

We are IntechOpen, the world's leading publisher of Open Access books Built by scientists, for scientists

6,900

Open access books available

186,000

International authors and editors

200M

Downloads

Our authors are among the

154

Countries delivered to

TOP 1%

most cited scientists

12.2%

Contributors from top 500 universities



WEB OF SCIENCE™

Selection of our books indexed in the Book Citation Index
in Web of Science™ Core Collection (BKCI)

Interested in publishing with us?
Contact book.department@intechopen.com

Numbers displayed above are based on latest data collected.
For more information visit www.intechopen.com



Biomarkers of Aflatoxin Exposure and Its Relationship with the Hepatocellular Carcinoma

Alessandra Vincenzi Jager, Fernando Silva Ramalho,
Leandra Náira Zambelli and Carlos Augusto Fernandes Oliveira
*Universidade de São Paulo
Brazil*

1. Introduction

Mycotoxins are secondary metabolites produced by fungi that grow naturally in foodstuffs. They are able to generate a wide variety of toxic effects in vertebrates, including men (Coulombe, 1991). Toxigenic fungi may contaminate foodstuffs in the most different phases of production and processing, from cultivation to transport and storage. Mycotoxins show high chemical stability and may persist in the foodstuff even after fungi were removed by common manufacturing and packaging processes (Chu, 1991).

Diseases caused by mycotoxins are called mycotoxicoses. They are diffuse syndromes that cause lesions mainly in organs such as liver, kidneys, epithelial tissue (skin and mucous membranes) and central nervous system, depending on the type of the toxin. Two or more toxins may also occur simultaneously, leading to intensified toxic effects on the susceptible organism (Orsi et al., 2007).

Aflatoxins are mycotoxins produced by fungi in the genus *Aspergillus*, species *A. flavus*, *A. parasiticus* and *A. nomius* (Moss, 1998). These fungi are distributed worldwide, and their optimal growth conditions are relative humidity of 80-85% and temperature around 30°C (Coulombe, 1991).

Nowadays, 18 similar compounds are called aflatoxins. However, the most important in medical terms are types B₁, B₂, G₁ e G₂ (Coulombe, 1991). Aflatoxin B₁ (AFB₁), besides being the most frequently found in plant substrates, has the greatest toxigenic power. Aflatoxins B₂ (AFB₂), G₁ (AFG₁) and G₂ (AFG₂) have about 50, 20 and 10% of AFB₁ toxigenic power, respectively (Leeson et al., 1995).

AFB₁ is a genotoxic compound, and is considered to be one of the most potent natural mutagens. Liver carcinogenesis is the most important effect of chronic aflatoxin exposure. This toxicity has been widely demonstrated – mainly in relation to AFB₁ - in many animal species, including fish, birds, rodents, carnivores and primates (Busby & Wogan, 1984). Based on available studies, the *International Agency for Research on Cancer* (IARC) concluded, in 1987, that there was enough evidence to classify AFB₁ in Group 1 - human carcinogen (Rothschild, 1992).

One of the most important aspects in risk analysis of chemical substances is to determine the degree of human exposure (World Health Organization [WHO], 2002), a particularly difficult task for contaminants present in foodstuffs. However, it is possible to indirectly estimate the degree of exposure based on data on consumption of contaminated foodstuffs, and on the average occurrence of the toxin. In this estimation, the degree of exposure is

measured in terms of probable daily intake (PDI) per unit of body weight, and is generally expressed in ng/kg of body weight (BW) / day. In risk analysis, PDI is compared with tolerable daily intake (TDI) determined in toxicological studies. In spite of the genotoxic characteristic of this toxin, there is no consensus on tolerable daily intake of AFB₁.

Taking into account aflatoxin toxicity and the lack of an established TDI, several countries determined regulations on maximum aflatoxin levels allowed in foodstuffs. Table 1 summarizes some data of a report by the Food and Agriculture Organization of the United Nations (FAO, 2004). It may be noted that the European Community and the Mercosur standardized their regulations, although some countries kept some food items with additional country regulations. Foodstuffs characteristic of each country, frequency of consumption of these items and climate characteristics apparently influence maximum limits adopted in each region, although there is a consensus that these limits should comply with the ALARA (as lowest as reasonable accepted) criterion recommended by the FAO (2004).

2. AFB₁ biotransformation

Biotransformation is a process by which the body transforms foreign substances (xenobiotics) in new chemical compounds (metabolites), that is, a process in which the initial compound is modified to be eliminated by the biological system (Guengerich, 1999). After oral ingestion, AFB₁ is efficiently absorbed and biotransformed before urinary and fecal excretion (Figure 1).

Absorbed AFB₁ and its metabolites are excreted in urine and feces. Breastfeeding mothers who consume contaminated foodstuffs may also shed aflatoxins metabolites in their milk. Studies in animals demonstrated that in normal conditions, 50% of AFB₁ oral dose is quickly absorbed in the duodenum and reach the liver by the portal system (Wilson et al., 1985). AFB₁ is concentrated in the liver and, in lesser amounts, in the kidneys. It may also be found in mesenteric venous blood as free AFB₁ or as water-soluble metabolites (Wogan et al., 1967).

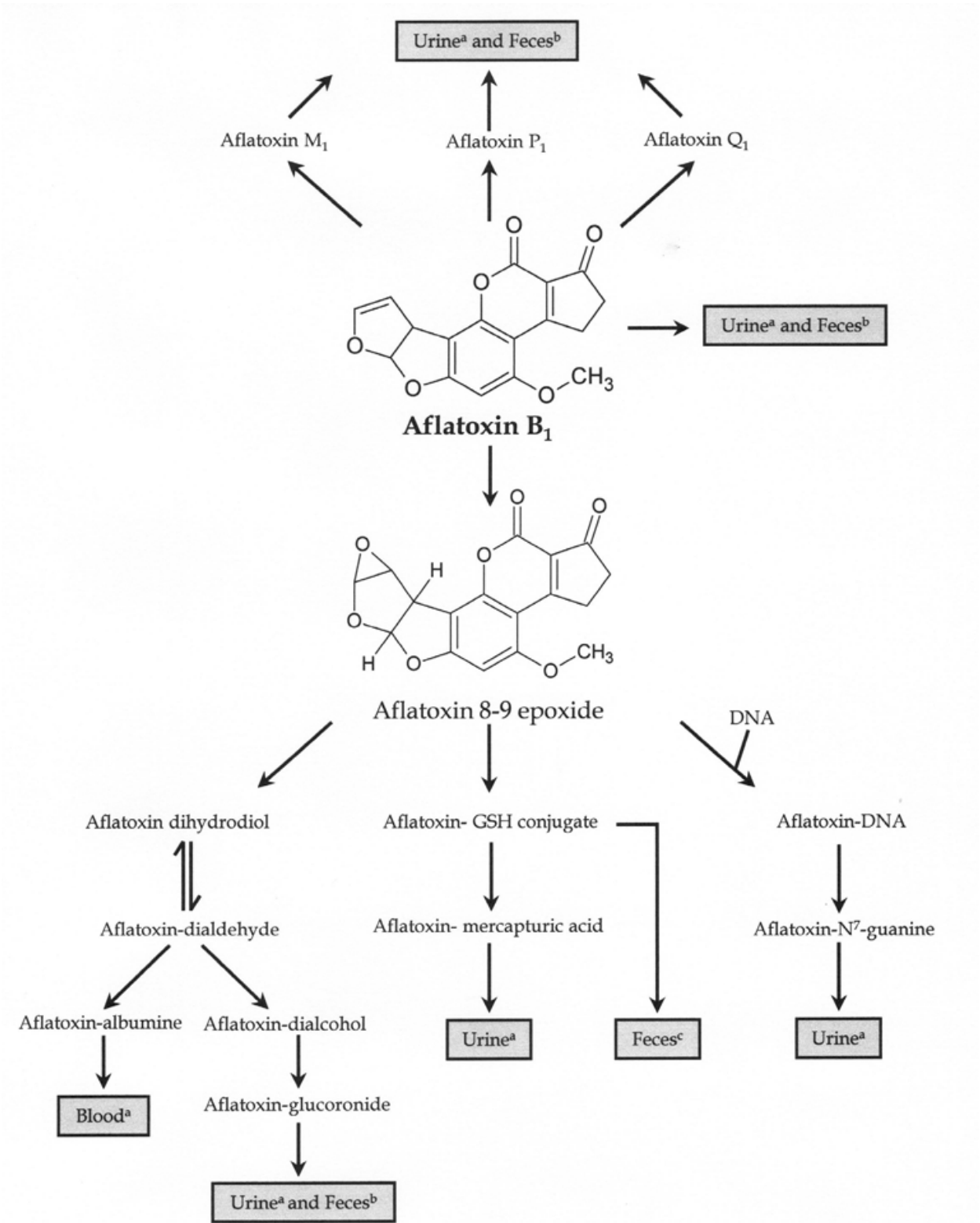
Enzymes of the cytochrome P450 (CYP) family, CYP1A2, CYP3A4 and CYP2A6, are responsible for the biotransformation of absorbed aflatoxins (Essigmann et al., 1982). These enzymes convert AFB₁ into its carcinogenic form, AFB-8,9-epoxide, which bonds covalently to DNA and serum albumin, producing AFB₁-N⁷-guanine and lysine adducts, respectively (Essigmann et al. 1977; Sabbioni et al. 1987). The bond between AFB₁ and DNA modifies the structure and biological activity of DNA, leading to the basic mutagenic and carcinogenic mechanisms of the toxin. Studies with rat livers showed that AFB₁-N⁷-guanine adducts may be removed after they are formed, leaving apurinic sites in the DNA molecule (Hsieh et al., 1991). Vacant sites tend to be filled with adenine, causing a guanine to thymine transversion and generating a highly significant point of mutation (Aguillar et al., 1993).

Besides being epoxidized, AFB₁ can be also oxidized into several other derivatives. The main hydroxylated metabolites are aflatoxin M₁ (AFM₁), aflatoxin Q₁ (AFQ₁), a demethylated metabolite, aflatoxin P₁ (AFP₁), and a reduced metabolite, aflatoxicol (Figure 1). AFM₁ may be activated to form AFM₁-8,9-epoxide, which binds to DNA and is excreted in urine as AFM₁-N⁷-guanine (Egner et al, 2003). AFQ₁ and AFP₁ are not significantly oxidized by human microsomes, and are not considered to be genotoxic (Raney et. al, 1992). Metabolites AFM₁, AFQ₁ and AFP₁ are not good substrates for epoxidation, are less genotoxic than AFB₁, and consequently, are considered detoxification products. However, because of the high toxicity reported for AFM₁, researchers should be cautious when labeling this compound a “detoxification product” (Neal et al., 1998).

Country	Food Product	Mycotoxin	Limit (µg/kg)
United States	All foods except milk	AFB ₁ , AFB ₂ , AFG ₁ , AFG ₂	20
	Milk	AFM ₁	0.5
Canada	Nuts and nut products	AFB ₁ , AFB ₂ , AFG ₁ , AFG ₂	15
Mercosur (Brazil, Argentina, Paraguay and Uruguay)	Peanuts, maize, and maize products	AFB ₁ , AFB ₂ , AFG ₁ , AFG ₂	20
	Fluid milk		
	Powdered milk	AFM ₁	0.5
Bosnia & Herzegovina	Wheat, maize, rice and cereals	AFB ₁ , AFG ₁	1
	Beans		5
China	Maize and maize products, peanut and peanut products, peanut oil, irradiated peanut	AFB ₁	20
	Rice, irradiated rice, edible vegetable oil		10
	Soy bean sauce, grain paste, vinegar, other grains, beans, fermented foods, fermented bean products, starch products, fermented wine, red rice, butter cake, pastry biscuit and bread, food additive alpha-amylase, food additive glucoamylase preparation, salad oil		5
India	All food products	AFB ₁ , AFB ₂ , AFG ₁ , AFG ₂ and AFM ₁	30
European Union	Groundnuts, nuts and dried fruits, processed products intended for direct human consumption or as ingredients in foodstuffs	AFB ₁	2
		AFB ₁ , AFB ₂ , AFG ₁ , AFG ₂	4
Australia and New Zealand	Peanuts and tree nuts	AFB ₁ , AFB ₂ , AFG ₁ , AFG ₂	15
Chile	All foods	AFB ₁ , AFB ₂ , AFG ₁ , AFG ₂	5
	Milk	AFM ₁	0.05
Japan	All foods	AFB ₁	10
Israel	Nuts, peanuts, maize flour, figs and their products, and other foods	AFB ₁	5
		AFB ₁ , AFB ₂ , AFG ₁ , AFG ₂	15
	Milk and milk products	AFM ₁	0.05
Italy	Infusion plants	AFB ₁	5
		AFB ₁ , AFB ₂ , AFG ₁ , AFG ₂	10
	Baby food	AFM ₁	0.01
Mexico	Cereals and cereal products	AFB ₁ , AFB ₂ , AFG ₁ , AFG ₂	20
	Corn flour for tortillas		12

EU member states: Austria, Belgium, Denmark, Finland, France, Germany, Greece, Ireland, Italy, Luxembourg, the Netherlands, Portugal, Spain, Sweden, the United Kingdom

Table 1. Limits for mycotoxin contamination in food products destined for human consumption in different countries.



Source: Adapted from Mykkanen et al., 2005.

Fig. 1. Pathways of aflatoxin B₁ biotransformation and excretion in humans.
(a) Experimental and human evidendence of excretion of this metabolite; (b) Scarce or no evidence available; (c) only experimental evidence available (no data for humans).

3. Role of aflatoxins in the etiology of hepatocellular carcinoma

Hepatocellular carcinoma (HCC) represents more than 80% of primary malignant tumors of the liver, and it is the 7th to 9th most common type of cancer worldwide affecting men and women, respectively. About 315,000 new cases of HCC are reported annually, a total of 4.1% of all malignant tumors in the world population. Although it is a relatively uncommon tumor, HCCs are aggressive, and mortality rates reach significant values, with about 312,000 deaths a year, and maximal survival rates of 5% in 5 years. Occurrence of HCC is associated with some degree of chronic liver disease in 90% of the cases, and it is an important cause of death in cirrhosis patients. HCC incidence has been growing, and may be directly related to the frequency of hepatitis C virus infection and longer survival of cirrhosis patients (Yang & Roberts, 2010).

HCC incidence in Africa and southeastern China is far greater than in the rest of the world. Besides known risk factors of western countries such as viral hepatitis and alcohol consumption, these populations are exposed to aflatoxin. The toxin is ingested in contaminated and stored foodstuffs, such as peanuts, maize, soybeans and rice. The association between AFB₁ and HCC is based on the ability of the toxin to induce a specific mutation of gene *p53* (Bressac et al., 1991).

In Brazil, HCC is not included among the ten most common types of cancers, probably because of underreporting. Estimates show that there are 2 to 3,000 diagnoses of the disease every year, with a national incidence of 1:100,000 inhabitants/year. Incidence of this neoplasm is greater in the north, northeast and southeast than in the south of the country. The greatest frequency occurs in the states of Amazonas, Bahia and Espírito Santo. (Pimenta & Massabki, 2010). In São Paulo, incidence is a little greater than the mean of the country, affecting about 2:100,000 inhabitants/year. In terms of mortality, HCC is the 7th death cause and is responsible for 4% of the deaths by cancer in Brazil, annually. HCC incidence rate in Brazil is associated with advanced cirrhosis in 71.2% of the cases, as observed in the rest of the world. However, serology for viral hepatitis is negative in 42% of the cases HCC, even with regional discrepancies (Gonçalves et al., 1997). This difference may be related to the exposure to AFB₁. This relationship, however, was not analyzed in the whole country.

Apparently, HCC progress is not an occasional event. Hepatocarcinogenesis seems to be a multifactorial process in which extrinsic stimuli induce gene changes in mature hepatocytes, leading to successive proliferation and cell death cycles that culminate in the production of monoclonal populations. Several lines of evidence suggest that hepatocarcinogenesis may begin in preneoplastic lesions, such as regenerative macronodules and low or high grade dysplastic nodules. Accumulation of genetic changes and new mutations in preneoplastic lesions would probably cause HCC (Theise et al., 2002).

In molecular terms, many derangements observed in HCC may be preferentially attributed to cirrhosis and inflammatory activity, and others are inherent to dysplastic nodules and to HCC itself. In early stages of chronic hepatitis, there are significant changes in the expression of growth factors, proteases and metalloproteinases, besides somatic changes, reduced apoptosis and increased expression of oncogenes and transcriptional factors. In general, these changes become more prominent and complex as the lesion progresses to fibrosis, cirrhosis, dysplastic nodules and, finally, HCC (Coleman, 2003). No tumor-suppressor gene exclusively associated with HCC has been identified. However, all molecular changes accumulated by chronic hepatitis and cirrhosis in repeated aggression / regeneration cycles, directly contribute to hepatocarcinogenesis (Fausto & Weber, 1993). Aneuploidy is also a common event in hepatocarcinogenesis. HCC is characterized by a considerable loss of heterozygosity, and includes several chromosomes, such as 1p, 4q, 6q, 8p, 8q, 9p, 13q, 16p, 16q, and 17p. Mutations

in several critical genes, such as *p73*, *p53*, *Rb*, *APC*, *DLC-1* (deleted in liver cancer), *p16*, *GSTP1*, *PTEN*, *IGF-2*, *BRCA2*, *SOCS-1*, *Smad2* and *Smad4*, β -catenin, *c-myc*, and *cyclin D1* were also identified (Fujimori et al., 1991; Tsuda et al., 1992).

Impaired control of cell cycle is an important event in carcinogenesis. The first observations involving carcinogenesis and cyclins were related to detection of the incorporation of Hepatitis B virus DNA to *cyclin A* gene in HCC (Wang et al. 1990), and to amplification of *cyclin D1* gene in some cell lineages of colon carcinoma (Leach et al., 1993). The *p16/cyclin D1/RB* pathway (retinoblastoma) may be considered the greatest cell cycle regulator. *RB* and *p16* act as tumor suppressor genes, and *cyclin D1* as an oncogene (Weinberg, 1995; Ito et al., 1999). Aberrant expression of both cyclin-dependent kinases (CDK) and CDK-inhibitors has an important role in HCC development. High expression of *cyclin D1* in HCC is variable, ranging from 6 to 76% in different studies. Among positive regulators of cell cycle, changes in *cyclin D1*, *A* and *B1* expression compared with normal tissues have been associated with increased cell growth and development of neoplasms (Ito et al., 1999).

Analysis of aberrant expression of *cyclin D1*, its biological role and its relationship with mutations in *p53* in cases of HCC demonstrated that *cyclin D1* was normally expressed in healthy livers, but it was highly reduced in 40% of the livers affected by HCC (Peng et al., 1998). Lower expression of *cyclin D1* RNA was associated with larger and less differentiated tumors. Increased expression of *cyclin D1* was observed in only 5.6% of the cases. On the other hand, *cyclin E* shows increased expression in 56% of the HCC cases. Overexpression of *cyclin E* was associated with little differentiation and with invasiveness, but not with tumor volume. Thus, decreased expression of *cyclin D1* and increased expression of *cyclin E* are intimately associated with mutation in *p53*. Besides, overexpression of *cyclin E* and concomitant loss of *p53* function seem to contribute to HCC progression (Peng et al., 1998; Jung et al., 2001).

There are three important inhibitors of cell cycle progression in the Cip/Kip family: *p27^{KIP1}*, *p21^{WAF1}*, and *p57^{KIP2}*. The most comprehensively studied of these inhibitors, in terms of clinical significance in the evolution of human cancer, is *p27^{KIP1}*. Expression of *p27^{KIP1}* is marked in non-proliferating cells, and it has important roles in the regulation of both quiescence and progression in G1 phase, by means of inhibition of cyclin / CDK complexes. Loss of *p27^{KIP1}* acts together with mutations of several oncogenes and suppressor genes, stimulating tumor growth. Reduced production of the protein synthesized by *p27^{KIP1}* is significantly involved in the stage and volume of primary tumors. Thus, *p27^{KIP1}* has been described as a crucial negative regulator of HCC progression. Its increased expression is considered an independent variable in favorable prognosis of HCC (Ito et al., 2001; Fiorentino et al., 2000). Reduced *p21^{WAF1}* expression is mainly related to mutation in gene *p53* in HCC, and also contributes to hepatocarcinogenesis. However, *p21^{WAF1}* loss was not identified as an independent factor in HCC bad prognosis (Ito et al., 2001). Compared with healthy livers, expression of *p57^{KIP2}* is significantly decreased in HCC lesions. This decreased expression of *p57^{KIP2}* was associated with highly aggressive tumors, characterized by more advanced stages, little differentiation, larger size, portal invasion, intense cellular growth and low disease-free survival rates (Ito et al., 2001).

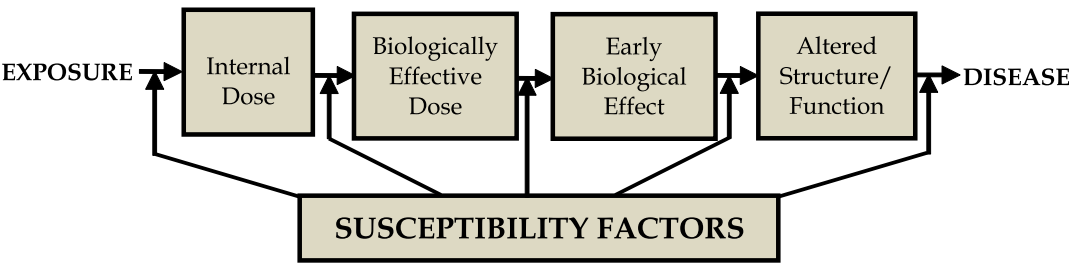
In terms of frequency, the most common molecular changes observed in HCC cases are *p53* (20-70%), *cyclin D* (11%), *p16^{Ink4}* (0-50%), *Rb* (15%) and β -catenin (16-26%), from which only *p53* mutation was reported to be associated with hepatitis B virus gene interaction and exposure to aflatoxin (Ozturk, 1999).

Although mutations in *p53* pathway have an important role in HCC pathogenesis in cases of cirrhosis, changes in cell cycle regulator genes *p21^{waf1/cip1}* and *p27^{Kip1}* are more involved in

HCC cases unrelated to cirrhosis (Tretiakova et al., 2010). These cases could be, in part, attributed to aflatoxin. Therefore, other molecular changes in genes p21^{waf1/cip1} and p27^{Kip1} should also be assessed in individuals exposed to aflatoxin.

4. Biomarkers of aflatoxin exposure

Biomarkers measure cellular, biological or molecular changes in biological tissues, cells or fluids, providing information on disease or exposure to a given substance. As biomarkers are used to measure or indicate biological processes, detection of specific biomarkers may aid identification, diagnosis and treatment of individuals who are affected and at risk, but still asymptomatic. Development of biomarkers for environmental agents should be based on specific knowledge on metabolism, formation of by-products and general mechanism of action (Groopman & Kensler, 1999). Biomarkers may be classified in four categories: internal dose, biologically effective dose, early biological response, and altered structure/function. (Figure 2). Internal dose is the amount of substance that is metabolized. Individual characteristics determine susceptibility to exposure, such as the ability to activate / detoxify carcinogens, ability to repair DNA changes, nutritional status and immunity, age, sex and socioeconomic status (WHO, 1993).



Source: Adapted from Groopman & Kensler (1999)

Fig. 2. Classification of biomarkers.

Biomarkers of exposure and effect for aflatoxins have been validated in comprehensive studies in animals and humans. Dose-response relationship between AFM₁ and AFB₁-N⁷-guanine levels and incidence of liver tumors was first established in animals (Groopman et al., 1992b). Biomarkers were then evaluated in humans to determine sensitivity, specificity, accuracy and reliability parameters. Later validation in epidemiological studies evaluated intra and inter-subject variability, the relationship biomarker-external dose and the feasibility of using them in large population studies (Groopman et al., 1992a; Groopman et al., 1992c). In a study carried out in Shanghai, People’s Republic of China, 18,244 volunteers were followed up for three years. The analysis included individual interviews on eating habits, possible previous exposure to aflatoxins, and collection of urine samples (Ross et al., 1992; Qian et al., 1994). Cases and controls were compared to detect associations between aflatoxin markers, infection by Hepatitis B virus (HBV), and hepatocellular carcinoma. Data showed a 340% increase in the relative risk for HCC when aflatoxin biomarkers were detected in urine. Relative risk in individuals showing positive results for HBV was 730% greater. Subjects who showed aflatoxin markers in urine and were positive for HBV infection had a relative risk of developing HCC 5,900% greater. These data support the relationship between the two major causes of HCC: HBV infection and exposure to AFB₁. Besides, when individual metabolites were stratified for HCC incidence, the presence of AFB₁-N⁷-guanine adduct led to 200 to 300% increase in the relative risk of developing HCC.

After the Shanghai study, a trial developed in Taiwan with about 15,000 volunteers also analyzed the relationship between HBV, exposure to AFB₁, and incidence of HCC; the results of this trial confirmed the findings of the previous study (Yu et al., 1997). Risk of developing HCC along with AFB₁ exposure was more pronounced among those individuals infected by HBV and with detectable levels of AFB₁-N⁷-guanine in their urine.

5. Occurrence of aflatoxin biomarkers in biological fluids

In past decades, several studies reported the presence of aflatoxins, metabolites and biomarkers in urine (Table 2). Zhu et al. (1987) analyzed 252 urine samples from inhabitants of Guangxi province, People's Republic of China, and reported a correlation between total daily ingestion of AFB₁ and excretion of AFM₁. Between 1.2 and 2.2 of AFB₁ ingested daily was found in urine as AFM₁, in levels ranging from 0 to 3.2 ng/mL. In a later study, the same urine samples were analyzed again and levels of AFB₁-N⁷-guanine adduct were also correlated with AFB₁ ingestion (Groopman et al., 1992c). Total amounts of AFB₁-N⁷-guanine excreted in urine in a three-day period ranged from < 50 and 3250 ng and about 0.2% of AFB₁ ingested was excreted in urine as AFB₁-N⁷-guanine. In the same study, levels of the metabolite AFP₁ did not show a significant statistical correlation between dietary exposure and excretion in urine, and the metabolite AFQ₁ was observed in few samples. Percentage of AFB₁ excreted in urine as any of these metabolites was 4.4% in women and 7.6% in men.

In another study, also carried out in Guangxi province, AFB₁-lysine adduct was determined in serum samples of 42 inhabitants and compared with AFB₁ ingestion and AFM₁ excretion in urine (Gan et al., 1988). Significant correlation coefficients were found between AFB₁-lysine levels in serum and AFM₁ in urine, and between AFB₁-lysine in serum and dietary exposure to AFB₁. It is estimated that 1.4 and 2.3% of AFB₁ ingested is covalently bound to albumin.

Qian et al. (1994) detected 55 cases of hepatocellular carcinoma. From these cases, urine samples of 50 individuals and 267 control samples were analyzed for levels of AFB₁-N⁷-guanine, AFM₁, AFP₁ and AFB₁. The metabolite detected in the greatest concentration was AFP₁, (0.59-16.0 ng/mL), whereas AFM₁ ranged from 0.17-5.2 ng/mL, and 0.3 to 1.81 ng/mL for AFB₁-N⁷-guanine adduct.

Wild et al. (1992) carried out a study with 20 individuals in Gambia, West Africa, and also confirmed the validity of AFB₁-lysine as a biomarker. Parallel evaluation of the same individuals by Groopman et al. (1992a) for AFB₁-N⁷-guanine in urine, confirmed not only the correlation between this metabolite and AFB₁, ingestion, but also demonstrated the correlation between levels of AFB₁-lysine in serum and AFB₁-N⁷-guanine in urine. AFG₁ was the most frequent metabolite observed in urine, as a consequence of the high concentration of aflatoxin found in the foodstuffs consumed by the individuals analyzed, compared with other studies in which the diet analyzed did not have AFG₁. Besides, metabolites AFQ₁ and AFP₁ were also determined, and AFM₁ was observed in some samples.

Levels of AFB₁-N⁷-guanine adducts in urine (Groopman et al., 1992a; Groopman et al., 1992c) and AFB₁-lysine in blood (Gan et al., 1988) show the biological effective dose of aflatoxin to which the individual has been exposed. Concentration of AFB₁-N⁷-guanine in urine shows exposure to AFB₁ in a 1 to 2-day period, whereas concentration of AFB₁-lysine in serum indicates 2 to 3-month exposure (Wild et al., 1992).

Urinary and fecal excretion of metabolites AFQ₁ e AFM₁ and urinary excretion of AFB₁-N⁷-guanine were evaluated in 83 university students in China (Mykkanen et al., 2005). Mean fecal AFQ₁ concentration (137 ng/g, moist weight) was about 60 times greater than mean

AFM₁ concentration (2.3 ng/g, moist weight). In urine, mean AFQ₁ concentration was 10.4 ng/mL, and 0.04 ng/mL and 0.38 ng/mL for AFM₁ and AFB₁-N⁷-guanine, respectively. The authors emphasized that, compared with other studies, differences in concentrations and frequencies of AFQ₁ and AFM₁ in their study may be attributed to differences in age and diet of the subjects. Participants of this study were young adults, 18-24 years of age, whereas in previous trials, individuals were 25 to 65 years old. Expression of CYP3A enzymes, which produce AFQ₁, decreases about 25-40% with age in animals and humans, and consumption of foodstuffs rich in flavonoids, such as green tea, may increase AFQ₁ formation by activation of these enzymes.

In Brazil, Scussel et al. (2006) evaluated the presence of AFB₁-lysine adduct in blood samples of 50 subjects in the city of Sao Paulo, in 1999. The adduct was detected in 62% of the samples, in a concentrations ranging from 0 - 57.3 pg AFB₁-lysine/ mg blood albumin. Mean concentration in positive samples was 14.9 pg/mg. Sixty-five urine samples from inhabitants of the city of Piracicaba, state of Sao Paulo, were analyzed for AFM₁ and 65% of them showed concentrations greater or equal to 1.8 pg/mL, with mean concentration of 5.96 pg/mL (Romero et al., 2010). Correlation between probable aflatoxin intake - estimated by means of questionnaires on the frequency of consumption - and AFM₁ levels in urine were not significantly correlated.

AFM₁ is also excreted in milk during lactation, and several studies demonstrated the presence of this metabolite in human milk. In the Arab Emirates, AFM₁ was detected in milk in concentrations ranging from 5 to 3400 pg/mL (Abdulrazzaq et al., 2003). In Australia, AFM₁ levels ranged from 28 to 1031 pg/mL, and in Thailand, from 39 to 1736 pg/mL (El-Nezami et al., 1995). In a study carried out in Gambia (Zarba et al., 1992), 0.09 to 0.43 % AFB₁ ingested in the diet was excreted in milk as AFM₁. In Brazil, this metabolite was studied in samples collected from human milk banks. From 50 samples analyzed, only one was contaminated by AFM₁ at a concentration of 0.024 ng/mL (Navas et al., 2005). In a recent study carried out with 160 lactating mothers in Iran, AFM₁ was detected in 157 samples, with concentrations ranging from 0.3 to 26.7 ng/kg (Sadeghi et al, 2009).

Aflatoxins were also detected in samples of umbilical cord blood, demonstrating they can cross the placenta, starting exposure to this carcinogen in the uterus (Wild et al., 1991; Turner et al., 2007).

Quantitative determination of several metabolites in complex matrices, such as serum and urine, requires specific and sensitive methods for a large number of samples. Particularly for AFB₁-lysine adduct in serum, methods may include radioimmunoassay (RIA; Gan et al., 1988), enzyme linked immunosorbent assay (ELISA; Wild et al., 1992), or purification with immunoaffinity columns followed by separation by high performance liquid chromatography (HPLC) and detection by fluorescence (Wild et al., 1992; Wang et al., 1996). As all these methods require antibodies for detection and/or purification, results will necessarily reflect the capacity, specificity and/or sensitivity of the antibody (Wang et al., 2001). Results obtained using ELISA, RIA and fluorescence were significantly different (Sheabar et al., 1993; Wild et al., 1990). ELISA is highly sensitive, but it is less specific and shows higher concentration of AFB₁-lysine due to the concomitant detection of adducts from reactions with other amino acids and ingestion of aflatoxins of similar structure, such as AFG₁. HPLC-fluorescence is specific for AFB₁-lysine, but it is not sensitive enough for epidemiological studies.

A recently developed method combines solid phase extraction and liquid chromatography-mass spectrometry (HPLC-MS/MS), showing high specificity and sensitivity (McCoy et al., 2005). The method uses a stable isotope internal standard to correct recovery and equipment variability. This method showed to be adequate for routine quantification of adducts in human serum (Scholl et al., 2006b).

Sample/ Country	Aflatoxin and metabolite	no. samples	% positive samples	Level ^a		Ref.
				Mean	Range	
Urine/ Brazil	AFM ₁	69	65	5.96 pg/mL	1.8-39.9 pg/mL	Romero et al. (2010)
Urine/ Bosnia and Herzegovina	AFB ₁ HCC patients	30	100	N.S.	0.05-0.26 µg/kg	Aljicevic & Hamzic (2010)
	Control group	30	100		0.05-0.15 µg/kg	
Urine/ China	AFM ₁	145	54	NS	0.003-0.243 ng/mL	Sun et al.(1999)
Urine/ China	AFM ₁	42	NS	NS NS	0.01-3.2 ng/mL 40-4800 ng/day	Zhu et al. (1987)
Urine/ China	AFM ₁	29	89	192 ng/day	0.9-3569 ng/day	Wang et al. (2001)
	AFB ₁ -N ⁷ -G		41	407	64.9-1789	
	AFB ₁ -Merc		89	103	6.6-494	
	AFQ ₁		26	92.2	77.3-137	
	AFP ₁		30	664	80.4-3569	
Urine/ China	AFB ₁ -N ⁷ -G (placebo)	39				El-Nezami et al (2006)
	Beginning		59.5	0.54 ng/mL	0.29-1.03 ng/mL	
	Week 3		64.3	0.63	0.34-1.16	
	Week 5		57.1	0.46	0.25-0.86	
	Final		54.2	0.45	0.24-0.83	
	AFB ₁ -N ⁷ -G (intervention with probiotics ^c)	44				
	Beginning		51.3	0.42 ng/mL	0.22-0.82 ng/mL	
	Week 3		43.6	0.27	0.15-0.47	
	Week 5		38.5	0.19	0.11-0.31	
	Final		61.5	0.45	0.26-0.79	
Urine/ China	AFM ₁ (intervention with GTP)	352	100			Tang et al. (2008)
	Beginning Placebo			59.41 pg/mg	0.42-141.99 pg/mg	

Sample/ Country	Aflatoxin and metabolite	no. samples	% positive samples	Level ^a		Ref.
				Mean	Range	
				crea	crea	
	500 mg GTP			60.85	0.59-746.10	
	1000 mg GTP			40.12	0.52-308.27	
	<u>1st month</u>					
	Placebo			61.67	0.52-881.39	
	500 mg GTP			15.03	0.38-64.27	
	1000 mg GTP			20.06	0.77-51.50	
	<u>3rd month</u>					
	Placebo			78.66	0.24-1276.25	
	500 mg GTP			16.12	0.18-222.35	
	1000 mg GTP			25.95	0.12-338.85	
	AFB₁ - Merc (intervention with GTP)					
	<u>Beginning</u>					
	Placebo			8.67 pg/mg	0.43-41.15 pg/mg	
				crea	crea	
	500 mg GTP			10.31	0.38-50.77	
	1000 mg GTP			9.32	0.60-67.71	
	<u>1st month</u>					
	Placebo			9.95	0.09-57.92	
	500 mg GTP			79.53	1.57-362.47	
	1000 mg GTP			79.48	0.30-465.62	
	<u>3rd month</u>					
	Placebo			6.11	0.43-50.58	
	500 mg GTP			97.76	11.32-501.48	
	1000 mg GTP			96.60	18.20-560.30	
Urine/ China	Total aflatoxin (AFB ₁ -N ⁷ -G, AFP ₁ . AFB ₁ . AFQ ₁)	42	NS	NS NS	1.5-2.3 ng/mL 3300-6600 ng/day	Groopman et al (1992c)
Urine/ China	AFM ₁	317	67	NS	0.17-5.2 ng/mL	Qian et al.(1994)
	AFB ₁ -N ⁷ -G		49	NS	0.3-1.81	
	AFP ₁		53	NS	0.59-16	
	AFB ₁		71	NS	NE	

Sample/ Country	Aflatoxin and metabolite	no. samples	% positive samples	Level ^a		Ref.
				Mean	Range	
Urine/ China	AFM ₁	83	83	0.04 ng/mL	0.01-0.33 ng/mL	Mykkanen et al. (2005)
	AFQ ₁			10.4	3.4-23.3	
	AFB ₁ -N ⁷ -G			0.38	0.0-2.15	
Urine/ Egypt	AFB ₁	20	30	NS	< 1.5 ng/mL	Al- Saadany (1993)
	AFM ₁			NS	< 2.5 ng/mL	
	AFG ₁			1.1 ^b		
Urine/ Egypt	AFB ₁	60	61	NS	0.01-0.15 ng/mL	Hatem et al. (2005)
Urine / Egypt (Children 1-2.5 years old)	AFB ₁	50	38	189 ^b pg/mL	- pg/mL	Polychona ki et al. (2008)
	AFB ₂			1.4	0.8-2.2	
	AFG ₁			76.6	72.1-81.1	
	AFG ₂			2.2	0.9-8.0	
	AFM ₁			5.5	5.0-6.2	
Guinea (Children 2-4 years old)	AFB ₁	50	86	2682 pg/mL	179-18000 pg/mL	
	AFB ₂			5.7	0.6-43	
	AFG ₁			709 ^b	-	
	AFG ₂			19.0	1.4-199	
	AFM ₁			97.0	8.0-801	
Urine/ Gambia	AFB ₁ -N ⁷ -G (AFG ₁ . AFM ₁ . AFP ₁ . AFQ ₁ also detected)	20	NS	NS	48.2-7099 ng/day	Groopman et al (1992b)
Urine/ United States	AFB ₁ -lys	184	20.6	3.84 pg/mg alb	1.01-16.57 pg/mg alb	Johnson et al. (2010)
	AFM ₁		11.7	223.85 pg/mg crea	1.89-935.49 pg/mg alb	
Serum/ Benin	AFB ₁ -alb	480	99	NS	5-1064 pg/mg alb	Gong et al. (2002)
Serum/ Brazil	AFB ₁ -alb	50	62	14.9 pg/mg alb	0-57.3 pg/mg alb	Scussel et al. (2006)
Serum/ Egypt	AFB ₁	60	61	NS	0.04-0.69 ng/mL	Hatem et al. (2005)
Serum/ Egypt	AFB ₁	20	55	NS	< 4.5 ng/mL	Al- Saadany (1993)
	AFM ₁			NS	<0.5	
	AFM ₂			0.2 ^b ng/mL	-	

Sample/ Country	Aflatoxin and metabolite	no. samples	% positive samples	Level ^a		Ref.
				Mean	Range	
Serum/ Gambia	AFB ₁ -alb	20	NS	44 pg/mg alb	NS	Wild et at. (1992)
Serum/ Gambia	AFB ₁ -alb	357	100		NS	Wild et at. (2000)
	Feb/Mar			83.2 pg/mg alb		
	July/Aug			34.9 pg/mg alb		
Serum/ China	AFB ₁ -alb	64	100	0.9972 pmol/mg alb	0.3325-2.2703 pmol/mg alb	Jiang et al.(2005)
Serum/ Gambia	AFB ₁ -alb	444	100	NS	2.2-459 pg/mg alb	Turner et al. (2000)
Serum/ Gambia	AFB ₁ -alb	117	100	29.3 pg/mg alb	2.2-254 pg/mg alb	Wild et at. (1993)
Serum/ Guinea	AFB ₁ -alb	600	95	NS	9.4-22 pg/mg alb	Sylla et al. (1999)
Serum/ Ghana	AFB ₁ -alb (pregnant women)	755	100	10.9 pg/mg alb	0.44-268.73 pg/mg alb	Shuaib et al. (2010)
Serum/ Ghana	AFB ₁ -alb	507	100	0.94 pmol/mg alb	0.1-4.4 pmol/mg alb	Tang et al. (2009)
Serum/ Gambia	AFB ₁ -alb					Turner et al. (2007)
	Mothers	119	100	38.9 pg/mg alb	23.3-64.1 pg/mg alb	
	Umbilical cord	99	48.5	2.5	2.5-7.9	
	Children (4 months)	118	11	2.5	2.5-2.5	
Serum/ China	AFB ₁ -alb	42	NS	NS	30-340 pg/mg alb	Gan et al. (1988)

Table 2. Aflatoxins and metabolites in human urine and serum. (a) The unit is expressed only in the first row; (b) Only one positive sample; (c) *Lactobacillus rhamnosus* LC705 and *Propionibacterium freudenreichii* subsp. *shermanii* (1:1, m:m), 2 – 5 × 10¹⁰ colony forming units/d. NS, not specified; GTP, green tea polyphenols; alb, albumin; crea, creatinine.

Methods used to determine AFB₁-N⁷-guanine adduct include immunoassays (Groopman et al., 1992a), HPLC with UV detection (Groopman et al., 1992c), or fluorescence (Wang et al., 1999; Mykkanen et al., 2005). Egner et al. (2006) described a method using HPLC-MS/MS in the analysis of AFB₁-N⁷-guanine in urine, also based on the use of stable isotope internal

standard. Precision and accuracy were far superior than previous procedures. Together with the analysis of AFB₁-lysine, determination of these two biomarkers in urine and serum samples is precise, accurate, specific and selective. Determination of residual aflatoxin and metabolites AFM₁, AFP₁ and AFQ₁ in urine has been carried out using HPLC-fluorescence (Tang et al., 2008; Polychronaki et al., 2008; Romero et al., 2010). However, HPLC-MS/MS has recently been used successfully to determine AFB₁, AFB₂, AFG₁, AFG₂, AFM₁ and AFP₁ in urine (Everley et al., 2007).

6. Concluding remarks

Current concepts derived from intensive research on biotransformation, mechanisms of toxicity and evidence of the role of aflatoxins in the etiology of human liver cancer were summarily presented in this chapter. AFB₁ exerts its effects after conversion to the reactive compound AFB₁-epoxide by means of cytochrome P450-dependent enzymes. This epoxide can form derivatives with cellular macromolecules, including proteins, RNA and DNA. Reaction with DNA occurs with guanines in codon 249 of tumor suppressor gene *p53*. Although mutations in *p53* pathway have an important role in HCC pathogenesis other molecular changes in genes *p21^{waf1/cip1}* and *p27^{Kip1}* should also be assessed in individuals exposed to aflatoxin.

Primary biotransformation of AFB₁ also produces hydroxylated and less toxic derivatives, such as aflatoxins Q₁ and P₁. Intra and interspecies differences in the pathways of activation/detoxification are directly related to the susceptibility of animals to aflatoxin effects. In humans, individual biomonitoring of AFB₁ metabolites such as AFB₁-N⁷-guanine have demonstrated that aflatoxins constitute an important risk factor for hepatocellular carcinoma in highly exposed populations. Some of these studies also show synergism between aflatoxins and hepatitis B virus in the development of human HCC. Based on these concepts, and taking into account the frequent detection of aflatoxins in foodstuffs worldwide, further investigations are needed to assess the level of dietary exposure to these toxins and its impact on human health.

7. References

- Abdulrazzaq, Y.M.; Osman, N.; Yousif, Z.M. & Al-Falahi, S. (2003). Aflatoxin M₁ in breast-milk of UAE women. *Annals of tropical paediatrics*, Vol.23, No.3, (September 2003), pp.173-179, ISSN 0272-4936
- Aguillar, F.; Hussain, S.P. & Ceerutti, P. (1993). Aflatoxin B₁ induces the transversion of G-T in codon 249 of the p53 tumor suppressor gene in human hepatocytes. *Proceeding of the National Academic Sciences of the United States of America*, Vol.90, No.18, (September 1993), pp.8586-8590, ISSN 0027:8424
- Aljicevic, M. & Hamzic, S. (2010). Aflatoxin in the urine of patients suffering from HCC and HBV. *Healthmed*, Vol.4, No.4, (January 2010), pp.852-856, ISSN 1840:2291
- Al-Saadany, A.; El-Hennawy, S.; Amra, H.; Iman Abd El-Reheim & Ezzat, S. (1993). A study of aflatoxins and Kwashiorkor in Benha. *Benha Medical Journal*, Vol.10, No.3, (September 1993), pp.125-138, ISSN 1110-208X
- Bressac, B.; Kew, M.; Wands, J. & Ozturk, M. (1991). Selective G-mutation to T-mutations of p53 gene in hepatocellular carcinoma in Southern Africa. *Nature*, Vol.350, No.6317, (April 1991), pp. 429-431, 1991, ISSN 0028-0836

- Busby, W.F. & Wogan, G.N (1984). Aflatoxins. In: *Chemical carcinogens*, Searle, C.E., , pp. 945-1136, American Chemical Society, ISBN 0841208697 , Washington, D.C., USA
- Chu, F.S. (1991). Mycotoxins: food contamination, mechanism, carcinogenic potential and preventive measures. *Mutation Research*, Vol.259, No. 3-4, (March-April 1991), pp.291-306, ISSN 0921-8262
- Coleman, W.B. (2003). Mechanisms of human hepatocarcinogenesis. *Current Molecular Medicine*, Vol. 3, No.6, (September 2003), pp. 573-588, 2003, ISSN 1566-5240
- Coulombe, R.A. (1991). Aflatoxins. In: *Mycotoxins and phytoalexins*, Sharma, R.P. & Salunkhe, p.103-143, CRC Press, ISBN 0849388333, Boca Raton
- Egner, P.A.; Groopman, J.D.; Wang, J.S.; Kensler, T.W. & Friesen, M.D. (2006). Quantification of Aflatoxin-B₁-N⁷-Guanine in Human Urine by High-Performance Liquid Chromatography and Isotope Dilution Tandem Mass Spectrometry. *Chemical Research in Toxicology*, Vol.19, No.9, (September 2006), pp.1191-1195, ISSN 0893-228X
- Egner, P.A.; Yu, X.; Johnson, J.K.; Nathasingh, C.K.; Groopman, J.D.; Kensler, T.W. & Roebuck, B.D. (2003). Identification of aflatoxin M₁-N⁷-guanine in liver and urine of tree shrews and rats following administration of aflatoxin B₁. *Chemical Research in Toxicology*, Vol.16, No.9, (September 2003), pp.1174-1180, ISSN 0893-228X
- El-Nezami, H.; Nicoletti, G.; Neal, G.E.; Donohue, D.C. & Ahokas, J.T. (1995). Aflatoxin M₁ in human breast milk samples from Victoria, Australia and Thailand. *Food and Chemical Toxicology*, Vol.33, No.3, (March 1995), pp.173-179, ISSN 0278-6915
- El-Nezami, H.S.; Polychronaki, N.N.; Ma, J.; Zhu, H.; Ling, W.; Salminen, E.K.; Juvonen, R.O.; Salminen, S.J.; Poussa, T. & Mykkanen, M. (2006). Probiotic supplementation reduces a biomarker for increased risk of live cancer in young men from Southern China. *The American Journal of Clinical Nutrition*, Vol.83, No.5, (May 2006), pp.1199-1203, ISSN 0002-9165
- Essigmann, J.M.; Croy, R.G.; Nadzan, A.M.; Busby, W.F.; Reinhold, V.N.; Buchi, G. & Wogan, G.N. (1977). Structural identification of the major DNA adduct formed by aflatoxin B₁ in vitro. Proceedings of the National Academy of Sciences of the United States of America, Vol.74, No.5, pp.1870-1874, ISSN 0027-8424
- Essigmann, J.M.; Croy, R.G.; Bennett, R.A. & Wogan, G.N. (1982). Metabolic activation of aflatoxin B₁: patterns of DNA adduct formation, removal, and excretion in relation to carcinogenesis. *Drug Metabolism Reviews*, Vol.13, No.4, pp. 581-602, ISSN 0360-2532
- Everley, R.A.; Ciner, F.L.; Zhang, D.; Scholl, P.F.; Groopman, J.D. & Croley, T.R. (2007). Measurement of aflatoxin and aflatoxin metabolites in urine by liquid chromatography-tandem mass spectrometry. *Journal of Analytical Toxicology*, Vol.31, No.3, (April 2007), pp.150-156, ISSN 0146-4760
- Fausto, N. & Webber, E.M. (1993). Control of liver growth. *Critical Reviews in Eukaryotic Gene Expression*, Vol.3, No.2 pp.117-135, ISSN 1045-4403
- Fiorentino, M.; Altimari, A.; D'Errico, A.; Cukor, B.; Barozzi, C.; Loda, M. & Grigioni, W.F. (2000). Acquired expression of p27 is a favorable prognostic indicator in patients with hepatocellular carcinoma. *Clinical Cancer Research*, Vol.6, No.10, (October 2000), pp.3966-3972, ISSN 1078-0432
- Food and Agriculture Organization of the United Nations - FAO. (2004). Worldwide regulations for mycotoxins in food and feed in 2003. FAO Food and Nutrition Paper No. 81. pp. 1728-3264, Food and Agriculture Organization of the United Nations, ISBN 9251051623, Rome, Italy.

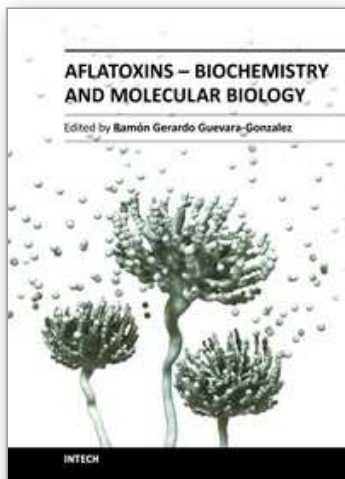
- Fujimori, M.; Tokino, T.; Hino, O.; Kitagawa, T.; Imamura, T.; Okamoto, E.; Mitsunobu, M.; Ishikawa, T.; Nakagama, H. & Harada, H. (1991). Allelotype study of primary hepatocellular carcinoma. *Cancer Research*, Vol.51, No.1, (January 1991), pp.89-93, ISSN 0008-5472
- Gan, L.S.; Skipper, P.L.; Peng, X.C.; Groopman, J.D.; Chen, J.S.; Wogan, G.N. & Tannenbaum, S.R. (1988). Serum albumin adducts in the molecular epidemiology of aflatoxin carcinogenesis: correlation with aflatoxin B₁ intake and urinary excretion of aflatoxin M₁. *Carcinogenesis*, Vol.9, No.7, (July 1988), pp.1323-1325, ISSN 0143-3334
- Gonçalves, C.S.; Pereira, F.E.L. & Gayotto, L.C.C. (1997). Hepatocellular carcinoma in Brazil: report of a national survey (Florianopolis, SC, 1995). *Revista do Instituto de Medicina Tropical de São Paulo*, Vol.39, No.3, (May-June 1997), pp.165-70, 1997, ISSN 0036-4665
- Gong, Y.Y.; Cardwell, K.; Hounsa, A.; Egal, S.; Turner, P.c.; Hall, A.J. & Wild, C.P. (2002). Dietary aflatoxin exposure and impaired growth in young children from Benin and Togo: cross sectional study. *British Medical Journal*, Vol.325, No.7354, (July 2002), pp.20-21, ISSN 0959-535X
- Groopman, J.D.; Hall, A.J.; Whittle, H.; Hudson, G.J.; Wogan, G.N.; Montesano, R. & Wild, C.P. (1992a). Molecular dosimetry of aflatoxin-N⁷-guanine in human urine obtained in The Gambia, West Africa. *Cancer epidemiology, biomarkers & prevention*, Vol. 1, No.3, (March-April 1992), pp.221-227, ISSN 1055-9965
- Groopman, J.D.; Hasler, J.A.; Trudel, L.J.; Pikul, A.; Donahue, P.R. & Wogan, G.N. (1992b). Molecular dosimetry in rat urine of aflatoxin-N⁷-guanine and other aflatoxin metabolites by multiple monoclonal antibody affinity chromatography and immunoaffinity/high performance liquid chromatography. *Cancer Research*, Vol.52, No.2, (January 1992), pp.267-274, ISSN 0008-5472
- Groopman, J.D. & Kensler, T.W. (1999). The light at the end of the tunnel for chemical-specific biomarkers: daylight or headlight? *Carcinogenesis*, Vol.20, No.1, (January 1999), pp.1-11, ISSN 0143-3334
- Groopman, J.D.; Zhu, J.Q.; Donahue, P.R.; Pikul, A.; Zhang, L.S.; Chen, J.S. & Wogan, G.N. (1992c). Molecular dosimetry of urinary aflatoxin-DNA adducts in people living in Guangxi Autonomous Region, People's Republic of China. *Cancer Research*, Vol.52, No.1, (January 1992), pp.45-52, ISSN 00088-5472
- Guengerich, F.P. (1999). Cytochrome P-450 3A4: regulation and role in drug metabolism. *Annual Review of Pharmacology Toxicology*, Vol.39, pp.1-17, ISSN 0362-1642
- Hatem, N.L.; Hassab, H.M.; Abd Al-Rahman, E.M.; El-Deeb, S.A. & El-Sayed Ahmed, R.L. (2005). Prevalence of aflatoxins in blood and urine of Egyptian infants with protein-energy malnutrition. *Food & Nutrition Bulletin*, Vol.26, No.1, pp.49-56, ISSN 0379-5721
- Hsieh, D.P.H. & Atkinson, D.N. Bisfuranoid mycotoxins: their genotoxicity and carcinogenicity. (1991). *Advances in Experimental Medicine and Biology*, Vol.283, pp.525-532, ISSN 0306437376
- International Agency for Research on Cancer - IARC. (1993). Some Naturally Occurring Substances: Food Items and Constituents, Heterocyclicaromatic Amines and Mycotoxins. In: *IARC monographs on the evaluation of carcinogenic risks to humans*. pp. 245-395, International Agency for Research on Cancer, ISBN 928321256-8, Lyon, France
- Ito, Y.; Matsuura, N.; Sakon, M.; Miyoshi, E.; Noda, K.; Takeda, T.; Umeshita, K.; Nagano, H.; Nakamori, S.; Dono, K.; Tsujimoto, M.; Nakahara, M.; Nakao, K.; Taniguchi, N. & Monden, M. (1999). Expression and prognostic roles of the G1-S modulators in

- hepatocellular carcinoma: P27 independently predicts the recurrence. *Hepatology*, Vol.30, No.1, (July 1999), pp.90-99, ISSN 0270-9139
- Ito, Y.; Takeda, T.; Sakon, M.; Tsujimoto, M.; Monden, M.; Matsuura, N. (2001). Expression of p57/Kip2 protein in hepatocellular carcinoma. *Oncology*, Vol.61, No.3, pp.221-225, ISSN 0030-2414
- Jiang, Y.; Jolly, P.E.; Ellis, W.O.; Wang, J.S.; Phillips, T.D. & Williams, J.H. (2005). Aflatoxin B₁ albumin adduct levels and cellular immune status in Ghanaians. *International Immunology*, Vol.17, No.6, (June 2005), pp.807-814, ISSN 0953-8178
- Johnson, N.M.; Qian, G.; Xu, L.; Tietze, D.; Marroquin-Cardona, A.; Robinson, A.; Rodriguez, M.; Kaufman, L.; Cunningham, K.; Wittmer, J.; Guerra, F.; Donnelly, K.C.; Williams, J.H.; Wang, J.-S. & Phillips, T.D. (2010). Aflatoxin and PAH exposure biomarkers in a U.S. population with a high incidence of hepatocellular carcinoma. *Science of the Total Environment*, Vol.408, No.23, (November 2010), pp.6027-6031, ISSN 0048-9697
- Jung, Y.J.; Lee, K.H.; Choi, D.W.; Han, C.J.; Jeong, S.H.; Kim, K.C.; Oh, J.W.; Park, T.K. & Kim, C.M. (2001). Reciprocal expressions of cyclin E and cyclin D1 in hepatocellular carcinoma. *Cancer Letters*, Vol.168, No.1, (July 2001), pp.57-63, ISSN 0304-3835
- Leach, F.S.; Elledge, S.J.; Sherr, C.J.; Willson, J.K.; Markowitz, S.; Kinzler, K.W. & Vogelstein, B. (1993). Amplification of cyclin genes in colorectal carcinomas. *Cancer Research*, Vol.53, No.9, (May 1993), pp.1986-1989, ISSN 00088-5472
- Leeson, S.; Diaz, G.J. & Summers, J.D. (1995). *Poultry metabolic disorders and mycotoxins*. Guelph, University Books, ISBN 096956001, Ontario, Canada
- McCoy, L., Scholl, P. F., Schleicher, R., Groopman, J. D., Powers, C. & Pfeiffer, C. M. (2005). Analysis of aflatoxin B₁-lysine adducts in serum by isotope dilution liquid chromatography tandem mass spectrometry. *Rapid communications in mass spectrometry*, Vol.19, No.16, pp.2203-2210, ISSN 0951-4198
- Moss, M.O. Recent studies of mycotoxins. (1998). *Journal of Applied Microbiology*, Vol.84, Supplement, pp.62S-76S, ISSN 1364- 5072
- Mykkanen, H.; Zhu, H.; Salminen, E.; Juvonen, R.O.; Ling, W.; Ma, J.; Polychronaki, N.; Kemplainen, H.; Mykkanen, O.; Salminen, S. & El-Nezami, H. (2005). Fecal and urinary excretion of aflatoxin B₁ metabolites (AFQ₁, AFM₁ and AFB-N⁷-guanine) in young Chinese males. *International Journal of Cancer*, Vol.115, No.6, (July 2005), pp.879-84, ISSN 0020-7136
- Navas, A.; Sabino, M. & Rodriguez-Amaya, D. B. Aflatoxin M₁ and ochratoxin A in a human milk bank in the city of São Paulo, Brazil. (2005). *Food Additives and Contaminants*, Vol.22, No.5, (May 2005), pp. 57-462, ISSN 0265-203X
- Neal, G.E.; Eaton, D.L.; Judah, D.J. & Verma, A. Metabolism and toxicity of aflatoxins M₁ and B₁ in human-derived in vitro systems. (1998). *Toxicology and Applied Pharmacology*, Vol.151, No.1, (July 1998), pp.152-158, ISSN 0041-008X
- Ozturk, M. (1999). Genetic aspects of hepatocellular carcinogenesis. *Seminars in Liver Disease*, Vol.19, No.3, pp.235-242, ISSN 0272-8087
- Peng, S.Y.; Chou, S.P. & Hsu, H.C. (1998). Association of downregulation of cyclin D1 and of overexpression of cyclin E with p53 mutation, high tumor grade and poor prognosis in hepatocellular carcinoma. *Journal of Hepatology*, Vol.29, No.2, (August 1998), pp.281-289, ISSN 0168-8278
- Pimenta, J.R. & Massabki, P.S. (2010). Carcinoma hepatocelular: um panorama clínico. *Revista da Sociedade Brasileira de Clínica Médica*, Vol.8, pp.59-67, ISSN 1679-1010
- Polychronaki, N.; Wild, C.P.; Mykkanen, H.; Amra, H.; Abdel-Wahhab, M.; Sylla, A.; Diallo, M.; El-Nezami, H. & Turner, P.C. (2008). Urinary biomarkers of aflatoxin exposure

- in young children from Egypt and Guinea. *Food and Chemical Toxicology*, Vol.46, No.2, (February 2008), pp.519-526, ISSN 0278-6915
- Orsi, R.; Oliveira, C.A.F.; Dilkin, P.; Xavier, J.; Direito, G.; Correa, B. (2007). Effects of oral administration of aflatoxin B₁ and fumonisin B₁ in rabbits (*Oryctolagus cuniculus*). *Chemico-Biological Interactions*, Vol. 170, No.3 (December 2007), pp. 201-208, ISSN 0009-2797
- Qian, G.S.; Ross, R.K.; Yu, M.C.; Yuan, J.M.; Gao, Y.T.; Henderson, B.E.; Wogan, G.N. & Groopman, J.D. (1994). A follow-up study of urinary markers of aflatoxin exposure and liver cancer risk in Shanghai, People's Republic of China. *Cancer epidemiology, biomarkers & prevention*, Vol.3, No.1, (January February 1994), pp.3-10, ISSN 1055-9965
- Raney, K.D.; Shimada, T.; Kim, D.H.; Groopman, J.D.; Harris, T.M. & Guengerich, F.P. (1992). Oxidation of aflatoxins and sterigmatocystin by human liver microsomes: significance of aflatoxin Q₁ as a detoxication product of aflatoxin B₁. *Chemical Research in Toxicology*, Vol.5, No.2, (March-April 1992), pp.202-10, ISSN 0893-228X
- Romero, A.C.; Ferreira, T.R.B.; Dias, C.T.S.; Calori-Domingues, M.A. & Gloria, E.M. (2010). Occurrence of AFM₁ in urine samples of a Brazilian population and association with food consumption. *Food Control*, Vol.21, No. 4, (April 2010), pp.554-558, ISSN 0956-7135
- Ross, R.K.; Yuan, J.M.; Yu, M.C.; Wogan, G.N.; Qian, G.S.; Tu, J.T.; Groopman, J.D.; Gao, Y.T. & Henderson, B.E. (1992). Urinary aflatoxin biomarkers and risk of hepatocellular carcinoma. *Lancet*, Vol.339, No.8799, (April 1992), pp.943-946, ISSN 0140-6736
- Sabbioni, G.; Skipper, P.L.; Buchi, G. & Tannenbaum, S.R. (1987). Isolation and characterization of the major serum albumin adduct formed by aflatoxin B₁ in vivo in rats. *Carcinogenesis*, Vol.8, No.6, (June 1987), pp.819-824, ISSN 0143-3334
- Sadeghi, N.; Oveisi, M.R.; Jannat, B.; Hajimahmoodi, M.; Bonyani, H. & Jannat, F. (2009). Incidence of aflatoxin M₁ in human breast milk in Tehran, Iran. *Food Control*, Vol.20, No.1, (January 2009), pp.75-78, ISSN 0956-7135
- Scholl, P.F.; Turner, P.C.; Sutcliffe, A.E.; Sylla, A.; Diallo, M.S.; Frisen, M.D.; Groopman, J.D. & Wild, C.P. (2006). Quantitative comparison of aflatoxin B₁ serum albumin adducts in humans by isotope dilution mass spectrometry and ELISA. *Cancer epidemiology, biomarkers & prevention*, Vol.15, No.4, (April 2006), pp.823-826, ISSN 1055-9965
- Scussel, V.M.; Haas, P.; Gong, Y. Y.; Turner, C. P. & Wild, C. P. (2006). Study of aflatoxin exposure in a Brazilian population using an aflatoxin-albumin biomarker. *11th International IUPAC Symposium on Mycotoxins and Phycotoxins. Proceedings*, pp.197-202, ISBN 978-90-8686-007-4
- Sheabar, F.Z.; Groopman, J.D.; Qian, G.-S. & Wogan, G.N. (1993). Quantitative Analysis of Aflatoxin Albumin adducts. *Carcinogenesis*, Vol.14, No.6, (June 1993), pp. 1203-1208, ISSN 0143-3334
- Shuaib, F.M.B.; Jolly, P.E.; Ehiri, J.E.; Yatich, N.; Jiang, Y.; Funkhouser, E.; Person, S.D.; Wilson, C.; Ellis, W.O.; Wang, J-S. & Williams, J.H. (2010). Association between birth outcomes and aflatoxin B₁ biomarker blood levels in pregnant women in Kumasi, Ghana. *Tropical Medicine and International Health*, Vol. 15, No.2, (February 2010), pp.160-167, ISSN 1360-2276
- Sun, Z.; Lu, P.; Gail, M.H.; Pee D.; Zhang, Q.; Ming, L.; Wang, J.; Wu, Y.; Liu, G. & Zhu, Y. (1999). Increased risk of hepatocellular carcinoma in male hepatitis B surface antigen carriers with chronic hepatitis who have detectable urinary aflatoxin metabolite M₁. *Hepatology*, Vol.30, No.2, (August 2010), pp.379-383, ISSN 0270-9139

- Sylla, A.; Diallo, M.S.; Castegnaro, J. & Wild, C.P. (1999). Interactions between hepatitis B virus infection and exposure to aflatoxins in the development of hepatocellular carcinoma: a molecular epidemiology approach. *Mutation Research*, Vol.428, No.1-2, (July 1999), pp.187-196, ISSN 0027-5107
- Tang, L.; Meng, T.; Xu, L.; Luo, H.; Huang, T.; Yu, J.; Zhang, L.; Gao, W.; Cox, S.B. & Wang, J-S. (2008). Modulation of aflatoxin biomarkers in human blood and urine by green tea polyphenols intervention. *Carcinogenesis*, Vol.29, No.2, (February 2008), pp.411-417, ISSN 0143-3334
- Tang, L.; Xu, L.; Afriyie-Gyawu, E.; Liu, W.; Wang, P.; Tang Y.; Wang, Z.; Huebner, H.J.; Ankrah, N.-A.; Ofori-Adjei, D.; Williams, J.H.; Wang, J.-S. & Phillips, T.D. (2009). Aflatoxin-albumin adducts and correlation with decreased serum levels of vitamins A and E in an adult Ghanaian population. *Food additives and contaminants*, Vol.26, No.1, pp.108-118, ISSN 0265-203X
- Theise N.D.; Park Y.N. & Kojiro M. (2002). Dysplastic nodules and hepatocarcinogenesis. *Clinics in Liver Disease*, Vol.6, No.2, (May 2002), pp.497-512, ISSN 1089-3261
- Tretiakova, M.S.; Shabani-Rad, M.T.; Guggisberg, K.; Hart, J.; Anders, R.A. & Gao, Z.H. (2010). Genomic and immunophenotypical differences between hepatocellular carcinoma with and without cirrhosis. *Histopathology*, Vol.56, No.6, (May 2010), pp.683-693, ISSN 0309-0167
- Tsuda, H.; Oda, T.; Sakamoto, M. & Hirohashi, S. (1992). Different pattern of chromosomal allele loss in multiple hepatocellular carcinomas as evidence of their multifocal origin. *Cancer Research*. Vol.52, No.6, (March 1992), pp.1504-1509, ISSN 0008-5472
- Turner, P.C.; Collison, A.C.; Cheung, Y.B.; Gong, Y.Y.; Hall, A.J.; Prentice, A.M. & Wild, C.P. (2007). Aflatoxin exposure in utero causes growth faltering in Gambia infants. *International Journal of Epidemiology*, Vol.36, No.5, (October 2007), pp.1119-1125, ISSN 0300-5771
- Turner, P.C.; Mendy, M.; Whittle, H.; Fortuin, M.; Hall, A.J. & Wild, C.P. (2000). Hepatitis B infection and aflatoxin biomarker levels in Gambian children. *Tropical Medicine & International Health*, Vol.5, No.12, (December 2000), pp.837-841, ISSN 1360-2276
- Wang, J.; Chenivesse, X.; Henglein, B. & Brechot, C. (1990). Hepatitis B virus integration in a cyclin A gene in a hepatocellular carcinoma. *Nature*, Vol.343, No.6258 (February 1990), pp.555-557, ISSN 0028-0836
- Wang, J.S.; Huang, T.; Su, J.; Liang, F.; Wei, Z.; Liang, Y.; Luo, H.; Kuang, S.Y.; Qian, G.S.; Sun, G.; He, X.; Kensler, T.W. & Groopman, J.D. (2001). Hepatocellular carcinoma and aflatoxin exposure in Zhuqing Village, Fusui County, People's Republic of China. *Cancer epidemiology, biomarkers & prevention*, Vol.10, No.2, (February 2001), pp.143-146, ISSN 1055-9965
- Wang, J.-S.; Shen, X.; He, X.; Zhu, Y.-R.; Zhang, B. C.; Wang, J.-B.; Qian, G.-S.; Kuang, S.-Y.; Zarba, A.; Egner, P. A.; Jacobson, L. P.; Munoz, A.; Helzlsouer, K. J.; Groopman, J. D. & Kensler, T. W. (1999). Protective alterations in phase 1 and 2 metabolism of aflatoxin B₁ by oltipraz in residents of Qidong, People's Republic of China. *Journal of the national cancer institute*, Vol.91, No.4, (February 1999), pp.347-354, ISSN 0027-8874
- Wang, L.Y.; Hatch, M.; Chen, C.J.; Levin, B.; You, S.L.; Lu, S.N.; Wu, M.H.; Wu, W.P.; Wang, L.W.; Wang, Q.; Huang, G.T.; Yang, P.M.; Lee, H.S. & Santella, R.M. (1996). Aflatoxin exposure and risk of hepatocellular carcinoma in Taiwan. *International Journal of cancer*, Vol.67, No.5, (September 1996), pp.620-625, ISSN 0020-7136
- Weinberg, R.A. (1995). The retinoblastoma protein and cell cycle control. *Cell*, Vol.81, No.3 (May 1997), pp.323-330, ISSN 0092-8674

- Wild, C.P.; Fortuin, M.; Donato, F.; Whittle, H.C.; Hall, A.J.; Wolf, C.R. & Montesano, R. (1993). Aflatoxin, liver enzymes, and hepatitis B virus infection in Gambian children. *Cancer epidemiology, biomarkers & prevention*, Vol.2, No.6, (November–December 1993), pp.555-61, ISSN 1055-9965
- Wild, C.P.; Hudson, G.J.; Sabbioni, G.; Chapot, B.; Hall, A.J.; Wogan, G.N.; Whittle, H.; Montesano, R. & Groopman, J.D. (1992). Dietary intake of aflatoxins and the level of albumin-bound aflatoxin in peripheral blood in The Gambia, West Africa. *Cancer epidemiology, biomarkers & prevention*, Vol.1, No.3, (March–April 1992), pp.229-234, ISSN 1055-9965
- Wild, C.P.; Jiang, Y.-Z.; Sabbioni, G.; Chapot, B. & Montesano, R. (1990). Evaluation of methods for quantification of aflatoxin-albumin adducts and their application to human exposure assessment. *Cancer Research*, Vol.50, (January 1990), pp.245-250 ISSN 0008-5472
- Wild, C.P.; Rasheed, F.N.; Jawla, M.F.; Hall, A.J.; Jansen, L.A. & Montesano, R. (1991). In-utero exposure to aflatoxin in West Africa. *The Lancet*, Vol. 337, No. 8756, (June 1991), pp.1602, ISSN 0140-6736
- Wild, C.P.; Yin, F.; Turner, P.C.; Chemin, I.; Chapot, B.; Mendy, M.; Whittle, H.; Kirk, G.D. & Hall, A.J. (2000). Environmental and genetic determinants of aflatoxin-albumin adducts in the Gambia. *International Journal of cancer*, Vol.86, No. 1, (April 2000), pp.1-7, pp. ISSN 0020-7136
- Wilson R.; Ziprin, R.; Ragsdale, S. & Busbee, D. (1985). Uptake and vascular transport of ingested aflatoxin. *Toxicology Letters*, Vol.29, No.2-3, (December 1985), pp.169-176, ISSN 0378-4274
- Wogan, G.N.; Edward, G.S. & Shank, R.C. (1967). Excretion and tissue distribution of radioactivity from aflatoxin B₁-C¹⁴ in rats. *Cancer Research*, Vol. 27, No.10P1, 1729-1736, ISSN 0008-5472
- World Health Organization (WHO). (1993). *Biomarkers and risk assessment: concepts and principles*, World Health Organization, ISBN 9241571551, Geneva
- World Health Organization (WHO). (2002). *Evaluation of certain mycotoxins in food*, World Health Organization, ISBN 9241209062, Geneva
- Yang, J.D. & Roberts, L.R. (2010). Hepatocellular carcinoma: A global view. *Nature Review Natural Gastroenterology & Hepatology*. Vol.7, No. 8, (August 2010), pp. 448-458, ISSN 1759-5045
- Yu, M.W.; Lien, J.P.; Chiu, Y.H.; Santella, R.M.; Liaw, Y.F. & Chen, C.J. (1997). Effect of aflatoxin metabolism and DNA adduct formation on hepatocellular carcinoma among chronic hepatitis B carriers in Taiwan. *Journal of Hepatology*, Vol.27, No.2, (August 1997), pp.320-330, ISSN 0168-8278
- Zarba, A.; Wild, C.P.; Hall, A.J.; Montesano, R.; Hudson, G.J. & Groopman, J.D. (1992). Aflatoxin M₁ in human breast milk from The Gambia, West Africa, quantified by combined monoclonal antibody immunoaffinity chromatography and HPLC. *Carcinogenesis*, Vol.13, No.5, (May 1992), pp.891-894, ISSN 0143-3334
- Zhu, J.Q.; Zhang, L.S.; Hu, X.; Xiao, Y.; Chen, J.S.; Xu, Y.C.; Fremy, J. & Chu, F.S. (1987). Correlation of dietary aflatoxin B₁ levels with excretion of aflatoxin M₁ in human urine. *Cancer Research*, Vol.47, No.7, (April 1987), pp.1848-1852, ISSN 0008-5472



Aflatoxins - Biochemistry and Molecular Biology

Edited by Dr. Ramon G. Guevara-Gonzalez

ISBN 978-953-307-395-8

Hard cover, 468 pages

Publisher InTech

Published online 03, October, 2011

Published in print edition October, 2011

Aflatoxins – Biochemistry and Molecular Biology is a book that has been thought to present the most significant advances in these disciplines focused on the knowledge of such toxins. All authors, who supported the excellent work showed in every chapter of this book, are placed at the frontier of knowledge on this subject, thus, this book will be obligated reference to issue upon its publication. Finally, this book has been published in an attempt to present a written forum for researchers and teachers interested in the subject, having a current picture in this field of research about these interesting and intriguing toxins.

How to reference

In order to correctly reference this scholarly work, feel free to copy and paste the following:

Alessandra Vincenzi Jager, Fernando Silva Ramalho, Leandra Náira Zambelli and Carlos Augusto Fernandes Oliveira (2011). Biomarkers of Aflatoxin Exposure and Its Relationship with the Hepatocellular Carcinoma, Aflatoxins - Biochemistry and Molecular Biology, Dr. Ramon G. Guevara-Gonzalez (Ed.), ISBN: 978-953-307-395-8, InTech, Available from: <http://www.intechopen.com/books/aflatoxins-biochemistry-and-molecular-biology/biomarkers-of-aflatoxin-exposure-and-its-relationship-with-the-hepatocellular-carcinoma>

INTECH
open science | open minds

InTech Europe

University Campus STeP Ri
Slavka Krautzeka 83/A
51000 Rijeka, Croatia
Phone: +385 (51) 770 447
Fax: +385 (51) 686 166
www.intechopen.com

InTech China

Unit 405, Office Block, Hotel Equatorial Shanghai
No.65, Yan An Road (West), Shanghai, 200040, China
中国上海市延安西路65号上海国际贵都大饭店办公楼405单元
Phone: +86-21-62489820
Fax: +86-21-62489821

© 2011 The Author(s). Licensee IntechOpen. This is an open access article distributed under the terms of the [Creative Commons Attribution 3.0 License](https://creativecommons.org/licenses/by/3.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

IntechOpen

IntechOpen