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Medulloblastoma – Genetic Alterations

Esther Manor¹ and Lipa Bodner²

¹*Genetic Laboratories, Institute of Human Genetics, Soroka University Medical Center
Ben Gurion University of the Negev*

²*Department of Oral and Maxillofacial Surgery, Soroka University Medical Center
Ben Gurion University of the Negev
Israel*

1. Introduction

Medulloblastoma (MB) is an embryonal brain tumor of the cerebellum. MB accounts for 4-6% of all primary intracranial tumors. It is the most common malignant brain tumor in children it represents 15-30% of all pediatric brain tumors, with 85% of MBs being diagnosed in patients younger than 18 years of age. The peak incidence occurs between the ages 3-9 years, with about 35% metastasis disease (Crawford *et al.*, 2007; Polkinghorn and Tarbell 2007; Rossi *et al.*, 2008).

In contrast, medulloblastoma is rare in adults and accounts for less than 1% of primary intracranial malignancies in this age group (Brandes *et al.* 2009; Louis 2007; Padovani *et al.* 2007).

Based on histopathological features, the 2007 WHO classification of CNS tumors (Louis *et al.*, 2007; Gilbertson and Ellison 2008), has separated MB into five recognizable subtypes: the classic tumor which is the most common subtype of MB in general and in children in particular. The other four variants are: 1. desmoplastic/nodular which is the second most common (10-20% of the cases) in very young patients and adults, 2. MB with extensive nodularity is predominantly observed in infants (less than 3 years old), 3. anaplastic MB, 4. large cell MB is rare in infants, both of which account for 5-10% of the cases

The histological classification and clinical staging have proven to be less than ideal methods for stratification. Histological subtypes are often quite heterogeneous and exhibit highly variable clinical behavior, with anaplastic subtype typically associated with the worst prognosis, followed by classic and desmoplastic/nodular MB, which correlate with improved overall survival.

MB with extensive nodularity and desmoplastic/nodular MB in infants have a better outcome than the classic MB tumors, while large cell and anaplastic MB behave aggressively. Thus infants with desmoplastic MB should be stratified into a low risk therapeutic group while the large cell MB is rare but points to a poor prognosis that often presents with metastatic disease (Gilbertson and Ellison 2008).

The increasing recognition of MB as a heterogeneous disease, with histological and molecular variants that have distinct biological behavior, affected the disease classification and treatment.

In spite of current aggressive therapies, approximately one third of the patients eventually succumb to the disease due to metastasis. Furthermore the post-treatment sequelae, which

includes neurological, vascular and long-term neurocognitive impairments, can be devastating for survivors of the disease (Ribi et al., 2005).

It is imperative to gain a better understanding of the molecular alterations and other biological sequelae in medulloblastoma for future targeted therapies that are more effective and less toxic.

A combination of clinicopathological evaluation and assays based on molecular subgroups of the disease allow stratification of patients into risk groups and more tailored approach to therapy which should prevent the significant adverse effects among survivors.

Studies in the last 5 years have helped to build a consensus on the value and means of using molecular markers in the therapeutic stratification of childhood medulloblastoma (Gilbertson 2004 ; Packer and Vezina 2008 ; Pizer and Clifford 2009).

These studies have revealed the genetic heterogeneity within MB including dysregulation of several signaling pathways. The genetic heterogeneity appears to be the basis for differential response to treatment and give a new impact to the traditional histological subtyping and improve to some extent the accuracy of diagnosis and prognosis and as a consequence the stratification to risk groups (Packer and Vezina 2008).

Although, adult MB is considered different from their childhood counterparts in terms of tumor biology and clinical variables, yet, because of the high incidence of MB in childhood and low incidence in adults most of the studies have been done on the MB of childhood.

Some studies documented differences between childhood and adults tumors with regards to localization, cell of origin, histopathologic features, tumor cell differentiation, and treatment outcome. The adult's MB found to be also genetically distinct from the pediatric MB (Korshunov et al 2010).

Here we focused on the genetic alteration reported so far in MB and their significance in sub-typing, stratification, and medical care.

2. Heritability of medulloblastoma

Neither familial cases of MB, nor MB's cases affected by defined environmental factors have been described so far. Also, no specific chromosomal aberration has been described in more than about 40% of the MB cases. Unlike Ewing sarcoma, where t(11;21) can be found in about 90% of the cases, in MB no main specific gene alteration can be detected. Yet, there is a wide basis to believe that inheritable factors play a significant role in MB pathogenesis and prevalence. Some of the evidence that support this point of view are:

A. Age of onset- MB is most prevalent in childhood including about 3% in infancy, mainly medulloblastoma with extensive nodularity, while it is rare in adults. MB is mostly a childhood brain tumor with a peak in 3-9 years age. Inheritable markers are most probably expressed phenotypically at the early ages of life. The younger the patient is, the higher chance that the genetic part is more pronounced, and so in the adults, the latter the age of tumor appearance the higher chance for multi-factorial environmental cause. About 85% of the primary intracranial tumors diagnosed as MB in patients younger than 18 years of age while in adults about 1% of the primary intracranial tumors diagnosed as MB. It is well known that infants are the more vulnerable group with 25-60% rate of cure, while the overall 5-year survival rate in childhood is about 80%. Although, the environmental factors could not be totally excluded, yet the younger age of MB appearance with 35% metastatic, points toward a more pronounced genetic background rather than environmental (Rossi et al 2008, Bar and Stearns 2008).

B. Severity of the disease- the most prevalent MB sub-type in infancy is the desmoplastic which is considered the most aggressive sub-type especially when it also appear with metastasis , 35% of the childhood MB are metastatic at presentation. Metastatic phenotype considered as a high risk group with the worst prognosis. Cure rates are lower for infants than for older children. This might point either to the more significant-effective role of the inheritage genetics predisposition in infants then in elderly or adults MB. Also most probably radiotherapy can be particularly more adverse in the immature infant brain (Grill et al 2005; Packer and Vezina 2008; Rutkowski et al., 2009). The main consequence of carniospinal irradiation (CSI) is notably cognitive problems children's IQ can decrease by as much as 30 points in the standard risk group. The high risk group (metastasis, significant post operative residual tumor) remains poor 5-years event free survival (EFS) being 25-40% the worse prognosis is particularly in younger children

Although 5-years overall survival rates have reached 60-80% , survivors often face a variety of long term neurological , neuroendocrine , and social squeal as a result of conventional treatment regimens (surgery, radiotherapy, and chemotherapy (Pizer and Clifford 2009) (Packer and Vezina 2008; Rossi et al 2008, Bar and Stearns 2008). It is imperative to gain a better understanding of the molecular alterations in MB for future targeted therapies that are more effective and less toxic.

C. Predisposition- Predisposition to develop MB due to known genetic alterations- some MB cases are associated with known hereditary syndromes such as Gorlin, Turcot and Li-Fraumeni. (Reviewed by Ellison 2010, Northcott, 2011; Onvani 2010) will be discussed in the gene alteration part. All the above points support the suggestion that genetic factors cause predisposition for MB's development.

Most studies on MB have focused on genes that are already known to be involved in the pathogenesis of tumors, such as Wnt, SHH, Notch TP53. The specific genetic pathway/s in MB is/are still unknown.

Studying the different MB's subtypes with the most advanced technologies will significantly contribute to the overall understanding of MB pathogenesis in general and in particular in MB genetic knowledge.

3. Chromosomal aberration and its corroltion to clinicohistopatological classification

Cytogenetics and molecular genetics are two different levels of genetic studies. Both of them are in use for characterization of tumors in general and MB in particular. Nowadays, these tests in combination with other standard methods such immunohistochemistry, significantly improved the diagnosis, prognosis and consequently the treatment modalities and survival of the patients with malignant diseases. Cytogenetics, molecular cytogenetics (FISH, CGH) and molecular genetics are three methods that supplement each other, and their combination leads to the identification of the genes that are involved in tumorigenesis (tumor suppressor genes and oncogenes) and thus can give complete information for prognostic and diagnostic purposes. Although, impressive progress has been achieved in the nano molecular genetics, next generation sequencing, proteomics and transcriptomics, in the last decade, yet, we can not give up the "macro-tests": the old gold standard methods such as cytogenetics and histology. The nano tests are still new and very expensive to implicate to routine work up, they are in a developing stage and the accumulated experience with their use is still limited. There is still missing information regarding the whole range of polymorphism and how to differentiate it

from significant changes. Also the exact interactions between all the nano tests (molecular genetics, proteomics and transcriptomics) are only partially understood so far. Moreover the significance and the importance of the emerging field of epigenetics is still not clear. The field of epigenetics is dealing with changes that do not affect the molecular genetics itself but rather the changes in the expression of the gene/s by changing methylation, acetylation, phosphorylation and ubiquitination (Bernstein et al. 2007; Kouzarides 2007). Its effects include the most delicate interactions of all the living levels, starting from biochemical reaction and the interactions inside the cell compartments, cell to cell, in the organ, in the host and between the host and the environment.

Chromosome analysis of tumor cells plays an important role in the diagnosis, prognosis and follow up of many malignancies. It is a standard of care in hemato-oncology. Culturing solid tumor cells *in vitro* is essential for cytogenetic analysis. The success in getting specific chromosomal aberrations in solid tumors after *in vitro* culturing is limited because of technical problems such as: an overgrowth of cells from healthy part of the tissue mostly fibroblasts, limited growth of the abnormal tumor cells and low quality of the tumor cell metaphases and also the need for special and well trained cytogeneticists. These are probably the main reasons for the limited and what seems like a non specific chromosomal aberration in medulloblastoma. Only about 40% of the medulloblastomas demonstrate chromosomal aberrations. Presently, advanced technologies such as microarray CGH and next generation sequencing enable us to better evaluate chromosomal aberrations in medulloblastoma. It will take some time for the accumulation of new data regarding chromosomal aberrations using the new methods, mainly because reliable results should be done on a large cohort which include sufficient size group of all MB subtypes.

Copy number abnormalities of chromosome 17 isodicentric, isochromosome of chromosome 17q, loss of 17p, or gain of 17q are the most frequent chromosomal abnormality in medulloblastomas (Mitelman database 2011, Aldosari et al 2002; McCabe et al 2006) it is found in 30-50% of the studied cases (Reardon et al 1997). Isochromosome -isodicentric chromosome 17 is present in approximately one third of the tumors and in some cases is the only chromosomal aberration (Mitelman database 2011; Pfister et al 2009; Rickett and Paulus 2004). It has been observed in 25-30% of childhood MB. In fact there is a loss of 17p and a gain of 17q which is the most common isochromosome in cancer in general (Mitelman database 2011). This might point toward the fact that i(17q) is a general marker of neoplastic process rather than a specific marker of MB. Its role as an indicator for a poor outcome in MB stratification is controversial. The controversial evidence based mainly on the findings of i(17q) also in Wnt pathway subgroup which in consensus considered as a favorable outcome. The question whether to consider copy number abnormality of chromosome 17 as a marker for poor prognosis and to exclude those found to be Wnt pathway, is still in debate (Pan et al 2005; Pfister et al 2009).

Using more sophisticated and advanced techniques such as CGH to profile a panel of 27 primary MB (Reardon et al 1997) revealed frequent loss of 10q, 11, 16q, 17p, and 8p as well as recurrent gains of chromosomes 7, and 17q. These losses and gains were also confirmed by other techniques, such as G-banding, SKY and FISH (Aldosari et al 2002; Avet-Loiseau et al 1999; Bayani et al 2000; Eberhart et al 2002; Gilhuis et al 2000). Array CGH of 47 MB (Speicher and Carter 2005) showed gain of 17q, 7, and 1q and loss of 17p, 11p, 10q and 8.

There have also been reports, although less frequently of losses on chromosomes 10q, 11, 17p and 22 as well as gains on chromosomes 1q, 7 and 7q (Mitelman database 2011; Bayani et al 2000).

SKY analysis of 19 primary MB revealed structural aberrations involving chromosomes 7, 17, 3, 14, 10 and 22. (Bayani et al 2009).

Monosomy 6 is strongly associated with Wnt profile in MB (Clifford et al 2006). Chromosomal abnormalities including loss of chromosome X in females and loss of chromosome 8, have been found mainly in non SHH/Wnt subgroups.

4. Genetic alterations and its correlation with clinicohistopathological classification

Tumorigenesis is a process in which one of the pathways leading to normal cell development has lost its control and its normal function. In addition, the neovascularization, cell proliferation, differentiation, motility and apoptosis (death) are also altered.

There are two possible categories in tumor specific genetic alteration; one is inheritable or germ cells mutation and the other is acquired alteration. Also there is a difference between general tumorigenesis pathways and tumor specific pathway and general chromosomal aberration and tumor specific chromosomal aberration.

Usually, the tumor specific pathway is a part of the general tumorigenesis pathways but still it is predominant in one specific tumor subgroup, for example RB gene that causes mainly retinoblastoma, WT gene that causes mainly Willm's tumor, both are also causing predisposition to other tumors. Similarly, the acquired gene alteration occurring during the life time, such as BCR/ABL is causing chronic myeloid leukemia and EWSR1-FLI1 is causing Ewing sarcoma. Specific acquired aberration found in addition to the cancer they caused, also in other cancers, there its significance and its prognostic value is different, for instance BCR/ABL points to a favorable prognosis in CML and points to a poor prognosis in acute lymphocytic leukemia (ALL).

Among the life time acquired mutation, one should differentiate between the first event, such as alteration of a gene that causes the disease and has a diagnostic value and those mutations that are considered as the latter (the second or the third) events for example KRAS and TP53 mutation in adenomatose polyposis coli (APC) and MNP1 or FLT1 mutation in acute leukemia. Those gene alterations have mainly prognostic value and may play as a general event seen in other tumors as well.

Unfortunately, neither MB specific chromosomal changes nor MB specific gene were found to be a clear evidence for the possible pathogenesis of MB, neither in MB general nor in one of its subtypes. The understanding of MB pathogenesis is still limited, mainly because most of the MB studies are associated with genes that are already known to be involved in other tumors or syndromes, rather than studying the MB specific gene alterations as the first goal. Indeed, MB is associated with other (tumor's) syndromes such as Gorlin, Turcot and Li Fraumeni, in which several important developmental signal transduction pathways, including sonic hedgehog (SHH), Wingless (Wnt) and Notch signaling cascades. SHH, Wnt, TP53 and Notch signaling cascades are implicated in the cells migration and localization in the cerebrum, and their proliferation and differentiation. The alterations in these pathways by any component of each signaling pathway may lead to tumorigenesis (Marino 2005; Ingham and Placzek 2006). For example, Gorlin's syndrome characterized by a germ line mutation in the patched homologue 1 (PTCH1) located on 9q22 (Frandon et al 1992) that served as a negative regulator of SHH gene during normal cerebella development. Mutation in PTCH1 gene can cause predisposition to different tumor development including MB with incidence of 5-20% (Friedrich 2007).

Turcot syndrome is characterized by the mutation in the APC a tumor suppressor gene that is predisposed mainly (about 90% during life time) to colon cancer adenomatous polyposis coli (APC), (Hamilton et al 1995) but also predisposed to a lesser extent to other tumors including MB (Huang et al 2000). The APC gene regulates the Wnt signaling pathway.

Recent advances in gene expression profiling techniques have led to the generation of several molecular classification schemes in MB (Thompson et al 2006; Kool et al 2008; Northcott et al 2010 epub).

In general, based on the molecular studies done so far, three main subgroups were defined: SHH, Wnt and non SHH/Wnt pathways which include another 2 subgroups which are less distinct (Thompson et al 2006; Kool et al 2008; Onvani et al 2010; Ellison 2010; Northcott et al 2011). SHH pathway subgroup accounts for 15-25% cases of MB and have a poor prognosis, Wnt pathway subgroup accounts for 15-20% cases of MB and mostly has favorable prognosis, and non SHH/Wnt subgroups account for 60% cases of MB.

According to the Thompson's (2006) classification based on molecular and FISH examination 5 distinct subtypes identified (A to E) including subgroup B in which Wnt pathway and monosomy 6 have been found and subgroup D in which SHH pathway has been found. Kool et al (2008) further corroborated Thompson's classification using CGH and defined 5 subgroups: (A)-Wnt signaling subgroup, (B)-SHH signaling pathway, C and D expression of neural differentiation genes, D and E expression of photoreceptor genes. Subgroups C, D and E are genetically closely related and most often associated with metastatic appearance mainly subgroup E.

SHH pathway MB associated with desmoplastic phenotype (Behesti 2009) and also with large cell and anaplastic MB. They are both reported mainly in infancy and childhood. Metastatic disease at presentation characterizes some SHH pathway mostly large cells and anaplastic MB. Less than half of the SHH pathway MB have PTCH1 mutations or show copy number loss at the PTCH1 locus, 9q22, mutations in SMOH and SUFU are rare thus there must be other undiscovered SHH pathway/s (Thompson et al 2006; Kool et al 2008; Northcott et al 2011).

In contrast Wnt pathway is mostly associated with classic MB (Fattet et al 2009) it has a favorable outcome it tends to be present in childhood in the pre-teen years (6-13 years) but almost do not present in infancy. Most of the children with this tumor survived (Thompson et al 2006). Metastatic disease at presentation is rare in Wnt pathway. There is no report on desmoplastic medulloblastoma with Wnt pathway and also large cell and anaplastic MB with Wnt is rare. The APC protein is a regulator of Wnt signaling that function in a complex with other components to regulate several important developmental processes, including proliferation and specification of neural progenitor cells during early cerebellar development (Sillitoe and Joyner 2007). APC function as a tumor suppressor through CTNNB1 (β -catenin) of sporadic cases of MB, a downstream component of the Wnt signaling pathway (Hamilton et al 1995) this account for 15% of the cases. β -catenin by itself also activates transcription of several oncogenes such as MYC and CCND1 resulting in enhanced cell proliferation (Ellison et al 2005).

Only a small part of the Wnt pathway MBs are found to be carrying mutations in CTNNB1 gene-captured by identification of the nuclear activity of β -catenin in the MB. Mutations in APC and AXIN1/2 are rare, thus there are still undiscovered components also in Wnt pathway (Ellison et al 2005; Fattet et al 2009; Northcott et al 2011).

Most of non SHH/Wnt MBs have a classic pathology and present in infancy and childhood. About half of the large cell and anaplastic MBs are non SHH/Wnt. Metastatic disease at presentation also falls into the non SHH/Wnt tumor subgroup.

Other molecular markers beside the above three identified groups is the amplification of MYC family (MYC and MYCN proto-oncogenes), that account for 4-15% of MBs (Rossi et al 2008; McCabe et al 2006). MYCN is an early transcriptional target of the SHH pathway and activation by SHH promotes the expression of the cell cycle proteins cyclinD1 and CyclinD2 leading to GCP proliferation (Behesti and Marino 2009). A high expression level of MYC is reported to cause progression of MB to an anaplastic phenotype and has been linked to a poor prognosis while even though MYCN shows some association with large cells and anaplastic MB yet it is less established as a marker for an adverse outcome (Aldosari et al 2002; Pfister et al 2009).

Both SHH and Wnt subgroups rarely show copy number abnormalities of chromosome 17, amplification of MYC and MYCN or any other widespread ploidy changes. MB occurred also in 5% of the Li Fraumeni syndrome's patients carrying mutation in the suppressor gene TP53 (Northcott 2009). Other known pathways are involved in the normal cerebellar development found also to be aberrant in some of the MB. For example a disturbance of the RAS-MAP upregulation through downstream components such as MAP2K1, MAP2K2, and MAPK1/3. It is found to be correlated with metastatic behavior (Gilbertson and Clifford 2003). Also, overexpression of the EGF receptor family member ERBB2 is linked to metastatic behavior. A number of proto-oncogenes in MB such as CDK6, PDGFRA, KIT and MYCL1 have been found to be amplified by array CGH. A single copy numbers gains of MET locus on chromosome 7q in 38.5% of the cases in 13 MBs (MacBabe 2006; Tong et al 2004).

The Notch pathway was found to be implicated in MBs pathogenesis in a number of studies. Notch promote granule cell precursor (GCP) proliferation and prevents their differentiation (Behesti and Marino 2009). Increased copy number of Notch2 has been found in 15% of the studied MBs. Also, mutated Hes1 gene that unregulated the Notch pathway have been found in a small study group, although it has been associated with poor prognosis and outcome, its role and implication should be carefully considered and further examined (Fan et al 2004; Thompson et al 2006; Kool et al 2008; Northcott et al 2009).

Due to technology limitations and the availability research strategies most molecular studies of MBs, we are still missing the MB specific molecular markers. Hopefully, MB subtypes specific markers will be discovered by using the advanced technologies which enables us to study MB at different levels: DNA by next generation sequencing, gene expression by transcriptomic and proteomic, epigenomic and miRs.

An accurate classification and the stratification will implicate the medical care. An optimal classification will differentiate between MB in each age interval (infancy, childhood and adult) and between the subgroups in each age.

There is a general consensus that a better understanding of the disease biology should allow us to develop more effective and less harmful treatments of MB.

Northcott et al (2011) generated a class prediction algorithm, an 8-gene classification model which successfully predicted the survival status for 47 out of the 60 patients profiled. The markers of the cerebellar differentiation (β -NAP, NSCL1, TRKC) and component of the extracellular matrix (lysyl hydroxylase[PLOD], collagen TypeVai and elastin) predict favorable prognosis. While genes involved in cell proliferation and metabolism (MYBL2, enolas 1, LDH, HMGI[y], and cytochrome C oxidase) as well as ribosomal protein coding genes predict poor prognosis.

5. MicroRNA (miR) and its correlation with clinicohistopathological classification

The field on miRNA emerged in the last few years, concentrates on small non coding, single strand RNA molecules that are found to play a regulatory role on gene expression.

Differential microRNA (miR) expression analysis has also contributed to our knowledge on MB pathogenesis. The miRs are the short fragments of endogenous noncoding RNA that play an important role in the developmental processes by regulating gene expression. Target mRNAs are either degraded or translated by specific miRNAs. Ferretti et al (2008) performed one of the earliest expression profiles on MB. Northcott et al (2009) identified amplification of miR17/92 polycistron proto-oncogene in 6% of pediatric MBs and showed upregulation of miR17/92 expression in a large percentage of primary cases. Similarly, Pierson et al. (2008) demonstrated decreased expression of miR-124a in primary MB as well as in MB cell line. Onvani et al (2010) have reported that miR124a as a negative regulator of CDK6 which are found to be overexpressed in MB (Mendrzyk et al 2005). Additional oncogenic targets, such as miR-30b and miR-30d have also been proposed through miRNA profiling (Lu et al 2009)

Northcott et al (2009) have described subgroup classification based on transcription profile, using mRNA and miRNA examination. They identified 4 distinct subgroups including the well known Wnt and SHH and another two independent subgroup C and D. It has been found that mir17/92, cluster of oncogenic miRNAs, was highly expressed in SHH tumors in association with MYCN expression, while group C was found to be correlated with MYC.

High-resolution SNP array profiling on a group of >200 MB revealed high-level amplification of miR-17/92 on 13q31 (Northcott et al 2009).

It has been found that miR-17/92 and related paralogs (miR-106a/363 and miR-106b/25) were identified as the most highly upregulated miRNAs in MB when compared with normal cerebellum in this analysis. The combination of miR 17/92 amplification and consistent overexpression suggested miR-17/92 as a key player in MB pathogenesis. There is evidence that miR17/92 might cooperate with SHH in MB, as it showed preferential upregulation in SHH subtype.

Recently the role of miRNA in MB has been studied on 14 primary cases using profile of 248 miRNAs showing a general biased downregulation in MB cells as compared to the control cells . A subset of 86 miRNAs which were previously reported to be expressed in neuronal tissues and/or cancer studied in cohort of 34 among them two neuronal candidates miR-9 and miR-125a were chosen for functional analysis. Induction of their expression resulted in the decrease in tumorigenic features: promoted apoptosis, inhibited cell proliferation and impaired anchorage-independent growth. Moreover loss of miR9 and miR125a correlated well with upregulation of truncated trkC which was identified as a target for posttranscriptional repression (Northcott 2009).

In order to discriminate between miRs deregulation in SHH-driven MB from non SHH-cases, 31 MBs studied using a set of 250 miRs. Two groups were defined GIL1-high and GIL1-low. A set of 34 miRs was found with a significant differential expression between the two classes. For functional analysis of the GIL1-high class three candidates (miR-125b, miR-324-5p, and miR-326) exhibiting reduced expression were chosen based on their predicted capacity to target the SHH family members, Smo and Gil1. They were proven to repress Smo mRNA level in the MB cell line (Ferretti et al 2008).

SNPs array profiling of more than 200 MB revealed copy number aberration of multiple unreported regions including high level amplification of miR17/92

Another miRNAome study of 90 MB revealed that four distinct molecular subgroups can be described. These subtypes include the well characterized Wnt and SHH subgroups and another 2 subgroup designated C and D (Northcott et al 2009).

The miR-17/92 was the most highly expressed in SHH-driven MB but also in tumors exhibiting high MYCN (SHH) and MYC (group C,Wnt) expression. MiR-17/92 transcriptional upregulation by N-Myc and Myc and confirming miR-17/92 aberrancy in a large percentage of the MBs (□ about 60%). Based on experiments on mice it was suggested by Northcott et al (2009) that miR 17/92 cooperate with SHH signaling to promote and/or enhance CGNP proliferation.

The miRs studies are still in their initial stage and we are still far from understanding their exact role and how they affect processes in MB pathogenesis.

6. Epigenomic and its correlation with clinicohistopathological classification

Until recently the thought was that genetic alteration is the main cause of each tumor development and progress. Researchers invest their efforts in finding a tumor specific pattern of genetic alteration. Over the past few years another aspect has arisen which is the deregulation of epigenetics to malignancy (Jones and Baylin 2002; Bernstein et al 2007; Kouzarides 2007; Kongham et al 2008; Esteller 2008).

Epigenetics involves nongenetic DNA modifications that result in changes in gene expression. These changes include gene promoter methylation on cytosine residues, most frequently of the CpG islands of the promoter, as well as their histone code. These changes are found to be involved in pathogenesis of tumors including MB, through hypermethylation of promoter of tumor suppressor genes and consequently silencing them (Kongham et al 2008).

Fruhwald et al (2001) showed methylation of up to 1% of all CpG island in 17 primary medulloblastomas, it was linked to poor prognosis.

Epigenetics is defined as "mitotically heritable changes in gene expression that are not accompanied by modifications in primary DNA sequence". It is highly correlated with the MB class were ZIC and NSCL1, encoding transcription factors that are specific for cerebellar granule cells point to the MB arising from cerebellar granule cells.

Anderton et al (2008) have identified tumor-specific methylation of COL1A2 in 77% of the studied primary MBs (46 out of 60) and showed an age-dependent methylation pattern for this gene in desmoplastic tumors, which presented COL1A2 as a potential MB subtype biomarker. Kongkham et al. (2008) identified serine protease inhibitor kunitz-type2 (SPINT2/HAI-2), an HGF/cMET signaling inhibitor, as a novel tumor suppressor gene that is frequently silenced by promoter hypermethylation in MBs (Kongkham et al 2008). Also Waha et al (2007) found promoter hypermethylation-induced reduction of SCG5 expression in primary 16 out of 23 primary cases compared to normal cerebellar controls (Waha 2007), points to its possible role in pathogenesis of MB. Furthermore, promoter hypermethylation-mediated silencing of CASP8, HIC1 and RASSF1A tumor suppressor genes has also been discovered in more than 30% of MBs by various groups (Lindsey et al 2004).

Pfister et al (2007) showed a striking association between samples classified as either "low methylators" or "high methylators" and patient outcome, where the "high methylators" group exhibited reduced overall survival.

Also the GLI C2H2-type zinc-finger protein family member ZIC2 was identified as hypermethylated and thus it has been silenced (Pfister et al 2007).

It has been found that EHMT1 function as part of a transcriptional repressor complex that mediates gene silencing by promoting dimethylation of H3K9 (Tachibana et al 2005), a

repressive epigenetic modification (Bernstein et al 2007; Kouzarides 2007) in the promoter regions of target genes. An obvious correlation between loss of EHMT1 leads to H3K9 hypomethylation.

Another study using microarray-based differential methylation hybridization (Waha et al 2007) identified hypermethylation of the SCG5 (secretory granule, neuroendocrine protein 1[7B2 protein] gene) in 16 out of 23 (70%) primary MB. Expression of SCG5 found to be downregulated in the MB in comparison with normal cerebral controls. Another gene that found to be down regulated is SPINT2 that was found in 41 out of 56 primary MB. Stable expression of SPINT2 resulted in attenuation of the malignant phenotype: inhibiting cell proliferation, anchorage-independent growth in soft agar and cell motility, of cell lines (Kongham et al 2008). This study suggested that SPINT2 is a suppressor gene. Treatment of MB cell lines suggesting that SCG5 is a suppressor gene (Fruhwald et al 2001) or increased gene with demethylation agent (5-aza-2'-deoxycytidine) reduced colony formation expression (Anderton et al 2008; Kongham et al 2008).

The MB epigenomic studies demonstrate that not only the genetic alteration can cause loss of control in a cell and transformation to a malignant cell but also other mechanism can cause malignancy, implicated epigenetic gene silencing as important mechanism of the tumor suppressor gene inactivation in MB.

7. Summary

The reported genetic alterations of MB, either chromosomal or molecular, are so far not specific. The main distinct subgroups are the Wnt and the SHH both account for 30-40% of MB. These pathways are common in many other tumors, suggesting they are not exclusive to MB. There are some other pathways involved in MB pathogenesis that might be more specific to MB. Similarly, the chromosomal aberration i(17q) which is found in about 40% of MBs, as well as in many other tumors including in chronic myelocytic leukemia (CML) can possibly be a secondary chromosomal aberration. Some of the features found in different subtypes but not in an equal distribution, for instance i(17q) were found in 34%, 36%, 12% in classic, large cell and desmoplastic MB respectively and are associated with poor prognosis (Gillbertson & Ellison 2008). Another example NOTCH and PDGF, they both have been found in A and B Kool's subtypes and also subtypes C, D and E share increased expression of neural differentiation genes. Another example is the Wnt- β catenin, found in all subtypes defined in Northcott's classification. This overlapping between the different subtypes, points toward the fact that some of the alterations are not, necessarily, a specific prognostic marker. Studies have been done on a mice model and on cell lines in order to learn more about pathogenesis of MB are important; however the data learned from these experimental systems should be first corroborated with the data learned from human primary MBs, before going to conclusions on the pathogenesis of human MB. This is the reason for focusing in this chapter mainly on the studies done on primary human MBs studies.

Due to the variable results, the exact and specific chromosomal changes in MB which is a crucial event in pathogenesis, is still unknown.

Gilberston and Ellison (2008) wrote in their review: "Genomic-seeing the wood and the trees". The literature is full of studies (over 200 papers) on genetic alteration in MB trying to understand the MB pathogenesis, some of them corroborate with each other. Most of the studies have been done from different research points of view and emphasis, using different technologies. Presently, there are at least 5 different suggestions for classification and

stratification of MB: Thompson's (2006), WHO (2007), Kool's (2008), Pefister's (2008) and Northcott's (2011). It is very difficult to combine them to one clear cut classification (Table 1).

*THOMPSON'S (2006) CLASSIFICATION (46)	WHO CLASSIFICATION	PROGNOSIS	MOLECULAR ALTERATION	CHROMOSOMAL ABERRATION	AGE OF ONSET
A				Gain:17q Loss: 17p	
B	Classic MB	Favorable	Wnt- β catenin \uparrow CTNNB1- predominant,A PC, AXIN1mutation	Monosomy 6	≥ 3 years
C			SHH PTCH,SUFU mutation	Gain:17q Loss: 17p	
D	Desmoplastic		SHH, PTCH,SUFU mutation		≤ 3 years
E				Gain:17q Loss: 17p	
*KOOL'S(2008) CLASSIFICATION (52)	WHO CLASSIFICATION	PROGNOSIS	MOLECULAR ALTERATION	CHROMOSOMAL ABERRATION	AGE OF ONSET
A	classic		Wnt β -catenin mutation NOTCH, PDGF	Monosomy 6	Older children
B	desmoplastic		SHH PTCH1 mutation NOTCH, PDGF	9q loss	Young children years and adults
C	classic	metastases	Neural differentiation genes	17 alteration, loss of X chromosome (females)	children
D	classic	metastases	Neural differentiation genes Photoreceptor genes	17 alteration, loss of X chromosome (females)	children
E	classic	metastases	Photoreceptor genes	loss of X chromosome (females)	Young children
*PFISTER'S(2009) CLASSIFICATION (80/260)	WHO CLASSIFICATION	PROGNOSIS	Frequency		OS
MYC/MYCN amplification+10ch.	Large cell/anaplastic	Poor prognosis methastases	6%/4%/10%		13%

aber./ MYC/MYCN amplification +6q-gain+17qgain +10ch.aber.					
6q-gain+10ch.aber.		Poor prognosis	8%		16%
17q-gain/17p-loss/i(17q) +7.5ch.aber.		Poor prognosis	48%/39%/30-48%		56%
6q ,17qbalanced +4ch.aber.					90%
6qdeletion +2ch.aber.		Favorable prognosis	12%		100%
*NORTHCOTT'S (2010) CLASSIFICATION (103)**	WHO CLASSIFICATION	PROGNOSIS	MOLECULAR ALTERATION	CHROMOSOMAL ABERRATION	AGE OF ONSET
Wnt			MYC↑ Wnt-β catenin	Monosomy 6	Distributed age median 9-10 years, 3:1 F/M
SHH	Desmoplastic-predominant Anaplastic, large cell		MYCN↑ Wnt-β catenin	Del9q Isochromosome 9p Gain: 3q, 20q, 21q,2 Loss:10q,14	Infants ≤3 years-most common, adults≥16 years
C	Desmoplastic Anaplastic, large cell(23%)	Metastases(46.5%) Worst prognosis	OTX2↑,FOXG1 B↑, MYC(8q24)↑, Wnt-β catenin Neural development*	Isochromosome (17q) Gain:1q,17q,8 Loss:10q, 5qdistal, 16q,11p ,8p	Childhood peak 3-10 years
D	Desmoplastic Anaplastic, large cell (8%)	Metastases(29.7%)	OTX2↑,FOXG1 B↑ Wnt-β catenin, Neural development*	Isochromosome (17q) Gain: 17q,8 Loss:11p ,X(females), 8p,8q	Distributed age median 9-10 years

WHO (2007): Classic, desmoplastic, MB with extensive nodularity, anaplastic, Large cell

*For more details see: Louis 2007, Thompson *et al* 2006, Kool *et al* 2008, Pifster *et al* 2009, Northcott *et al* 2010 J. Clin.Oncol., Parenthesis-Year of publication.

** See table 2 in Ellison 2010, Table 2 in Huse and Holland 2010, Table 1-Northcott 2010 J. Clin.Oncol. Parenthesis -Number of samples

Table 1. Medulloblastoma classification according to: WHO- Louis *et al* 2007, Thompson *et al* 2006, Kool *et al* 2008, Pifster *et al* 2009, Northcott *et al* 2011.

There is also early evidences that epigenetics and miRs might play a role in MB pathogenesis and can be used as a prognostic tool. However , the data regarded to epigenetics and miRs in MB is still limited and uncompleted, as part of the studies done on MB cell line which might point to a candidate involved genes with no assurance for there role in the MB tumor. There are few studies on primary MBs, thus any conclusion from this data is still immature. This

emphasized the need for further studying the MB pathogenesis for either specific germ line mutation or other specific level of alteration (transcriptome, proteome, epigenome levels). These should be done as a multi-center study, on a large size of cohort including sufficient number of samples of each MB subtype including adult and childhood MBs. The study should be performed uniformly using different levels of examinations: histologic, cytogenetic, molecular, transcriptome, proteome, epigenetics, and miRs. Hopefully such a study will provide us with more personalized medical care with less adverse side effects.

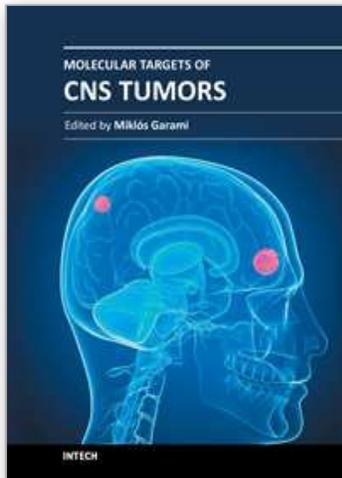
8. References

- Aldosari, N., Bigner, S.H., Burger, P.C., Becker L, Kepner JL, Friedman HS, and McLendon RE. 2002. MYCC and MYCN oncogene amplification in medulloblastoma. A fluorescence in situ hybridization study on paraffin sections from the Children's Oncology Group. *Arch. Pathol. Lab. Med.* 126:549-544.
- Anderton, J.A., Lindsey, J.C., Lusher, M.E., Gilbertson, R.J., Bailey. S., Ellison, D.W., and Clifford, S.C. 2008. Global analysis of the medulloblastoma epigenome identifies disease-subgroup-specific inactivation of COLIA2. *Neuro Oncol* 10:981-994
- Avet-Loiseau, H., Venuat, A.M., Terrier-Lacombe, M.J., Lellouch-Tubiana, A., Zerach, M., and Vassal, G. 1999. Comparative genomic hybridization detects many recurrent imbalances in central nervous system primitive neuroectodermal tumors in children. *Brit J Cancer* 79:1843-1847
- Bar, E.E., and Stearns, D. 2008. New developments in medulloblastoma treatment: the potential of a cyclosporine-lovastatin combination. *Expert Opin Invest Drugs* 17:185-195
- Bayani, J., Zielenska, M., Marrano, P., Kwan Ng, Y., Taylor, M.D., Jay, V., Rutka, J.T., and Squire, J.A.. 2000. Molecular primitive neuroectodermal tumors by using conventional banding, comparative genomic hybridization, and spectral karyotyping. *J Neurosurg* 2000;93:437-438
- Behesti, H., and Marino, S. 2009. Cerebellar granule cells: insights into proliferation, differentiation and role in medulloblastoma pathogenesis. *Int J Biochem Cell Biol* 41:435-445
- Bernstein, B.E., Meisner, A., and Lander, E.S.. 2007. The mammalian epigenome. *Cell* 128: 669-681
- Brandes, A.A., Franceschi, E., Tosoni, A., Reni, M., Gatta, G., Vencht, C., and Kortmann, R.D. 2009. Adult neuroectodermal tumors of posterior fossa (medulloblastoma) and of supratentorial sites (stPNET). *Crit Rev Oncol Hematol* 71:165-179
- Clifford, S.C., Lusher, M.E., Lindsey, J.C., Langdon, J.A., Gilbertson, R.J., Straughton, D., and Ellison, D.W. 2006. Wnt/Wingless pathway activation and chromosome 6 loss characterize a distinct molecular sub-group of medulloblastomas associated with a favorable prognosis. *Cell Cycle* 5:2666-2670
- Crawford, J.R., MacDonald, T.J., and Packer, R.J. 2007. Medulloblastoma in childhood; new biological advances. *Lancet Neurol* 6:1073-1085
- Eberhart, C.G., Kratz, J.E., Schuster, A., Goldthwaite, P., Cohen, K.J., Perlman, E.J., and Burger, P.C. 2002. Comparative genomic hybridization detects an increased number of chromosomal alterations in large cell/anaplastic medulloblastomas. *Brain Pathol* 12:36-44
- Ellison, D. 2002. Classifying the medulloblastoma: insight from morphology and molecular genetics. *Neuropathol Appl Neurobiol* 28:257-282

- Ellison, D.W., Onilude, O.E., Lindsey, J.C., Lusher, M.E., Weston, C.L., Taylor, R.E., Pearson, A.D., and Clifford, S.C. 2005. Beta-Catenin status predicts a favorable outcome in childhood medulloblastoma. *J Clin Oncol* 23:7951-7957
- Ellison, D.W. 2010. Childhood medulloblastoma: novel approaches to the classification of a heterogeneous disease. *Acta Neuropathol* 120:305-316
- Esteller, M. 2008. Epigenetics in cancer. *N Eng J Med* 358: 1148-1159
- Fattet, S., Haberler, C., Legoix, P., Varlet, P., Lellouch-Tubiana, A., Lair, S., Manie, E., Raquin, M.A., Bours, D., Carpentier, S., Barillot, E., Grill, J., Doz, F., Puget, S., Janoueix-Lerosey, I., and Delattre, O. 2009. Beta-catenin status in paediatric medulloblastoma: correlation of immunohistochemical expression with mutational status, genetic profiles and clinical characteristics. *J Pathol* 218:86-94
- Ferretti, E., De Smaele, E., Miele, E., Laneve, P., Po, A., Pelloni, M., Paganelli, A., Di Marcotulli, L., Caffarelli, E., Screpanti, L., Bozzoni, L., and Gulino, A. 2008. Concerted microRNA control of Hedgehog signalling in cerebellar neuronal progenitor and tumor cells. *EMBO J* 27:2616-2627
- Farndon, P.A., Del Mastro, R.G., Evans, D.G., Kilpatrick, M.W. 1992. Location of gene for Gorlin syndrome. *Lancet* 339:581-582
- Friedrich, R.E. 2007. Diagnosis and treatment of patients with nevoid basal cell carcinoma syndrome (Gorlin-Goltz syndrome-GGS). *Anticancer Res* 27:1783-1787
- Fruhwald, M.C., O'Dorisio, M.S., Dai, Z., Tanner, S.M., Balster, D.A., Gao, X., Wright, F.A., and Plass, C. 2001. Aberrant promoter methylation of previously unidentified target genes is a common abnormality in medulloblastomas-implications for tumor biology and potential clinical utility. *Oncogene* 16:5033-5042
- Gilbertson, R.J. 2004. Medulloblastoma: signalling a change in treatment. *Lancet Oncol* 5; 209-18
- Gilbertson, R.J., and Ellison, D.W. 2008. The origins of medulloblastoma subtypes. *Annu Rev Pathol Mech Dis* 3:341-365
- Gilbertson, R.J., and Clifford, S.C. 2003. PDGFRB is overexpressed in metastatic medulloblastoma. *Nat Genet* 35:197-198
- Grill, J., Saint-Rose, C., Jouvret, A., Gentet, J.C., Lejars, O., Frappaz, D., Doz, F., Rialland, X., Pichon, F., Bertozzi, A.I., Chastagner, P., Couanet, D., Habrand, J.L., Raquin, M.A., Le Deley, M.C., and Kalifa, C. 2005. Treatment of medulloblastoma with postoperative chemotherapy alone: an SFOP prospective trial in young children. *Lancet Oncol* 6:573-580
- Hamilton, S.R., Liu, B., Parsons, R.E., Papadopoulos, N., Jen, J., Powell, S.M., Krush, A.J., Berk, T., Cohen, Z., Tetu, B. 1995. The molecular basis of Turcot's syndrome. *N Eng J Med* 332:839-847
- Huang, H., Mahler-Araujo, B.M., Sankila, A., Chimelli L, Yonekawa Y, Kleihues P, Ohgaki H. 2000. APC mutations in sporadic medulloblastomas. *Am J Pathol* 156:433-437
- Ingham, P.W., and Placzek, M. 2006. Orchestrating oncogenesis : variations on a theme by sonic hedgehog. *Nat. Rev Genet* 7:841-850
- Jones, P.A. , and Baylin, S.B. 2007. The epigenomics of cancer. *Cell* 128:683-692
- Kongham, P.N., Northcott, P.A., Ra, Y.S., Nakahara, Y., Mainprize, T.G., Croul, S.E., Smith, C.A., Taylor, M.D., and Rutka, J.T. 2008. An epigenetic genome-wide screen identifies SPINT2 as a novel tumor suppressor gene in pediatric medulloblastoma. *Cancer Res* 68:9945-9953
- Kool, M., Koster, J., Bunt, J., Hasselt, N.E., Lakeman, A., van Sluis, P., Troost, D., Meeteren, N.S., Caron, H.N., Cloos, J., Mrcic, A., Ylstra, B., Grajkowska, W., Hatmann, W., Pietsch, T., Ellison, D., Clifford, S.C., and Versteeg, R. 2008. Integrated genomics

- identifies five medulloblastoma subtypes with distinct genetic profiles, pathway signatures and clinicopathological features. *PLoS One* 3:e3088
- Korshunov, A., Remke, M., Werft, W., Benner, A., Ryzhova, M., Witt, H., Sturm, D., Wittman, A., Scholter, A., Felsberg, J., Reifenberg, G., Rutkowski, S., Scheurlen, W., Kulozik, A.E., von Deimling, A., Lichter, P., and Pfister, M. Adult and pediatric medulloblastomas are genetically distinct and require different algorithms for molecular risk stratification. *J Clin Oncol* 2010
- Kouzarides, T. 2007. Chromatin modifications and their function. *Cell* 128:693-705
- Lindsey, J.C., Lusher, M.E., Anderton, J.A., Bailey, S., Gilbertson, R.J., Pearson, A.D., and Ellison, D.W. 2004. Identification of tumour-specific epigenetic events in medulloblastoma development by hypermethylation profiling. *25:661-668*
- Louis, D.N. 2007 WHO classification of tumors of the central nervous system. International Agency for Research on Cancer, Lyon.
- Lu, Y., Ryan, S.L., Elliott, D., Bingell, G.R., Futreal, P.A., Ellison, D.W., Bailey, S., Clifford, S.C. 2009. Amplification and overexpression of Hsa-miR-30b, Hsa-miR-30d and KHDRBS3 at 8q24.22-q24.23 in medulloblastoma. *PLoS One* 4, e6159
- Marino, S. 2005. Medulloblastoma : developmental mechanisms out of control. *Trends Mol. Med* 11:17-22
- McCabe, M.G., Ichimura, K., Liu, L., Plant, K., Backlund, L.M., Pearson, D.M., and Collins, V.P. 2006. High-resolution array-based comparative genomic hybridization of medulloblastomas and supratentorial primitive neuroectodermal tumors. *J Neuropathol Exp Neurol* 65:549-561
- Mendrzyk, F., Radlwimmer, B., Joos, S., Kokocinski, F., Benner, A., Stange, D.E., Neben, K., Fiegler, H., Carter, N.P., Reifenberger, G., Korshunov, A., and Lichter, P. 2005. Genomic and protein expression profiling identifies CDK6 as novel independent prognostic marker in medulloblastoma. *J Clin Oncol* 23:8853-8862
- Mitelman, F., Johansson, B., Mertens, F. (eds) (2011) Mitelman database of chromosome aberrations in cancer. Retrieved from: <http://cgap.nci.nih.gov/Chromosomes/Mitelman> Database last updated Feb 11, 2011
- Northcott, P.A., Nakahara, Y., Wu, X., Feuk, L., Ellison, D.W., Croul, S., Mack, S., Kongkham, P.N., Peacock, J., Dubuc, A., Ra, Y.S., Zilberberg, K., McLeod, J., Scherer, S.W., Sunil Rao, J., Eberhart, C.G., Grajkowska, W., Gillespie, Y., Lach, B., Grundy, R., Pollack, I.F., Hamilton, R.L., Van Meter, T., Carlotti, C.G., Boop, F., Binger, D., Gilbertson, R.J., Rutka, J.T., and Taylor, M.D. 2009. Multiple recurrent genetic events converge on control of histone lysine methylation in medulloblastoma. *Nat Genet* 41:465-472
- Northcott, P.A., Rutka, J.T., Taylor, M.D. 2010. Genomics of medulloblastoma: from Giesma-banding to next-generation sequencing in 20 years. *Neurosurg Focus* 28: E6
- Northcott, P.A., Korshunov, A., Witt, H., Hielschner, T., Eberhart, C.G., Mack, S., Bouffet, E., Clifford, S.C., Hawkins, C.E., French, P., Rutka, J.T., Pfister, S., and Taylor, M.D. 2011. Medulloblastoma comprises four distinct molecular variants. *J Clin Oncol* 29:1408-14
- Onvani, S., Etame, A.B., Smith, C.A., and Rutka, J.T. 2010. Genetics of medulloblastoma: clues for novel therapies. *Expert Rev Neurother* 10:811-823
- Packer, R.J., and Vezina, G. 2008. Management of and prognosis with medulloblastoma: therapy at the crossroad. *Arch Neurol* 65:1419-1424
- Padovani, L., Sunyach, M.P., Perol, D., Mercier, C., Alapetite, C., Haie-Meder, C., Hoffstetter, S., Muracciole, X., Kerr, C., Wagner, J.P., Lagrange, J.L., Maire, J.P., Cowen, D., Frappaz, D., and Carrie, C. 2007. Common strategy for adult and

- pediatric medulloblastoma: A multicenter series of 253 adults. *Int J Radiat Oncol Biol Phys* 68: 433-440
- Pan, E., Pellarin, M., Holmes, E., Smirnov, I., Misra, A., Eberhart, C.G., Burger, P.C., Biegel, J.A., and Feuerstein, B.G. 2005. Isochromosome 17q is a negative prognostic factor in poor-risk childhood medulloblastoma patients. *Clin Cancer Res* 11:4733-4740
- Pfister, S., Remke, M., Benner, A., Menderzyk, F., Toedt, G., Felsberg, J., Wittmann, A., Devens, F., Gerber, N.U., Joos, S., Kulozik, A., Reifenberger, G., Rutkowsky, S., Wiestler, O.D., Radlwimmer, B., Scheurlen, W., Lichter, P., and Korshunov, A. 2009 Outcome prediction in pediatric medulloblastoma based on DNA copy-number aberrations of chromosomes 6q and 17q and the MYC and MYCN loci. *J Clin Oncol* 27:1627-1636
- Pierson, J., Hostager, B., Fan, R., and Vubhakar, R. 2008. Regulation of cyclin dependent kinase 6 by microRNA 124 in medulloblastoma. *J Neurooncol* 90:1-7
- Pizer, B.L., and Clifford, S.C. 2009. The potential impact of tumor biology on improved clinical practice for medulloblastoma: progress towards biologically driven clinical trials. *Brit J Neurosurg* 23:364-375
- Polkinghorn, W.R., and Tarbell, N.J. 2007. Medulloblastoma: Tumorigenesis, current clinical paradigm, and effort to improve risk stratification. *Nat Clin Pract Oncol* 4:295-304
- Reardon, D.A., Michalkiewicz, E., Boyett, J.M., Sublett, J.E., Entrek, R.E., Ragsdale, S.T., Valentine, M.B., Behm, F.G., Li, H., Heideman, R.L., Kun, L.E., Shapiro, D.N., and Look, A.T. 1997. Extensive genomic abnormalities in childhood medulloblastoma by comparative genomic hybridization. *Cancer Res* 57: 4042-4047
- Ribi, K., Rely, C., Landolt, M.A., Allber, F.D., Boltshauser, E., and Grotzer, M.A. 2005. Outcome of medulloblastoma in children: long-term complications and quality of life. *Neuropediatrics* 36:357-365
- Rickett, C.H., and Paulus, W. 2004. Comparative genomic hybridization in central and peripheral nervous system tumors of childhood and adolescence. *J Neuropathol Exp Neurol* 63:399-417
- Rossi, A., Caracciolo, V., Russo, G., Reiss, K., and Giordano, A. 2008. Medulloblastoma: from molecular pathology to therapy. *Clin Cancer Res* 14:971-976
- Rutkowski, S., Gerber, N.U., von Hoff, K., Gnekow, A., Bode, U., Graf, N., Emser, A., Ottensmeiser, H., Deinlein, F., Schlegel, P.G., Kortmann, R.D., Pietsch, T., and Kuehl, J. 2009 Treatment of early childhood medulloblastoma by postoperative chemotherapy and deferred radiotherapy. *Neuro Oncol* 11:201-210
- Sillitoe, R.V., and Joyner, A.L. 2007. Morphology, molecular codes and circuitry produce the three-dimensional complexity of the cerebellum. *Annu Rev Cell Dev Biol* 23:549-577
- Speicher, M.R., and Carter, N.P. 2005. The new cytogenetics: blurring the boundaries with molecular biology. *Nat Rev Genet* 2005. 6:782-792
- Thompson, M.C., Fuller, C., Hogg, T.L., Dalton, J., Finkelstein, D., Lau, C.C., Chintagumpala, M., Adesina, A., Ashley, D.M., Kellies, S.J., Taylor, M.D., Curran, T., Gajjar, A., and Gilbertson, R.J. 2006. Genomics identifies medulloblastoma subgroups that are enriched for specific genetic alterations. *J Clin Oncol* 24:1924-1931
- Tong, C.Y., Hui, A.B., Yin, X.L., Pang, J.C., Zhu, X.L., Poon, W.S., and Ng, H.K. 2004. Detection of oncogene amplifications in medulloblastomas by comparative genomic hybridization and array-based comparative genomic hybridization. *J Neurosurg* 100 (2 Supp. Pediatr):187-193
- Waha, A., Koch, A., Hartman, W., Milde, U., Felsberg, J., Hubner, A., Mikeska, T., Goodyer, C.G., Sorensen, N., Lindberg, I., Wiestler, O.D., Pietsch, T., and Waha, A. 2007. SGNE1/7B2 is epigenetically altered and transcriptionally downregulated in human medulloblastomas. *Oncogene* 26:5662-5668



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University Campus STeP Ri
Slavka Krautzeka 83/A
51000 Rijeka, Croatia
Phone: +385 (51) 770 447
Fax: +385 (51) 686 166
www.intechopen.com

InTech China

Unit 405, Office Block, Hotel Equatorial Shanghai
No.65, Yan An Road (West), Shanghai, 200040, China
中国上海市延安西路65号上海国际贵都大饭店办公楼405单元
Phone: +86-21-62489820
Fax: +86-21-62489821

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