal & Khalil, 2004; Wynne & Khalil, 2003) and decreased (Ba *et al.*, 2001; Gonzales *et al.*, 2004) the vasodilator effect of endothelial NO. In our experimental rat model, we have found that the release of endothelial NO was not modified by orchidectomy, either in mesenteric artery (Blanco-Rivero *et al.*, 2007) or aorta (Blanco-Rivero *et al.*, 2006). However, orchidectomy reduced the endothelium-dependent vasodilator response induced by Acetylcholine (Ach) in mesenteric artery while increased in rat aorta (Fig. 3).



Fig. 3. Concentration-response curve to acetylcholine (ACh) in mesenteric artery and aortic segments from control and orchidectomized rats. Results are expressed as a percentage of a previous tone with noradrenaline. Number of animals is indicated in parentheses (from Blanco-Rivero *et al.*, 2007; Ferrer *et al.*, 1999).

These results could indicate that orchidectomy, depending on the vessel, may alter the sensitivity to NO and/or the release of factors other than NO, that will be revised in the next sections.

3. Androgens and production of reactive oxygen species

Reactive oxygen species are involved in metabolising NO (Gryglewski *et al.* 1986, Ferrer *et al.* 2000, 2001) and it has been reported that can induce oxidative processes associated with cardiovascular disorders (Harrison 1994; Munzel *et al.*, 1997). Among all the reactive oxygen species, superoxide anion plays a critical role since it is a source of many other reactive oxygen intermediates (Beckman & Koppenol, 1996). Therefore, both production and removal of superoxide anion are important contributors to the maintenance of appropriated levels of this oxygen species. Since several studies have demonstrated androgens antioxidants properties of androgens (Békési *et al.*, 2000; Yorek *et al.*, 2002), the effect of the orchidectomy on the production of superoxide anion was analyzed, by measuring the lucigenin chemiluminescence (control =117 \pm 43 U/mg/min, n= 5; orchidectomized= 546 \pm 58 U/mg/min, n= 5; p < 0.001). A similar result was obtained with acetylcholine-induced superoxide anion in aortas from the same animals (control: 34.7 \pm 5.6 U/mg/ min, n= 4; orchidectomized: 132.8 \pm 3.2, n= 4; p< 0.001). These results agree with previous studies showing the antioxidants properties of androgens (Békési *et al.*, 2000; Yorek *et al.*, 2002). The elimination of superoxide anion within the vessels is fundamental since this oxygen specie can reduce the NO bioavailability.

Within the vessel wall, SODs transform superoxide anion to hydrogen peroxide (Oury *et al.*, 1996, Price *et al.*, 2000, Muzykantov, 2001). Although three SOD isoforms have been identified: cytosolic Cu/ZnSOD, mitochondrial MnSOD, and extracellular ecSOD –which is also Cu/Zn-dependent (Strehlow *et al.*, 2003), we have focused on analyzing the participation of Cu/ZnSOD since it is the predominant isoform in peripheral vessels (Namgaladze *et al.*, 2005), and this enzyme therefore plays a crucial role in the pathogenesis of vascular dysfunction (Wolin, 2002). We analyzed the expression and activity of SODs because there was no information about modulation of these enzymes by endogenous male sex hormones. We found that both the expression and activity of Cu/ZnSOD were increased in aortic segments from orchidectomized rats (Fig. 4). These results were in line with other studies that described an increased expression and/or activity of SOD in cardiovascular pathologies, in which superoxide anion overproduction exists (Kobayashi *et al.*, 2002, Tanaka *et al.*, 2005).



Fig. 4. Representative western blot for Cu/ZnSOD and) α -actin expression and SOD activity in aortic segments from control (C) and from orchidectomized (Cx) rats (from Blanco-Rivero *et al.*, 2006).

In addition, these results indicated that the increases in both expression and activity of Cu/ZnSOD could be a compensatory mechanism to eliminate the elevated superoxide anion formation induced by orchidectomy in male rats. On the other hand, additional mechanisms may exist to try to maintain vasodilator function. One of these mechanisms could be the activation of calcium-dependent potassium channels by superoxide anion (Ferrer *et al.*, 1999). In that report, we confirmed that the calcium dependent potassium channels (K_{Ca}) activation was specifically induced by superoxide anion, since the NO synthesis was inhibited.

However, the vascular response depends on the vascular bed analyzed. For example, in mesenteric artery from orchidectomized rats, the vascular ROS-induced relaxation was due to the production of peroxynitrite and hydrogen peroxide H_2O_2 . The increased formation of peroxynitrite in arteries from orchidectomized rats was studied by immunohistochemical localization and was reinforced by functional analysis on vasodilator response induced by sodium nitroprusside (SNP):

The presence of SOD increased the response to SNP in segments from orchidectomized rats which could be mediated through the decreased peroxynitrite formation by removing superoxide anion, and the simultaneous increased of H_2O_2 formation, which exert

vasodilator action (Martín *et al.*, 2005; Rubanyi & Vanhoutte, 1986; Wei *et al.*, 1996). The simultaneous incubation of arteries with SOD and catalase reversed the vasodilator response to SNP, indicating that the H₂O₂ synthesized in the presence of SOD, would participate in that vasodilator effect, when the formation of peroxynitrite is inhibited (Fig. 5).



Fig. 5. Effect of orchidectomy, and effect of superoxide dismutase (SOD) and SOD plus catalase on the concentration-response curves to sodium nitroprusside (SNP) in mesenteric artery segments from orchidectomized rats. Results are expressed as a percentage of the inhibition of contraction induced by noradrenaline. Number of animals is indicated in parentheses. *p<0.01 compared with control (from Martín et al., 2005).

At this point, it has been discussed that the loss of male sex hormones did not modify:

(1) the nNOS-derived NO release induced by EFS although it did increase the NO metabolism through superoxide anion and peroxynitrite generation; however, the functional role of the nNOS-derived NO release was more pronounced in arteries from orchidectomized animals, due to products generated from the NO metabolism, such as peroxynitrite and hydrogen peroxide, that seem to be able to compensate for the loss of NO bioavailability, probably through their direct vasodilator effect,

(2) the eNOS-derived NO release induced by Ach in both aorta and mesenteric artery; however the functional role of the endothelial NO release seem to be different in both arteries: Ach-induced relaxation is totally dependent of NO because the Ach-induced relaxation is inhibited when the synthesis of NO is blocked (Blanco-Rivero *et al.*, 2007), as occur in aorta from control rats, while in aorta from orchidectomized rats important relaxation exist even after NO synthesis was blocked (Ferrer *et al.*, 1999).

These differences could be due to the existence of regulatory mechanisms turned on by the loss of gonadal function that include, at least, prostanoids production.

4. Androgens and prostanoids

Endothelial cells also release vasoconstrictor and vasodilator prostanoids, originated from the arachidonic acid metabolism through the cyclooxygenase (COX) pathway, to regulate vascular tone (Blanco-Rivero *et al.*, 2005; Feletou & Vanhoutte, 2006; Henrion *et al.*, 1997).

Two major isoforms of COX exist: COX-1 is expressed constitutively and is usually abundant in all animal and human endothelial cells, whereas endothelial COX-2 is induced mainly during inflammatory responses in nearly all animals. Depending on the vessel studied and the agonist used to activate cells, different prostanoids can be released and contribute to vasomotor response depending on the prostanoid receptor that result activated. Among all the prostanoids, one of the most studied is thromboxane A₂ (TXA₂), which has been implicated as a mediator in diseases such as myocardial infarction, hypertension, stroke and bronchial asthma (FitzGerald *et al.*, 1987; Narumiya *et al.*, 1999; Noll& Luscher, 1998). However, little information is available on the effect of androgens on vascular effects of endogenous TXA₂. Thus, orchidectomy has been reported to either decrease (Gonzales *et al.*, 2005) or not modify (Blanco-Rivero *et al.*, 2006) TXA₂ synthase expression. Likewise, the contractile effect induced by the TXA₂ mimetic, U-46619, was not modified by orchidectomy in mesenteric (Blanco-Rivero *et al.*, 2006) and cerebral (Gonzales *et al.*, 2005) rat arteries.

On the other hand, the role of COX-derivates, other than TXA₂, such as prostaglandin (PG) I_{2} , PGF_{2 α} and PGE₂, which can act as vasoconstrictors under certain pathological conditions, is the objective of numerous studies (Félétou et al., 2010; Gluais et al., 2005; Rapoport & Williams, 1996). However, to the best of our knowledge studies analyzing the effect of endogenous male sex hormones in the involvement of these prostanoids in vascular function were lacking, and therefore this was one of our objectives. As above commented, we have previously demonstrated that EFS induced similar contractile response (Fig.1) in mesenteric arteries from control and orchidectomized rats, responses that appear to be mediated by NA release from adrenergic nerve terminals and the subsequent activation of α -adrenoceptors (Martín *et al.*, 2005); in addition, we found that the contractile response to exogenous NA was decreased by orchidectomy, suggesting the EFS could increase NA release in arteries from orchidectomized rats; however, we later demonstrated the EFSinduced NA release was not modified by orchidectomy (Blanco-Rivero et al., 2006) which indicated that other vasoconstrictor factors could be released when the artery was electrically stimulated. We observed that the EFS induced a greater TXA₂ formation in arteries from orchidectomized than control rats, which was in line with other reports showing increased TXA₂ release after activation of muscarinic (Blanco-Rivero et al., 2007) or α_2 -adrenoceptors (Blanco-Rivero *et al.*, 2006), and it confirmed the endothelial and smooth muscle cells as sources of TXA₂ production. The increased TXA₂ release could be the contractile factor that was released when the artery is electrically stimulated, and explains the non modification of the EFS-induced response in arteries from control and orchidectomized rats, in spite of the fact that the NA response was diminished in arteries from the latter animals.

It has been previously reported that prostanoids other than TXA₂, i.e. prostaglandin (PG) I₂ increased neuronal NO release (Ferrer *et al.*, 2004) and taking into account that in our experimental conditions the main neurotransmitters involved in that response elicited by EFS were NO and NA, we investigated the regulation of the NA and NO release and function by TXA₂. We observed that in arteries from control rats, the TXA₂ synthesis inhibition, with furegrelate, increased the neuronal NO release, which was in line with reports describing an inhibitory effect of TXA₂ on inducible (Yamada *et al.*, 2003) and endothelial (Miyamoto *et al.*, 2007) NO release. The vasodilator response induced by the NO donor, sodium nitroprusside was also increased by furegrelate. Concerning the effects of TXA₂ on NA release, inhibition (Nishihara *et al.*, 2000) or non-modification (Rump &

Schollmeyer, 1989) have been reported. In our experimental model, the inhibition of TXA2 synthesis did not modify either NA release or its vasomotor effect, which indicated that endogenous TXA₂ did not alter the function of sympathetic innervations on arteries from control rats. By contrast, in arteries from orchidectomized rats, endogenous TXA₂ did not regulate either the release/function of neuronal NO or NA. The results obtained in mesenteric arteries from control animals explain the decreased EFS-induced response in the presence of furegrelate; however, did not explain the unaltered EFS-induced response in arteries from orchidectomized rats, since to remove a constrictor substance a decreased in EFS-induced vasoconstriction would be expected. Since cross-talk between TXA₂ and PGI₂ has been reported (Cheng et al., 2002; Martorell et al., 2008) and joint increases in PGI2 and TXA2 synthesis have been shown in pathological conditions (Caughey et al., 2001; FitzGerald, 1991), the effect of TXA2 on PGI2 production was analyzed, as well as, its dependence of the hormonal status. We observed that the inhibition of endogenous synthesis of TXA₂ did not modify the release of PGI₂ in arteries from control rats, while it did greatly increase PGI₂ release in arteries from orchidectomized rats, which could work as a contractile factor after activation of TXA₂ receptors. Taking together these results, the loss of gonadal function in male rats increased the non-endothelial TXA₂ release in mesenteric arteries, and regulates the EFS-induced response through different mechanisms.

As previously commented, orchidectomy increased the TXA2 release in endothelial-intact mesenteric arteries (Blanco-Rivero et al., 2006; 2007). Similar effect was also observed in aorta segments from comparable animals (Martorell et al., 2008), however, in this case the relaxation to ACh was increased. Based on this result, the level of expression of COX-2 as well as the production of prostanoids derived from COX-2 other than TXA₂ was investigated. We found that COX-2 expression, in contrast to observations in mesenteric artery (Blanco-Rivero et al., 2006), was increased in aortas from orchidectomized rats indicating that endogenous male sex hormones act differently depending on the specific vessel. Our results also show that, in aorta from orchidectomized rats, COX-2 derivatives could also be increased and play a role in the regulation of vascular function. To test this hypothesis, we analyzed the effect of the COX-2 inhibitor NS-398 on the ACh-induced response. In contrast to our assumptions, we found that NS-398 did not modify the AChinduced relaxation in either group of rats, apparently indicating the lack of participation of COX-2 derived products in the ACh response, probably due to the equilibrium between the inhibition of prostanoids formation and phosphodiesterase inhibition (Klein et al., 2007), which allowed us to speculate that the contribution of different prostanoids to the vasodilator response mediated by ACh could be regulated by endogenous male sex hormones.

Once we had established that orchidectomy increased TXA₂ release, we analyzed the possible role of this prostanoid in the response to ACh by analyzing the effect of the TXA2 synthase inhibitor, furegrelate, and the TP receptor antagonist, SQ29,548, on the vasodilator response to ACh. We observed that neither substance had any effect on the ACh-induced response in arteries from control animals, indicating that TXA₂ did not participate in that response, in agreement with reports in other rat strains (Gluais *et al.*, 2005; Rapoport & Williams, 1996). However, in arteries from orchidectomized rats, furegrelate enhanced the vasodilator response to the TXA₂ mimetic U-46619 was similar in arteries from control and orchidectomized rats demonstrated that sensitivity to TXA2 is not modified by orchidectomy, which agrees with reports in cerebral (Gonzales *et al.*, 2005) and mesenteric

(Blanco-Rivero et al., 2006) arteries; additionally, it also shows that differences in the TXA2 involvement in the ACh-response are due to increased synthesis rather than increased sensitivity to TXA2. However, the incubation with SQ29,548 did not affect the ACh-induced relaxation, which seems to contradict the results obtained with furegrelate. However, since interactions among different prostanoids have been reported (Bachschmid et al., 2005; Cheng et al., 2002), it is possible to hypothesize that when TXA2 synthesis is inhibited, the production of other prostanoids, which counterbalance the TXA₂ effect, could be increased. Therefore, we investigated the effect of inhibiting PGI₂ synthesis (with TCP) on the AChinduced response, observing that it decreased the vasodilator response to ACh to a greater extent in arteries from orchidectomized than in those of control rats, which would indicate a greater involvement of this vasodilator prostanoid in the former arteries. Therefore, the release and vasomotor effect of PGI2 was investigated. The ACh-induced PGI2 release was increased in arteries from orchidectomized rats, probably due to the superoxide anion overproduction observed in aortas from orchidectomized rats (Blanco-Rivero et al., 2006), supporting the concept of redox regulation of vascular prostanoid synthesis proposed by Bachschmid et al. (2005). Moreover, the increased production of PGI₂ is in line with that reported in human syndromes involving platelet activation in which PGI₂ biosynthesis is elevated along with TXA₂ (Caughey et al., 2001; FitzGerald, 1991). It is known that PGI₂ can induce both vasodilation, through activation of prostacyclin receptors (IP) and thereby increasing cyclic-AMP, and vasoconstriction through activation of TP receptors (Blanco-Rivero et al., 2005). In the present study, we found that exogenous PGI₂ induced relaxation in rat aorta, and that it was decreased in arteries from orchidectomized rats, which could be due to differences in the expression of IP receptors rather than differences in cell signalling operating after receptor activation; we have observed that the relaxation induced by the activator of adenylate cyclase, forskolin, was similar in arteries from control and orchidectomized rats (unpublished data).

Since considerable evidence exists for cross-talk between the TXA₂ and PGI₂ systems (Cheng *et al.*, 2002), we analyzed the functional effect of inhibiting the synthesis of both prostanoids. We observed that co-incubation of arteries with TCP plus furegrelate, or TCP plus SQ29,548, reversed the decreased response to ACh caused by TCP in arteries from control rats, showing the existence of a balance between TXA₂ and PGI₂ in these arteries. However, in arteries from orchidectomized rats, the co-incubation with TCP plus furegrelate did not modify the decreased ACh response caused by TCP, indicating the participation of prostanoids other than PGI₂ and TXA₂ that could induce contraction. Moreover, these prostanoids would activate TP receptors since co-incubation with TCP and SQ29,548 completely reversed the decrease in the ACh response induced by TCP.

Among COX-2 derivatives, other than TXA₂ and PGI₂, that can activate TP receptors, PGE₂ is the most plausible candidate (Gluais *et al.*, 2005; Blanco-Rivero *et al.*, 2007) since the ACh-induced PGF_{2α} production and its vasoconstrictor effect were both very limited. Therefore, we investigated the ACh-induced PGE₂ release, as well as its vasoconstrictor effect. We found that both ACh-induced PGE₂ production and PGE₂-induced vasoconstrictor response were greater in arteries from orchidectomized than in those of control rats. Consequently, the ACh-induced PGE₂ release, under inhibited synthesis of TXA₂ and PGI₂, was analyzed. We found the ACh-induced PGE₂ production further increased, probably as a consequence of increased PGH₂ production and subsequent transformation into PGE₂ (Frein *et al.*, 2005); and, what is more important, the PGE₂ increase was more pronounced in arteries from orchidectomized than in those of control rats.



Fig. 6. Scheme showing the effect of orchidectomy on different mechanisms involved in the ACh-induced relaxation (see text for explanation).

This finding confirms our hypothesis that when the synthesis of PGI₂ and TXA₂ was inhibited, the release of PGE₂ was increased in arteries from orchidectomized rats, but raises the question as to why the PGE₂ produced in the presence of TCP plus furegrelate did not affect the AChinduced relaxation in arteries from control animals. The possible explanation could be that the PGE2 release was not sufficient to induce a vasomotor effect and/or the PGE2-induced contraction in arteries from control rats was diminished as a consequence of different expression of EP receptor subtypes. By itself, this finding is of physiological relevance, since PGE₂ release and the vasoconstrictor effect are both increased in orchidectomized animals. In summary, in rat aorta orchidectomy enhances COX-2 expression, and induces an imbalance in the production and function of vasodilator and vasoconstrictor prostanoids, in such a way that the vasoconstrictor prostanoids predominate in the latter group. Additionally, we have previously reported a decreased NO bioavailability in aorta from orchidectomized rats (Blanco-Rivero et al., 2006) that would also counteract the vasodilator response to ACh. However, despite these findings, the vasodilator response to ACh is increased in aorta from orchidectomized rats, probably as a consequence of compensatory mechanisms, such as the activation of K_{Ca} channels by superoxide anion, the formation of which is increased in orchidectomized rats (Ferrer et al., 1999) (Fig. 6).

5. Androgens and NO release, oxidative stress and protein kinase C interactions

The results described above point to that orchidectomy increases the oxidative stress through superoxide anion formation, but in contrast to that expected, without modifying the NO release. What is the mechanism underlying this effect?.

In several types of vascular diseases, protein kinase C (PKC) activation is involved in the induction of oxidative stress through the increased expression and activity of NADPH oxidase and the eNOS uncoupling (Hadi & Swaidi, 2007; Vanhoutte, 2001). But increased

of PKC activity by different reactive oxygen species has been also reported (Balafanova *et al.*, 2002; Bapat *et al.*, 2001; Oeckler & Wolin, 2000). PKC is a ubiquitous enzyme that was originally described as calcium-activated, phospholipid-dependent protein kinase. Molecular cloning and biochemical analysis have revealed a family of PKC subspecies with closely related structures (Newton, 1995; Nishizuka, 1992). The activity of these proteins is critical for signal transduction of a wide range of biological responses (Chen, 2003; Nishizuka, 1984). The pathways through PKC exerts its vascular effects include actions on ion channels, cytoskeleton and cell adhesion proteins, transcription factors, other kinases and other proteins (Spitaler & Cantrell, 2004; Ward *et al.*, 2004). The participation of PKC in vascular smooth muscle contraction is well established (Khalil & van Breemen, 1988; Salamanca & Khalil, 2005); in addition, different investigations have also reveal that PKC can phosphorylate nNOS in the central nervous system leading to an increase (Nakane *et al.*, 1991; Okada, 1995) or a decrease (Bredt *et al.*, 1992; Dawson *et al.*, 1993) in neuronal NO production.

On the other hand, influence of gender on PKC activity and expression had been studied (Kanashiro & Khalil, 2001). In that work, the authors observed a gender-specific reduction in vascular smooth muscle reactivity in female rats with intact gonadal function compared with males, that was associated with a reduction in the expression and activity of different α -, δ -, and ζ -PKC isoforms; they also proposed that gender-specific differences in vascular reactivity and PKC activity were possible related to endogenous estrogen. However, little information is available on the specific effect of endogenous male sex hormones on PKC activity in vascular tissues.

As described in the section corresponding to neuronal NO release, we observed that the neuronal NO release was not modified by orchidectomy although the nNOS expression was diminished, which indicated that nNOS activity could be increased; regarding regulation of nNOS by male sex hormones both increase (Simoncini *et al.*, 2003; Weiner *et al.*, 1994) or decrease (Reynoso *et al.*, 2002; Singh *et al.*, 2000) have been reported. Since in our experimental model, orchidectomy induced an increased oxidative stress, we analyzed the possible differences in PKC activity as well as its involvement in neuronal NO release.

We observed that orchidectomy increased PKC activity in rat mesenteric arteries (Blanco-Rivero *et al.*, 2005), in contrast to the results obtained in aorta from Wistar-Kyoto rats (Kanashiro & Khalil, 2001); this discrepancy could be due to differences in the artery analyzed, since it has been described different enzyme properties and function in the same blood vessels from different species and in different vessels from the same species (Liou & Morgan, 1994; Kanashiro & Khalil, 1988;Khalil *etal.*, 1992). However, the fact that PKC was increased by orchidectomy is not totally surprising, since orchidectomy increased the formation of superoxide anion and peroxynitrite that has been described to act in cell signalling pathways, for instance, increased PKC activity (Balafanova *et al.*, 2002; Bapat *et al.*, 2001; Oeckler & Wolin, 2000).

The regulatory effect of PKC on nNOS activity in arteries from the orchidectomized and the control animals was analyzed by using DAF-2 as fluorescence probe to measure the modification in basal and EFS-induced NO release. The results showed that the PKC activator PDBu (Abdel-Latif, 1986; Nishizuka, 1984) induced a greater increase in EFS-induced NO release in arteries from control than from orchidectomized animals, while the PKC inhibitor, calphostin C (Kobayashi *et al.*, 1989), induced a stronger decrease in arteries from the control than the orchidectomized animals (Fig. 7).



Fig. 7. Effect of PDBu, calphostin, Gö-6976 or PKC ζ -PI on the basal-and EFS-induced NO release in mesenteric arteries from control and orchidectomized rats. Results are expressed as arbitrary units (AU). Number of animals=4-7. *p<0.05, **p<0.01 vs the respective basal NO release from control rats; #p<0.05 vs the respective basal NO release from orchidectomized rats (from Blanco-Rivero et al., 2005).

These results indicate that PKC is involved in nNOS activation in arteries from both control and orchidectomized rats. Because the degree of PKC activation is already greater in arteries from orchidectomized rats, PDBu and calphostin C showed less ability to respectively increase or diminish NO release in arteries from orchidectomized than from control rats. Conventional and novel PKC isoforms are dependent on dyacylglycerol (DAG) (Ward et al., 2004) and, since we use the pharmacological mimetic of DAG, PDBu, we stimulate both isoforms. Although selective inhibitors for these isoforms do not exist (Davies et al., 2000) it has been reported that certain compounds, such as the indolocarbazole Gö6976, are partially selective for conventional over the novel and atypical isoforms (Martiny-Baron et al., 1993; Ward et al., 2004). Consequently, we tested the effect of this inhibitor on nNOS activity, and observed that a concentration of 0.1 µM Gö6976 inhibited the EFS-induced neuronal NO release in arteries from control arteries while it did not affect the neuronal NO release in arteries from orchidectomized rats. A higher Gö6976 concentration (1 µM) was used but it only decreased the EFS-induced NO release in arteries from orchidectomized rats. All these results support the assumption that nNOS seems to be much more activated by PKC, and probably the conventional PKC isoforms, in arteries from the orchidectomized than from the control rats.

Since the atypical PKC ζ isoform has been described in vascular smooth muscle as modulating vascular responses to different agents (Cogolludo *et al.*, 2003; De Witt *et al.*, 2001), the effect of this isoform on nNOS activity was analyzed. We found that PKC ζ -PI decreased EFS-induced NO release to a similar extent in arteries from control and orchidectomized animals. This result indicates that PKC ζ isoform involvement in the regulation of neuronal NO release occurs and that this involvement is not modulated by endogenous male sex hormones.

Regarding endothelial NO release, it has been previously described that orchidectomy did not modify the Ach-stimulated endothelial NO release, despite the increased production of superoxide anion. Based on the results obtained about PKC regulation of nNOS activity, the action of PKC activation or inhibition on eNOS activity was also investigated in arteries from orchidectomized and control animals. The results obtained showed that neither the PKC activator, PDBu (Nishizuka, 1984), nor the PKC inhibitor, calphostin C (Kobayashi *et al.* 1989), modified ACh-induced NO release in arteries from control animals. In contrast, in arteries from orchidectomized rats, PKC activation increased the basal and the AChinduced NO release while PKC inhibition more strongly decreased both basal and AChinduced NO release (Fig. 8).

105



Fig. 8. Effect of PDBu, calphostin, Gö-6976 or PKCζ-PI on the basal - (white columns) and Ach - (dashed columns) induced NO release in mesenteric arteries from control and orchidectomized rats. Results are expressed as arbitrary units (AU). Number of animals=8. *p<0.05 vs the respective basal NO release; #p<0.05 vs basal NO release in non treated arteries (from Blanco-Rivero et al., 2007).

These results indicate that PKC participates in eNOS activity only in arteries from orchidectomized rats, and also seem to indicate that PKC apparently did not regulate eNOS activity in arteries from control animals, which contrast with the results reported with nNOS (Blanco-Rivero et al., 2005) in which we observed an nNOS activity modulation by PKC. In this respect, it is possible to speculate that very subtle differences could exist in the modulation of NOS isoforms. In this regard, it is important to keep in mind that regulatory mechanisms other than PKC could be working on eNOS, including different redox conditions (Polytarchou & Papadimitriou, 2005), phosphatases activity (Fleming et al., 2001) and/or other kinases (Ferrer et al., 2004) that, in their turn, regulate the intracellular environment and function. Additionally, since the pharmacological mimic of dyacylglycerol, PDBu, stimulate conventional and novel PKC isoforms (Ward et al., 2004) and since calphostin C is a non specific PKC inhibitor, we tested the effect of the PKC inhibitor Gö6976, which is partially selective for conventional over novel and atypical PKC isoforms (Martiny-Baron et al., 1993, Ward et al., 2004). We observed that this inhibitor also decreased basal and ACh-induced NO in arteries from orchidectomized rats, indicating the involvement of the conventional PKC isoforms in endothelial NO regulation. Since the

atypical PKCζ isoform has been reported to modulate vascular responses (Damron *et al.*, 1998, De Witt *et al.*, 2001, Cogolludo *et al.*, 2003) and neuronal NO release (Blanco-Rivero *et al.*, 2005), we also tested the possible involvement of this isoform in endothelial NO release. The fact that PKCζ-PI decreased the basal and ACh-induced NO release showed the participation of this isoform. Moreover, since the three PKC inhibitors that we used, calphostin C, Gö6976 and PKCζ-PI, diminished both basal and ACh-induced NO release, it seems that eNOS, like nNOS (Blanco-Rivero *et al.*, 2005), would have already been activated by PKC in arteries from orchidectomized rats.

These results show that PKC activity is enhanced in mesenteric arteries from orchidectomized rats, and this increase would be responsible for the higher nNOS and eNOS activity.

6. Conclusions

Orchidectomy alters different cell signalling pathways that are involved in vascular tone regulation. Orchidectomy increases: (i) the formation of superoxide anion and peroxynitrite; (ii) the activity of PKC; and (iii) the expression of COX-2, the production of prostanoids derived from COX-2, as well as their vasoconstrictor effect. These aspects seem to be physiologically relevant, since the balance between vasodilator/vasoconstrictor prostanoids is lost in favour of vasoconstrictor substances in arteries from orchidectomized rats. This situation could indicate a disadvantage in cardiovascular function in the absence of male sex hormones, thereby suggesting that testosterone has a beneficial influence on the vasculature. However, in the animals used in our study (6 months old) several compensatory mechanisms are working: reactive oxygen species are able to induce relaxation; PKC positively regulates nNOS and eNOS activity ensuring the maintenance of NO release; the activity and expression of SOD are increased in an attempt to compensate for the increased superoxide anion production. This intriguing information makes it essential to perform studies in vascular function taking into account different cell signalling pathways that are working simultaneously. Future studies in the research field of androgens on cell signaling pathways are needed, since they will be of important interest to implement therapeutic strategies that could improve vascular function.

7. Acknowledgements

This work was supported by grant from Fondo de Investigaciones Sanitarias (PI08831).

8. References

- Abdel-Latif A.A. (1986). Calcium mobilizing receptors, polyphosphoinositides, and the generation of second messengers. *Pharmacol Rev* 38: 227-272.
- Agostini M.C., Borda E.S., Gimeno M.F. *et al.* (1981). Differences in the effects of acetylcholine on the vas deferens from normal and castrated rats. A participation of adrenergic mechanisms. *Arch Int Pharmacodyn Ther* 250: 212-220.
- Axelband F., Dias J., Ferrão F.M. *et al.* (2011). M. Nongenomic signaling pathways triggered by thyroid hormones and their metabolite 3-iodothyronamine on the cardiovascular system. *J Cell Physiol* 226:21-28.

- Ba Z.F., Wang P., Koo D.J. *et al.* (2001). Attenuation of vascular endothelial dysfunction by testosterone receptor blockade after trauma and hemorrhagic shock. *Archives of Surgery* 136: 1158-63.
- Balafanova Z., Bolli R., Zhang J. *et al.* (2002). Nitric oxide (NO) induces nitration of protein kinase Cepsilon (PKCepsilon), facilitating PKCepsilon translocation via enhanced PKCepsilon -RACK2 interactions: a novel mechanism of no-triggered activation of PKCepsilon. *J Biol Chem* 277: 15021-7.
- Bain J. (2020). Testosterone and the aging male: to treat or not to treat? Maturitas 66: 16-22.
- Baker P.J., Ramey E.R. & Ramwell P.W. (1978). Androgen-mediated sex differences of cardiovascular responses in rats. *Am J Physiol* 235: H242-246.
- Bapat S., Verkleij A. & Post J.A. (2001). Peroxynitrite activates mitogen-activated protein kinase (MAPK) via MEK-independent pathway: a role for protein kinase C. *FEBS Lett* 499: 21-6.
- Barud W. Palusinski R., Beltkowski J. *et al.* (2002). Inverse relationship between total testosterone and anti-oxidized low-density lipoprotein antibody levels in ageing males. *Atherosclerosis* 164:282-288.
- Basaria S. & Dobs A.S. (2001) Hypogonadism and androgen replacement therapy in elderly men. *Am J Med* 110: 563-572.
- Bachschmid M., Schildknecht S. & Ullrich V. (2005). Redox regulation of vascular prostanoid synthesis by the nitric oxide-superoxide system. *Biochem Biophys Res Commun* 338:536-542.
- Békési G., Kakucs R., Várvíró S. *et al.* (2000). In vitro effects of different steroid hormones on superoxide anion production of human neutrophil granulocytes. *Steroids* 65: 889-894.
- Blanco-Rivero J., Balfagón G. & Ferrer M. (2005). Male castration increases neuronal nitric oxide synthase activity in the rat mesenteric artery through protein kinase C activation. *Journal of Vascular Research* 42: 526-534.
- Blanco-Rivero J., Balfagón G. & Ferrer M. (2005). Orchidectomy modulates α2-adrenoceptor reactivity in rat mesenteric artery through increased thromboxane A₂ formation. *Journal of Vascular Research* 43: 101-108.
- Blanco-Rivero J., Cachofeiro V., Lahera V. *et al.* (2005). Participation of prostacyclin in endothelial dysfunction induced by aldosterone in normotensive and hypertensive rats. *Hipertension* 46:107-12.
- Blanco-Rivero J., Sagredo A., Balfagón G. & Ferrer M. (2006). Orchidectomy increases expression and activity of Cu/Zn-superoxide dismutase, while decreases endothelial nitric oxide bioavailability. *J Endocrinol* 190: 771-778.
- Blanco-Rivero J., Sagredo A., Balfagón G. & Ferrer M. (2007). Protein kinase C activation increases endothelial nitric oxide release in mesenteric arteries from orchidectomized rats. *J Endocrinol* 192: 189-197.
- Blanco-Rivero J., Márquez-Rodas I., Xavier F. *et al.* (2007). Long-term fenofibrate treatment impairs endothelium-dependent dilation to acetylcholine by altering cyclooxygenase pathway. *Cardiovas Res* 75:398-340.
- Bolotina V.M., Najibi S., Palacino J.J. *et al.* (1994). Nitric oxide directly activates calciumdependent potassium channels in vascular smooth muscle. *Nature* 368: 850-853.

- Braga-Basaria M., Dobs A.S., Muller D.C. *et al.* (2006). Metabolic syndrome in men with prostate cancer undergoing long-term androgen-deprivation therapy. *J Clin Oncol* 24: 3979-3983.
- Bredt D.S., Ferris C.D. & Snyder S.H. (1992). Nitric oxide synthase regulatory sites. Phosphorylation by cyclic AMP-dependent protein kinase, protein kinase C and calcium/calmodulin protein kinase; identification of flavin and calmodulin binding sites. *J Biol Chem* 267: 10976-10981.
- Calderone V., Baragatti B., Breschi M.C. *et al.* (2002). Hormonal influence on the release of endothelial nitric oxide: gender related dimorphic sensitivity of rat aorta for noradrenaline. *J Pharm Pharmacol* 54: 523-528.
- Caughey G.E., Cleland L.G., Gamble J.R. & James M.J. (2001) Up-regulation of endothelial cyclooxygenase-2 and prostanoid synthesis by platelets. Role of thromboxane A2. *J Biol Chem* 276: 37839-37845.
- Ceballos G., Figueroa L., Rubio I. *et al.* (1999). Acute and nongenomic effect of testosterone on isolate and perfused rat heart. *J Cardiovasc Pharmacol* 33: 691-697.
- Chatrath R., Ronningen K.L., Severson S.R. *et al.* (2003). Endothelium-dependent responses in coronary arteries are changed with puberty in male pigs. *Am J Physiol* 285: H1168-1176.
- Chen D.C., Duckles S.P. & Krause D.N. (1999). Postjunctional α2-adrenoceptors in the rat tail artery: effect of sex and castration. *Eur J Pharmacol* 372: 247-252.
- Chen G.X. (2003). Selective protein kinase C inhibitors and their applications. *Curr Drug Targets Cardiovasc Haematol Disord* 3: 301-307.
- Cheng Y., Austin S.C., Rocca B. *et al.* (2002). Role of prostacyclin in the cardiovascular response to thromboxane A2. Science 296 539-541.
- Chodak G.W., Keane T. & Klotz L. (2002) Critical evaluation of hormonal therapy for carcinoma of the prostate. *Urology* 60: 201-208.
- Cogolludo A., Moreno L., Bosca L. *et al.* (2003). Thromboxane α₂-induced inhibition of voltage-gated K+ channels and pulmonary vasoconstriction: role of protein kinase Czeta. *Circ Res* 93: 656-63.
- Damron D.S., Nadim H.S., Hong S.J. *et al.* (1998). Intracellular translocation of PKC isoforms in canine pulmonary artery smooth muscle cells by ANG II. *Am J Physiol* 274: L278-288.
- Davies S.P., Reddy H., Caivano M. & Cohen P. (2000). Specificity and mechanism of action of some commonly used protein kinase inhibitors. *Biochem J* 351:95-105.
- Dawson T.M., Steiner J.P., Dawson V.L. *et al.* (1993). Inmunosuppressant FK506 enhances phosphorylation of nitric oxide synthase and protects against glutamate neurotoxicity. *Proc Natl Acad Sci* 90: 9808-9812.
- De Witt B.J., Kaye A., Ibrahim I.N. *et al.* (2001). Effects of PKC isozyme inhibitors on constrictor responses in the feline pulmonary vascular bed. *Am J Physiol* 280: L50-57.
- Duckles S.P. & Miller V.M. (2010). Hormonal modulation of endothelial NO production. *Eur J Physiol* 459: 841-851.
- Félétou M., Köhler R. & Vanhoutte P.M. (2010). Endothelium-derived vasoactive factors and hypertension: possible roles in pathogenesis and as treatment targets. *Curr Hypertens Rep* 12: 267-275.

- Félétou M. & Vanhoutte P.M. (2006). Endothelial dysfunction: a multifaceted disorder. *Am J Physiol Heart Circ Physiol* 291: 985-1002.
- Ferrer M., Alonso M.J., Salaices M. *et al.* (2000). Increase in neurogenic nitric oxide metabolism by endothelin-1 in mesenteric arteries from hypertensive rats. *J Cardiovasc Pharmacol* 36: 541-547.
- Ferrer M., Alonso M.J., Salaices M. *et al.* (2001). Angiotensin II increases neurogenic nitric oxide metabolism in mesenteric arteries from hypertensive rats. *Life Sci* 68: 1169-1179.
- Ferrer M., Encabo A., Conde M.V. et al. (1995). Heterogeneity of endothelium-dependent mechanisms in different rabbit arteries. *J Vasc Res* 32: 339-46.
- Ferrer M., Marín J. & Balfagón G. (2000). Diabetes alters neuronal nitric oxide release from rat mesenteric arteries. Role of protein kinase C. *Life Sci.* 66:337-345.
- Ferrer M., Tejera N., Marín J. *et al.* (1999). Androgen deprivation facilitates acetylcholineinduced relaxation by superoxide anion generation. *Clin Sci* 140: 1861-1868.
- Ferrer M., Salaices M. & Balfagón G. (2004). Endogenous prostacyclin increases neuronal nitric oxide release in mesenteric artery from spontaneously hypertensive rats. *Eur J Pharmacol* 506: 151-156.
- FitzGerald G.A. (1991). Mechanisms of platelet activation: thromboxane A2 as an amplifying signal for other agonists. *Am J Cardiol* 68: 11-15.
- FitzGerald G.A., Healy C. & Daugherty J. (1987). Thromboxane A2 biosynthesis in human disease. *Fed Proc* 46:154-8.
- Fleming I., Fisslthaler B., Dimmeler S. *et al.* (2001). Phosphorylation of Thr(495) regulates Ca(2+)/calmodulin-dependent endothelial nitric oxide synthase activity. *Circ Res* 88: E68-75.
- Förstermann U., Pollock J.S., Schmint H.H.H.W. *et al.* (1991). Calmodulin-dependent endothelium-derivaded relaxing factor/nitric oxide synthase activity is present in the particulate and cytosolic fractions of bovine aortic endothelium cells. *Proc Nat Acad Sci* 88: 1788-1792.
- Frein D., Schildknecht S., Bachschmid M. *et al.* (2005). Redox regulation: a new challenge for pharmacology. *Biochem Pharmacol* 70:811-823.
- Furchgott R.F. & Zawadzki J.V. (1980). The obligatory role of endothelial cells in the relaxation of arterial smooth muscle by acetylcholine. *Nature* 288: 373-376.
- Gluais P., Lonchampt M., Morrow J.D. *et al.* (2005). Acetylcholine-induced endotheliumdependent contractions in the SHR aorta: the Janus face of prostacyclin. *Br J Pharmacol* 146: 834-45.
- Gonzales R.J., Ghaffari A.A., Duckles S.P. *et al.* (2005). Testosterone treatment increases thromboxane function in rat cerebral arteries. *Am J Physiol Heart Circ Physiol* 289: 578-85.
- Gonzales R.J., Krause D.N. & Duckles S.P. (2004). Testosterone suppresses endotheliumdependent dilation of rat middle cerebral arteries. *Am J Physiol* 286: H552-H560.
- Greenberg S., George W.R., Kadowitz P.J. *et al.* (1974). Androgen-induced enhancement of vascular reactivity. *Can J Physiol Pharmacol* 52: 14-22.
- Gryglewski R.J., Palmer R.M.J. & Moncada S. (1986). Superoxide anion is involved in the breakdown of endothelium-derived vascular relaxing factor. *Nature* 320: 454-456.
- Guan X.B. & Dluzen D. (1991). Castration reduces potassium-stimulated norepinephrine release from superfused olfactory bulbs of male rats. *Brain Res* 568: 147-151.

Harrison D.G. (1994). Endothelial dysfunction in atherosclerosis. Basic Res Cardiol 89: 87-102.

- Henrion D., Dechaux E., Dowell F.J. *et al.* (1994). Alteration of flow-induced dilatation in mesenteric resistance arteries of L-NAME treated rats and its partial association with induction of cyclo-oxygenase-2. *Br J Pharmacol* 121:83-90
- Holmquist F., Persson K., Bodker A. *et al.* (1994). Some pre- and postjunctional effects of castration in rabbit isolated corpus cavernosum and urethra. *J Urol* 152: 1011-1016.
- Hutchison S.J., Sudhir K., Chou T.M. *et al.* (1997). Testosterone worsens endothelial dysfunction associated with hypercholesterolemia and environmental tobacco smoke exposure in male rabbit aorta. *J Am Coll Cardiol* 29: 800-807.
- Ishikawa M. & Quock R.M. (2003). Role of nitric-oxide synthase isoforms in nitrous oxide antinociception in mice. *J Pharmacol Exp Ther* 306: 484-489.
- Jones R.D., Jones H.T. & Channer K.S. (2004). The influence of testosterone upon vascular reactivity. *Eur J Endocrinol* 151: 29-37.
- Jones T.H. (2010). Testosterone deficiency: a risk factor for cardiovascular disease? *Trends Endocrinol Metab* 21: 496-503.
- Jones T.H. & Saad F. (2009). The effects of testorene on risk factors for, and the mediators of, the atherosclerotic process. *Atherosclerosis* 207: 318-327.
- Kapoor D., Malkin C.J. Channer K.S. *et al.* (2005). Androgens, insulin resistance and vascular disease in men. *Clin Endocrinol* 63: 239-250.
- Kanashiro C.A. & Khalil R.A. (2001). Gender-related distinctions in protein kinase C activity in rat vascular smooth muscle. *Am J Physiol* 280: C34-C45.
- Kawasaki H., Takasaki K., Saito A. *et al.* (1988). Calcitonin gene-related peptide acts as a novel vasodilator neurotransmitter in mesenteric resistance vessels of the rat. *Nature* 335: 164-167.
- Keating N.L., O'Malley A.J. & Smith M.R. (2006). Diabetes and cardiovascular disease during androgen deprivation therapy for prostate cancer. *J Clin Oncol* 24: 4448-4456.
- Khalil R.A. & van Breemen C. (1988). Sustained contraction of vascular smooth muscle: calcium influx or C-kinase activation?. *J Pharmacol Exp Ther* 244: 537-542.
- Kim E.J., Raval A.P. & Perez-Pinzon M.A. (2008). Preconditioning mediated by sublethal oxygen-glucose deprivation-induced cyclooxygenase-2 expression via the signal transducers and activators of transcription 3 phosphorylation. J Cereb Blood Flow Metab 28: 1329-1340.
- Klein T., Eltze M., Grebe T. *et al.* (2007). Celecoxib dilates guinea-pig coronaries and aortic rings and amplifies NO/cGMP signaling by PDE5 inhibition. *Cardiovasc Res* 75: 390-397.
- Kobayashi E., Nakano H., Morimoto & Tamaoki T. (1989). Calphostin C (UCN-1028C), a novel microbial compound is a highly potent and specific inhibitor of protein kinase C. *Biochem Biophys Res Commun* 159: 548-553.
- Kobayashi S., Inoue N., Azumi H. *et al.* (2002). Expressional change of the vascular antioxidant system in atherosclerotic coronary arteries. *Arterioscler Thromb Vasc Biol* 9: 184-190.
- Li Y.J. Duckles& S.P. (1992). Effect of endothelium on the actions of sympathetic and sensory nerves in the perfused rat mesentery. *Eur J Pharmacol* 210: 23-40.
- Lincoln T.M., Dey N. & Sellak H. (2001). cGMP-dependent protein kinase signaling mechanisms in smooth muscle: from the regulation of tone to gene expression. *J Appl Physiol* 91: 1421-1430.

- Liu D. & Dillon J.S. (2002). Dehydroepiandrosterone activates endothelial cell nitric-oxide synthase by a specific plasma membrane receptor coupled to G alpha (i2,3), *J Biol Chem* 277 :21379-21388.
- Martín M.C., Balfagón G., Minoves N. & Ferrer M. (2005). Androgen deprivation increases neuronal nitric oxide metabolism and its vasodilator effect in rat mesenteric arteries. *Nitric Oxide: Biology and Chemistry* 12: 163-176.
- Martiny-Baron G., Kazanietz M.G., Mischak H. *et al.* (1993). Selective inhibition of protein kinase C isozymes by the indolocarbazole Gö6976. *J Biol Chem* 268:9194-7.
- McNeill A.M., Kim N., Duckles S.P. *et al.* (1999). Chronic estrogen treatment increases levels of endothelial nitric oxide synthase protein in rat cerebral microvessels. *Stroke* 30: 2186-2190.
- Minoves N., Balfagón G. & Ferrer M. (2002). Role of female sex hormones in neuronal nitric oxide release and metabolism in rat mesenteric arteries. *Clin Sci* 103: 239-247.
- Mukherjee S., Coaxum S.D., Maleque M. *et al.* (2001). Effects of oxidized low density lipoprotein on nitric oxide synthetase and protein kinase C activities in bovine endothelial cells. *Cell Mol Biol* 47: 1051-1058.
- Munzel T., Heitzer T. & Harrison D.G. (1997). The physiology and pathophysiology of the nitric oxide/superoxide system. *Herz* 22:158-172.
- Murad F. (1997). What are the molecular mechanisms for the antiproliferative effects of nitric oxide and cGMP in vascular smooth muscle? *Circulation* 95: 1101-1103.
- Muzykantov V.R. (2001). Targeting of superoxide dismutase and catalase to vascular endothelium. *J Controlled Release* 71: 1-21.
- Miyamoto A., Hashiguchi. Y, Obi T. *et al.* (2007). Ibuprofen or ozagrel increases NO release and l-nitro arginine induces TXA(2) release from cultured porcine basilar arterial endothelial cells. *Vasc Pharmacol* 46. 85-90.
- Nakane M., Mitchell J., Forstermann U. & Murad F. (1991). Phosphorylation by calcium calmodulin-dependent protein kinase II and protein kinase C modulates the activity of nitric oxide synthase. *Biochem Biophys Res Commun* 180: 1396-1402.
- Namgaladze D., Shcherbyna I., Kienhofer J. *et al.* (2005). Superoxide targets calcineurin signaling in vascular endothelium. *Biochem BiophysRes Commun* 334: 1061-1067.
- Narumiya S., Sugimoto Y. & Ushikubi F. (1999). Prostanoid receptors: structures, properties, and functions. *Physiol Rev* 79:1193-226.
- Newton AC. (1995). Protein kinase C: structure, function and regulation. J Biol Chem 270: 28495-28498.
- Nielsen K.C. & Owman C. (1971). Contractile response and amine receptor mechanism in isolated middle cerebral artery of the cat. *Brain Res* 27: 25-32.
- Nishihara M., Yokotani K., Inoue S. & Osumi Y. (2000). U-46619, a selective thromboxane A2 mimetic, inhibits the release of endogenous noradrenaline from the rat hippocampus in vitro. *Jpn J Pharmacol* 82: 226-231.
- Nishizuka Y. (1992). Intracellular signaling by hydrolysis of phospholipids and activation of PKC. *Science* 258: 607-614.
- Nishizuka Y. (1984). The role of protein kinase C in cell surface signal transduction and tumour promotion. *Nature* 308: 693-698.
- Nguyen Dinh Cat A & Touyz RM. (2011) Cell Signaling of Angiotensin II on Vascular Tone: Novel Mechanisms. *Curr Hypertens Rep* 13:122-128.

- Noll G. & Luscher T.F. (1998). The endothelium in acute coronary syndromes. *Eur Heart J* 19: C30-C38.
- Oeckler R.A. & Wolin M.S. (2000). New concepts in vascular nitric oxide signalling. *Curr Atheroscler* 2: 437-444.
- Okada D. (1992). Two pathways of cyclic GMP production through glutamate receptormediated nitric oxide synthesis. *J Neurochem* 59: 1203-1210.
- Okada D. (1995). Protein kinase C modulates calcium sensitivity of nitric oxide synthase in cerebellar slices. *J Neurochem* 64: 1298-1304.
- Onoue S., Endo K., Yajima T. *et al.* (2002). Pituitary adenylate cyclase activating polypeptide regulates the basal production of nitric oxide in PC12 cells. *Life Sci* 71: 205-214.
- Orshal J.M. & Khalil R.A. (2004). Gender, sex hormones and vascular tone. *Am J Physiol* 286: R233-R249.
- Oury T.D., Day B.J. & Crapo J.D. (1996). Extracellular superoxide dismutase: a regulator of nitric oxide bioavailability. *Laboratory Investigation* 75: 617-36.
- Polytarchou C. & Papadimitriou E. (2005). Antioxidans inhibit human endothelial cell functions through down-regulation of endothelial nitric oxide synthase activity. *Eur J Pharmacol* 510: 1-38.
- Price D.T., Vita J.A. & Keaney J.F.Jr. (2000). Redox control of vascular nitric oxide bioavailability. *Antioxidants & Redox Signaling* 2: 919-935.
- Rapoport R.M. & Williams S.P. (1996). Role of prostaglandins in acetylcholine-induced contraction of aorta from spontaneously hypertensive and Wistar-Kyoto rats. *Hypertension* 28: 64-75.
- Reynoso R., Mohn C., Retory V. *et al.* (2002). Changes in the effect of testosterone on hypothalamic nitric oxide synthetase during sexual maturation. Its relationship with GnRH release. *Neuroendocrinol Lett* 23: 101-4.
- Rubanyi G.M. & Vanhoutte P.M. (1986). Oxygen-derived free radicals, endothelium, and responsiveness of vascular smooth muscle. *Am J Physiol* 120: H815-H821.
- Rump L.C. & Schollmeyer P. (1989). Effects of endogenous and synthetic prostanoids, the thromboxane A2 receptor agonist U-46619 and arachidonic acid on [3H]noradrenaline release and vascular tone in rat isolated kidney. Br J Pharmacol 97: 819-828.
- Saad F., Gooren L.J., Haider A. *et al.* (2008). A dose-response study of testosterone on sexual dysfunction and features of the metabolic syndrome using testosterone gel and parenteral testosterone undecanoate. *J Androl* 29: 102-105.
- Scordalakes E.M., Imwalle D.B. & Rissman E.F. (2002). Oestrogen's masculine side: medition of mating in male mice. *Reproduction* 124: 331-338.
- Shang Y. & Dluzen D.E. (2002). Castration increases nisoxetine-evoked norepinephrine levels in vivo within the olfactory bulb of male rats. *Neurosci Lett* 328: 81-84.
- Shanmugam N., Gaw-Gonzalo I.T. & Natarajam R. (2004). Molecular mechanisms of high glucose-induced cyclooxygenase-2 expression in monocytes. *Diabetes* 53: 795-802.
- Siddiqui A. & Shah B.H. (1997). Neonatal androgen manipulation differentially affects the development of monoamine systems in rat cerebral cortex, amygdale and hypothalamus. *Dev Brain Res* 98: 247-252.
- Simon D., Charles M.A., Nahoul K. *et al.*, (1997). Association between plasma testosterone and cardiovascular risk factors in healthy adult men: the telecom study. *J Clin Endocrinol Metab* 82: 682-689.

- Simoncini T., Mannella P., Fornari L. *et al.* (2003). Dehydroepiandrosterone modulates endotelial nitric oxide synthesis via direct genomic and nongenomic mechanisms. *Endocrinology* 144: 3449-55.
- Singh .R, Pervin S., Shryne J. *et al.* (2000). Castration increases and androgens decrease nitric oxide synthase activity in the brain: physiologic implications. *Proc Natl Acad Sci* 97: 3672-7.
- Smith M.R., Finkelstein J.S., McGovern F.J. *et al.* (2002). Changes in body composition during androgen deprivation therapy for prostate cancer. *J Clin Endocrinol Metab* 87: 599-603.
- Spitaler M. & Cantrell D.A. (2004). Protein kinase C and beyond. Nature Immunol 5:785-790.
- Strehlow K., Rotter S., Wassmann S. *et al.* (2003). Modulation of antioxidant enzyme expression and function by estrogen. *Circ Res* 93: 170-177.
- Sun C., Chen M., Mao J. *et al.* (2001). Biphasic effects of orchidectomy on calcitonin generelated peptide synthesis and release. *Neuroreport* 12: 3497-3502.
- Szmydynger-Chodobska J., Zink B.J. & Chodobski A. (2011) Multiple sites of vasopressin synthesis in the injured brain. *J Cereb Blood Flow Metab* 31:47-51.
- Tanaka M., Umemoto S., Kawahara S. *et al.* (2005). Angiotensin II type 1 receptor antagonist and angiotensin-converting enzyme inhibitor altered the activation of Cu/Zncontaining superoxide dismutase in the heart of stroke-prone spontaneously hypertensive rats. *Hypertens Res* 28: 67-77.
- Teoh H., Quan A. & Man R.Y. (2000). Acute impairment of relaxation by low levels of testosterone in porcine coronary arteries. *CardiovascRes* 45: 1010-1018.
- Tep-areenan P., Kendall D.A. & Randal M.D. (2003). Mechanisms of vasorelaxation to testosterone in the rat aorta. *Eur J Pharmacol* 465: 125-132.
- Tesauro M., Schinzari F., Caramanti M. *et al.* (2010). Cardiovascular and metabolic effects of ghrelin. *Curr Diabetes Rev* 6:228-35.
- Toda N. & Okamura T. (2003). The pharmacology of nitric oxide in the peripheral nervous system of blood vessels. *Pharmacol Rev* 55:271-324.
- Tracey W.R., Nakane M., Basha F. *et al.* (1995). In vivo pharmacological evaluation of two novel type II (inducible) nitric oxide synthase inhibitors. *Can J Physiol Pharmacol* 73: 665-669.
- Traish A.M. & Kypreos K.E. (2011). Testosterone and cardiovascular disease: an old idea with modern clinical implications. *Atherosclerosis* 214:244-248.
- Vanhoutte P.M. (1996). Endothelium-dependent responses in congestive heart failure. J Mol Cell Cardiol 28: 2233-2240.
- Villablanca A.C., Jayachandran M. & Banka C. (2010). Atherosclerosis and sex hormones: current concepts. *Clin Sci* 119:493-513.
- Ward J.P.T., Knock G.A., Snetkov V.A. & Aaronson P.I. (2004). Protein kinases in vascular smooth muscle tone -role in the pulmonary vasculature and hypoxic pulmonary vasoconstriction. *Pharmacol Ther* 104: 207-231.
- Wei E.P., Kontos H.A. & Beckman J.S. (1996). Mechanisms of cerebral vasodilation by superoxide, hydrogen peroxide, and peroxynitrite. *Am J. Physiol* 271: H1262-H1266.
- Weiner I., Lizasoain S.A., Baylis R.G. *et al.* (1994). Induction of calcium-dependent nitric oxide synthases by sex hormones. *Proc Natl Acad Sci* 91: 5212-5216.
- Wolin M.S. (2002). Interaction of oxidants with vascular signaling system. *Arterioscler Thromb Vasc Biol* 20: 1430-1442.

- Wynne F.L. & Khalil R.A. (2003). Testosterone and coronary vascular tone: implications in coronary artery disease. *J Endocrinol Invest* 26: 181-186.
- Yamada T., Fujino T., Yuhki K. *et al.* (2003). Thromboxane A2 regulates vascular tone via its inhibitory effect on the expression of inducible nitric oxide synthase. *Circulation* 108: 2381-2386.
- Yorek M.A., Coppey L.J., Gellett J.S. *et al.* (2002). Effect of treatment of diabetic rats with dehydroepiandrosterone on vascular and neural function. *Am J Physiol* 283: 1067-1075.





Basic and Clinical Endocrinology Up-to-Date

Edited by Dr. Fulya Akin

ISBN 978-953-307-340-8 Hard cover, 350 pages **Publisher** InTech **Published online** 17, October, 2011 **Published in print edition** October, 2011

This book provides the most up-to-date information on the basic and clinical aspects of endocrinology. It offers both researchers and clinicians experts, gold-standard analysis of endocrine research and translation into the treatment of diseases such as insulinoma, endocrine disease in pregnancy and steroid induced osteoporosis. Investigates both the endocrine functions of the kidneys and how the kidney acts as a target for hormones from other organ systems. Presents a uniquely comprehensive look at all aspects of endocrine changes in pregnancy and cardiovascular effects of androgens.

How to reference

In order to correctly reference this scholarly work, feel free to copy and paste the following:

Mercedes Ferrer (2011). Androgens and Vascular Function, Basic and Clinical Endocrinology Up-to-Date, Dr. Fulya Akin (Ed.), ISBN: 978-953-307-340-8, InTech, Available from: http://www.intechopen.com/books/basic-and-clinical-endocrinology-up-to-date/androgens-and-vascular-function

Open science | open minds

InTech Europe

University Campus STeP Ri Slavka Krautzeka 83/A 51000 Rijeka, Croatia Phone: +385 (51) 770 447 Fax: +385 (51) 686 166 www.intechopen.com

InTech China

Unit 405, Office Block, Hotel Equatorial Shanghai No.65, Yan An Road (West), Shanghai, 200040, China 中国上海市延安西路65号上海国际贵都大饭店办公楼405单元 Phone: +86-21-62489820 Fax: +86-21-62489821 © 2011 The Author(s). Licensee IntechOpen. This is an open access article distributed under the terms of the <u>Creative Commons Attribution 3.0</u> <u>License</u>, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

IntechOpen

IntechOpen